DEPARTMENT OF PATHOLOGY & LABORATORY MEDICINE

Residency Procedure Manual

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This manual represents the institutional guidelines, policies and procedures governing the selection, appointment, evaluation and promotion of residents at the University of Kansas School of Medicine. While every effort has been made to ensure the accuracy and comprehensiveness of the information presented, the content of this manual is subject to change. Unless otherwise noted, all policies included in and revisions of this document become effective upon their publication on www.kumc.edu. Individuals seeking the most recent additions or revisions should contact the Office of the Associate Dean for Graduate Medical Education.

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MISSION, GOALS AND PHILOSOPHY

The mission of the Department of Pathology & Laboratory Medicine at the University of Kansas School of Medicine is to provide excellent teaching, research, patient care, and community service to meet the health needs of Kansas and the community at large. Our aim is to provide a supportive work environment in which each individual can excel and pursue avenues that lead to national and international recognition. We will accomplish this by developing mechanisms that make optimal use of our human and financial resources.

Within this context the overall goals of our residency training program are to develop a pathologist with the following characteristics:

- A pathologist capable of communicating as a medical consultant to other clinicians and to patients, as well as being capable of optimally directing the management of anatomic and clinical laboratory enterprises. The pathologist understands the science and technology of laboratory medicine and assures the quality, clinical appropriateness, and usefulness of the data produced by that laboratory. The pathologist is a clinician first and foremost.
- A pathologist who has the skills to recognize, interpret, and communicate pathologic processes in the clinical practice of anatomic pathology.
- A pathologist who understands and consults on methods of diagnostic test development, test utilization in the context of both generally applicable as well as patient-specific clinical settings, and assay interpretation in the acute and chronic clinical management of patients. These activities include the pathologist's role in the development and implementation of integrated medical informatics that optimize patient care.
- A pathologist who has the skills to consult in these areas at the broader systems level, and in the various extant healthcare delivery models.
- A pathologist who understands the role of research, in its broadest definition, in clinical decision-making, test development, knowledge generation, and continuing education.

To accomplish these goals, our program offers a flexible but educationally intensive training program in Anatomic Pathology (AP) and Clinical Pathology (CP, Laboratory Medicine) in order to prepare each of our residents for certification by the American Board of Pathology. A core program provides training that will lead to basic competence in general pathology. Elective opportunities are offered to permit the development of specialty excellence in particular subspecialty fields within Pathology such as Surgical Pathology and Hematopathology. Research activity is encouraged for our residents.

Values that we feel are important are that the program will be balanced in AP and CP, and we will practice and teach state-of-the-art diagnostic pathology together with a strong foundation in pathogenesis and the molecular basis of disease. We encourage a strong and collegial relationship between faculty and residents and other members of the department. We believe that when our residents finish this program, they will have acquired outstanding skills and knowledge to obtain the fellowship, faculty or hospital position of their choice.
OVERALL EDUCATIONAL GOALS

Competencies that are common to all rotations are outlined below. Competencies that are specific to individual rotations are included with each sub-discipline. Residents will be given graduated responsibilities and will be evaluated at two general skill levels. Specific goals and learning objectives (Skill Levels) are described under each sub-discipline.

### Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Didactic lecture</th>
<th>DL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Faculty sign-out</td>
<td>FSO</td>
</tr>
<tr>
<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
</tr>
<tr>
<td>Role modeling</td>
<td>RM</td>
</tr>
<tr>
<td>Lab inspections</td>
<td>LI</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
<td>IC</td>
</tr>
<tr>
<td>Online tools</td>
<td>OT</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
<td>USC</td>
</tr>
<tr>
<td>Project</td>
<td>P</td>
</tr>
</tbody>
</table>

### Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Report review</th>
<th>RR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Direct observation</td>
<td>DO</td>
</tr>
<tr>
<td>Checklist</td>
<td>CL</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
</tr>
<tr>
<td>Standardized exam</td>
<td>SE</td>
</tr>
<tr>
<td>Practical slide exam</td>
<td>PSE</td>
</tr>
<tr>
<td>In-house written exam</td>
<td>IWE</td>
</tr>
<tr>
<td>360 multisource rating</td>
<td>360</td>
</tr>
<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
</tr>
</tbody>
</table>

### CORE COMPETENCY: PATIENT CARE

**Goal:**
Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of pathology services.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
</tbody>
</table>
Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.

Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of anatomic and clinical pathology.

Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity. Where clinically appropriate, consult on the use of laboratory-based therapeutics such as blood transfusion and other forms of cellular therapy.

Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.

Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.

Note: The American Board of Pathology requires the following for board eligibility: a minimum of 50 autopsies, 2,000 surgical specimens, 1,500 cytology specimens and 200 intraoperative consultations.

### CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to pathology.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>DL, FSO, JC, DSP, RM, IC, OT, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in laboratory medicine.</td>
<td>DL, FSO, JC, DSP, IC, OT, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, FSO, JC, RM, IC, OT</td>
<td>RR, DO, GR/FE, SE, IWE, PCL</td>
</tr>
<tr>
<td>Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.</td>
<td>DL, JC, RM, LI, OT, P</td>
<td>DO, CL, GR/FE, SE, IWE, 360</td>
</tr>
</tbody>
</table>
Recognize the unique aspects of laboratory medicine practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice.

Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), College of American Pathologists (CAP), and similar organizations.

Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations.

Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice.

Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytics in the local setting.

**CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**
*Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM, OT, P</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>FSO, JC, RM, OT, P</td>
<td>DO, CL, GR/FE</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM, USC</td>
<td>RR, DO, GR/FE, SE, PSE, IWE, 360</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>FSO, RM, LI</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT, P</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>
**CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS**

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to write articulate, legible, and comprehensive yet concise reports and consultation notes. Provide a clear and informative report, including a precise diagnosis whenever possible, a differential diagnosis when appropriate, and recommended follow-up or additional studies as appropriate.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, 360</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC, USC</td>
<td>DO, GR/FE, PSE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the anatomic pathology and clinical pathology service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE, PSE, 360</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>FSO, RM, IC, USC</td>
<td>RR, DO, GR/FE, PSE, 360</td>
</tr>
<tr>
<td>Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to transfusion medicine.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
<td>FSO, JC, RM, IC, USC</td>
<td>DO, GR/FE, PSE, 360</td>
</tr>
</tbody>
</table>

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## Core Competency: Professionalism

**Goal:**

*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>

## Core Competency: System-Based Practice

**Goal:**

*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the clinical laboratory in the healthcare system.</td>
<td>DL, FSO, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
<td>FSO, RM</td>
<td>RR, DO, GR/FE, SE, IWE, 360, PF</td>
</tr>
<tr>
<td>Task</td>
<td>Responsible</td>
<td>Additional Responsible</td>
</tr>
<tr>
<td>---------------------------------------------------------------------</td>
<td>-------------</td>
<td>------------------------</td>
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<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the laboratory regulatory environment,</td>
<td>DL, FSO,</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>including licensing authorities; federal, state, and local public</td>
<td>LI, OT</td>
<td></td>
</tr>
<tr>
<td>health rules and regulations; regulatory agencies such as the</td>
<td></td>
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<tr>
<td>Centers for Medicare and Medicaid Services and the US Food and</td>
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<tr>
<td>Drug Administration; and accrediting agencies such as The Joint</td>
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<tr>
<td>Commission (TJC), CAP, and the Accreditation Council for</td>
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<tr>
<td>Graduate Medical Education (ACGME).</td>
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<tr>
<td>Understand and implement policies to continually improve patient</td>
<td>FSO, RM,</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>safety as they relate to anatomic and clinical pathology at all</td>
<td>LI, P</td>
<td></td>
</tr>
<tr>
<td>levels.</td>
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</table>
**PROGRAM STRUCTURE**

**Overview of Training**

The Pathology Training Program offers 16 positions for residents on service-based rotations. Approximately four residents enter the program each year. In addition, there are 1-3 Cytopathology Fellows, 1-2 Surgical Pathology Fellows, 1-2 Hematopathology Fellows and 1-3 Post-Sophomore Fellows. The program offers a four-year, combined Anatomic Pathology/Clinical Pathology (AP/CP) program. The AP/CP program contains a core program consisting of 26, four-week blocks of AP, 20, four-week blocks of CP and six, four-week-blocks of electives for a total of 48 months of full-time pathology service.

The faculty works with the residents to design an elective program that will provide a sound educational experience in the resident's chosen area of concentration, as well as to flexibly adapt to the resident's level of expertise and career goals. During the elective time, residents are encouraged to spend time carrying out basic or applied research and/or time pursuing subspecialty training. We encourage residents to elect a combined program in AP/CP because we feel that there is a considerable overlap between many of the traditional areas of anatomic pathology and laboratory medicine.

**GENERAL PROGRAM OUTLINE**

| Core Anatomic Pathology  
(26, four-week blocks) | Core Laboratory Medicine  
(20, four-week blocks) | Electives  
(Six, four-week blocks) |
|-------------------------|-------------------------|-------------------------|
| • Surgical Pathology*  
  (18) | • Hematopathology  
  (3) | • Anatomic Pathology  
  Subspecialty |
| • Forensic Pathology  
  (1) | • Microbiology/Immunology/Virology  
  (3) | • Clinical Pathology  
  Subspecialty |
| • Cytopathology  
  (4) | • Transfusion Medicine  
  (3) | • Research |
| • Dermpath/Neuropath  
  (1) | • Chemistry  
  (2) | |
| • Pediatric Pathology  
  (1) | • Cytogenetics  
  (1) | |
| • Surg. Path Trainer  
  (.5) | • Molecular Pathology  
  (1) | |
| • Renal Path/EM  
  (.5) | • Flow Cytometry/Wet Heme  
  (2) | |
| | • Lab Management/Informatics  
  (1) | |
| | • VACP Integrated Clinical Pathology  
  (4) | [Hematopathology, Chemistry, Lab  
  Management] |

*Estimated number of blocks on Surgical Pathology; additional blocks of Surgical Pathology may be required as determined by Resident Education Committee (REC).
## PROGRAM OUTLINE

<table>
<thead>
<tr>
<th>Rotation</th>
<th>Required</th>
<th>PGY1</th>
<th>PGY2</th>
<th>PGY3</th>
<th>PGY4</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>KU Surgical Pathology/Autopsy</td>
<td>12 blocks</td>
<td>4</td>
<td>4</td>
<td>3</td>
<td>1</td>
<td>12</td>
</tr>
<tr>
<td>VA Surgical Pathology</td>
<td>6 blocks</td>
<td>5</td>
<td>1</td>
<td></td>
<td></td>
<td>6</td>
</tr>
<tr>
<td>Forensic Pathology</td>
<td>1 block</td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Cytopathology</td>
<td>4 blocks</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>4</td>
</tr>
<tr>
<td>DermPath/Neuropath</td>
<td>1 block</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Pediatric Pathology</td>
<td>1 block</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>KU Surg Path Trainer</td>
<td>.5 block</td>
<td></td>
<td></td>
<td></td>
<td>.5</td>
<td>.5</td>
</tr>
<tr>
<td>Renal Path/EM</td>
<td>.5 block</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>.5</td>
</tr>
<tr>
<td><strong>Total Anatomic</strong></td>
<td><strong>26</strong></td>
<td><strong>10</strong></td>
<td><strong>7</strong></td>
<td><strong>4</strong></td>
<td><strong>5</strong></td>
<td><strong>26</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Rotation</th>
<th>Required</th>
<th>PGY1</th>
<th>PGY2</th>
<th>PGY3</th>
<th>PGY4</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hematopathology</td>
<td>3 blocks</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
<td>3</td>
</tr>
<tr>
<td>Microbiology/Immunology/Virology</td>
<td>3 blocks</td>
<td>1</td>
<td></td>
<td>2</td>
<td></td>
<td>3</td>
</tr>
<tr>
<td>Transfusion Medicine</td>
<td>2 blocks</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td>2</td>
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One week will be arranged for PGY4s to attend MTN for histocompatibility testing training.
PGY-SPECIFIC GOALS

Definition

ACGME Common Program Requirements IV.A.2 states the following: “Competency-based goals and objectives for each assignment at each educational level, which the program must distribute to residents and faculty annually in either written or electronic form. These should be reviewed by the resident at the start of each rotation.”

Purpose

Each rotation has specific competency-based goals and objectives. Additionally, the following list of year-specific goals defines specific goals either in skills, knowledge, or professionalism that are appropriate for each year of training. Residents must strive to achieve these goals as well as the overall educational goals described in the previous pages. Goals may vary somewhat between levels depending on individual rotation schedules.

PGY 1 GOALS: Anatomic Pathology

Autopsy - By the end of the first year:

- The resident has completed their first three autopsies under direct supervision.
- The resident demonstrates a strong knowledge in gross and microscopic anatomy.
- The resident understands and applies the seven components of the autopsy, as appropriate, required by ACGME for participation credit.
- The resident demonstrates the ability to review and extract appropriate information from the clinical history prior to the autopsy.
- The resident recognizes indications for performing an autopsy and recognizes settings where the coroner should be contacted.
- The resident can independently perform a full autopsy, including removal of the brain and be able to correlate gross and microscopic findings with clinical history.

Surgical Pathology - By the end of the first year:

- The resident has documented direct supervision of grossing the first three specimens in the majority of specimens on the organ-based list of specimens.
- The resident can dictate informative gross dictations with proper cassette summaries and cut appropriate sections without direct supervision on all biopsies and simple routine specimens and most common cancer cases.
- The resident becomes efficient in managing cases, such that an appropriate turnaround time is observed.
- The resident can independently perform frozen section procedures and can report frozen section results following joint faculty/resident interpretation of frozen sections.
Cytopathology - *By the end of the first year:*

- Beginning understanding of fundamentals of cytopathology practice.

**PGY 1 GOALS: Clinical Pathology**

**Transfusion Medicine** - *By the end of the first year:*

- The resident should achieve competency in Skill Level 1.

**Chemistry** - *By the end of the first year:*

- The resident should achieve competency in Skill Level 1.

**Laboratory Hematology/Flow Cytometry** - *By the end of the first year:*

- The resident demonstrates the ability properly to work up a bone marrow biopsy. This includes correct identification of cells with appropriate cell count, write up and differential diagnosis.
- Demonstrates basic skills in hematologic correlation through special coagulation test consulting.
- Demonstrates basic skills with interpretation of ancillary data (flow cytometry, cytogenetics, molecular genetics, immunohistochemistry).

**Microbiology/Immunology and Molecular Pathology** - *By the end of the first year:*

- The resident should achieve competency in Skill Level 1.

**Laboratory Management and Informatics** - *By the end of the first year:*

- The resident should achieve competency in Skill Level 1.

**PGY 2 GOALS: Anatomic Pathology**

**Autopsy** - *By the end of the second year:*

- The resident should be able to perform autopsies independently and efficiently with minimal correction by the attending.
- The resident should be able to prepare and discuss autopsy findings at morbidity and mortality conferences, including preparation of gross and microscopic photographs.
- The resident can properly dissect the brain and spinal cord for gross examination independently and identify the majority of gross and microscopic neuropathology.
- The resident is ready to start supervising (in third year) junior residents in autopsy procedures.
**Surgical Pathology** - *By the end of the second year:*

- The resident has documented direct supervision of grossing the first three specimens for all specimens on the organ-based list of specimens.
- The resident demonstrates the ability to work up cases properly, including ordering appropriate histochemical and immunohistochemical stains.
- The resident demonstrates efficiency and professionalism in the handling of cases (turnaround time is kept to 48 hours, special stains, immunostains are ordered when the attending staff requests them, the resident does the follow-up on stains when they do not arrive when anticipated).
- The resident demonstrates an economy of sections that are adequate to provide all the necessary information, and minimizes the need to submit additional wet tissue.
- The resident demonstrates the ability to communicate appropriately to clinical colleagues, including impromptu drop-by visits and in CPC-type conferences.
- The resident is ready to start supervising (in the third year) junior residents in surgical pathology procedures.

**Cytopathology** - *By the end of the second year:*

- The resident demonstrates competency in recognizing inflammatory reactive repair, LGSIL, HGSIL and carcinoma on pap smears and is able to report them out with the latest version of the Bethesda System.
- The resident demonstrates an improvement in medical knowledge in cytology at sign-out.
- The resident is ready to start supervising (in the third year) junior residents in cytology.

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**PGY 2 GOALS: Clinical Pathology**

**Transfusion Medicine** - *By the end of the second year:*

- The resident should achieve competency in Skill Levels 1 and some of Level 2.
- The resident should have worked up patients for autologous donor and pheresis in a competent manner.
- The resident should have completed the rotation at the Community Blood Center.

**Chemistry** - *By the end of the second year:*

- The resident must achieve competency in Chemistry Skill Level 1 and most of Level 2.
- Residents should demonstrate the ability to do method validation, reference intervals and test utilization.
- Residents should be able to demonstrate the ability to perform laboratory accreditation and prepare the laboratory for accreditation.
Laboratory Hematology/Flow Cytometry - By the end of the second year:

- Same skills as Year 1, but be competent in some Skill Level 2 hematology.
- The resident should have familiarity with the clinical presentation and work-up of patients with coagulation problems.
- The resident should demonstrate the ability to synthesize flow cytometry, cytogenetics, and molecular studies with hematology findings.
- The resident should develop competency for Skill Level 1 in Flow Cytometry.

Microbiology/Immunology and Molecular Pathology - By the end of the second year:

- The resident should achieve competency in Skill Level 1 and some of Level 2.

Laboratory Management and Informatics - By the end of the second year:

- The resident should achieve competency in Skill Level 1 and some of Level 2.

PGY 3 GOALS: Anatomic Pathology

Surgical Pathology - By the end of the third year:

- The resident must be able to compose a gross and microscopic surgical pathology report which is ready for electronic signature, with minimal, if any, correction.
- The resident should be comfortable performing independent intraoperative consultation procedures.

Autopsy - By the end of the third year:

- The resident should have completed a rotation at Jackson County Medical Examiner’s Office and be competent in general forensic autopsy skills.
- The resident should have case logs of a minimum of 50 autopsies, of which all 50 autopsies can be shared with one other resident. All seven elements must be documented, except forensic cases where microscopics are taken only when deemed necessary.

Cytopathology - By the end of the third year:

- Residents should be reviewing and preparing for sign-out all types of cytopathology with minimal correction by the attending staff.
- The resident should be able to perform most fine needle aspirations without direct supervision, and produce diagnostic aspirations that are well-preserved, well-stained and with adequate cell button for ancillary studies.
- The resident must develop professionalism and interpersonal and communication skills that are respectful and compassionate toward patients, demonstrating cultural competence.
PGY 3 GOALS: Clinical Pathology

**Transfusion Medicine** - *By the end of the third year:*

- The resident must achieve competency in Skill Level 1.

**Chemistry** - *By the end of the third year:*

- The resident must achieve competency in Skill Level 1.

**Laboratory Hematology/Flow Cytometry** - *By the end of the third year:*

- The resident demonstrates the ability to work up properly a bone marrow biopsy and aspirate, including correct identification of cells, appropriate cell count, write up and differential diagnosis.
- Demonstrates basic skills in hematologic correlation through special coagulation test consulting.
- Demonstrates basic skills with interpretation of ancillary data (flow cytometry, cytogenetics, molecular genetics, immunohistochemistry).

**Microbiology/Immunology and Molecular Pathology** - *By the end of the third year:*

- The resident should achieve competency in Skill Level 1 and most of Level 2.

**Laboratory Management and Informatics** - *By the end of the third year:*

- The resident should achieve competency in Skill Level 1 and most of Level 2.

PGY 4 GOALS: Anatomic Pathology

**Surgical Pathology**

- The resident should be able to supervise junior residents in all aspects of the practice of surgical pathology.
- The resident should have demonstrated increased medical knowledge through performance on study set examinations, conferences and conducting clinical conferences.
- The resident is practice ready for billing, Medicare compliance and accreditation issues.
- The resident should have completed subspecialty related rotations including dermatopathology and pediatric pathology at Children's Mercy Hospital.
- The resident must document minimally 200 intraoperative consultations/frozen sections.
- The resident must document minimally 2,000 surgical pathology cases that they have reviewed and signed out.
- The resident should be competent in all Surgical Pathology Skill Levels 1 and 2.
Cytopathology

- The resident must have reviewed a minimum of 1,500 cytologies (pap smears, nongynecologic exfoliatives and fine needle aspirations).
- The resident should be able to supervise junior residents in all aspects of cytopathology.
- The resident is practice ready for billing, Medicare compliance and accreditation issues.
- The resident should have received training in ThinPrep and is eligible for ThinPrep certification.
- The resident should be competent in all Cytopathology Skill Levels 1 and 2.

Autopsy

- The resident should have completed all autopsy training, including the Medical Examiner's rotation.
- The resident should have minimally fifty (50) autopsies with gross and microscopic examination (see Year 3 goals; microscopics are taken as indicated on forensic cases).
- The resident should be competent in all Autopsy Skill Levels 1 and 2.

| PGY 4 GOALS: Clinical Pathology |

Transfusion Medicine

- The resident should be competent in all Transfusion Skill Levels 1 and 2.

Chemistry

- The resident should be competent in all Chemistry Skill Levels 1 and 2.

Laboratory Hematology/Flow Cytometry

- The resident should be competent in all Hematology and Flow Cytometry Skill Levels 1 and 2.

Microbiology/Immunology and Molecular Pathology

- The resident should be competent in all Microbiology, Immunology, Cytogenetics and Molecular Skill Levels 1 and 2.

Laboratory Management and Informatics

- The resident should be competent in all Skill Levels 1 and 2 for Laboratory Management and Informatics.
GENERAL RESIDENCY GOALS

Throughout the entire duration of residency training, the resident must also demonstrate the specific skills listed below in addition to obtaining competences as described in the previous pages for overall educational goals.

Professionalism:

- The resident has demonstrated professional conduct with regard to interpersonal interaction with peers (pathologists and clinicians), with clerical staff, histotechnologists, medical technologists (CLS), laboratory assistants, autopsy technicians, the Residency/Fellowship Program Coordinator, laboratory administration, and all other employees.
- The resident must have learned to assume responsibility over their cases. The resident must demonstrate a prioritization of educational mission, with the willingness and appreciation of teaching from attending staff, fellows and senior residents and other para-health professionals.
- Part of professionalism is checking e-mail daily and responding in a timely manner, entering duty hours and completing documents in a timely manner, performance of surveys in a timely manner and checking and emptying their mail box.
- The resident should keep ACGME Case Log updated.
- The resident must attend required lectures and conferences.
- The resident must keep certifications (e.g. BLS, ACLS, as required) and licensure (as applicable) current and renewed, without lapses.
- The resident has taken and passed the United States Medical Licensing Examination (USMLE) Step 3 by December of their PGY2 year.

Practice-Based Learning and Improvement:

- The resident must demonstrate self-motivation in the desire to critically review their work to continually find ways of improving their clinical and diagnostic skills. This includes demonstration of adequate and appropriate review of the literature, the ability to use a library or internet to investigate topics.
- The resident must submit a manuscript for publication or present an abstract a meeting prior to the completion of the program.
- The resident must participate in patient safety conferences.
- While on CP rotation, the resident must attend and participate in the Laboratory Quality Assurance monthly meeting.
- Each resident must complete a minimum of one Practice-Based Learning and Improvement Project (PBLI) project prior to graduation.
Interpersonal and Communication Skills:

- The resident must demonstrate growth in areas of interacting with peers and with attending staff. They should demonstrate understanding in what is told to them by appropriate and timely follow-up on assigned duties.
- The resident should be able to clearly communicate in a manner that is professional to clinicians, that is, to communicate reports from written reports, rather than from guessing from memory, and to ask them to read-back the report.
- The resident should seek help from attending staff when it is appropriate.

Systems-Based Practice:

- The resident must take the College of American Pathologists (CAP) inspector on-line training program.
- The resident must participate in CAP self-inspections and mock inspections at KUMC.
- When possible, the resident should also participate in external inspections.
- The resident must participate in level appropriate interdisciplinary conferences.
Residents in pathology acquire healthcare skills through a combination of learning theory and practicing in the workplace. PBLI is one of the six ACGME competency areas for all physicians in training. The primary goal of PBLI is for residents to demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practice. Limiting residents to an observational role in improvement activities can impede their attaining competency in PBLI. To ensure active participation by residents in systems-based quality improvement efforts, the training program requires all residents complete a minimum of one project in PBLI prior to graduation. PBLI projects may fall into one of two categories:

1. Clinical: Improves the clinical practice in which the resident is involved during training.
2. Educational: Improves the educational portion of the resident program or other educational program.

**RECENT PBLI PROJECTS**

- Creation of transfusion medicine practice guidelines manual for KU hospital staff.
- General protocol and survival guide for first year residents in pathology.
- Guide for handling Midwest Transplant Network after-hours specimens.
- Resident slide study set.

**PBLI PROCEDURE**

The resident should identify an area for improvement in the clinical practice or educational program through their work-based practice. They should then assemble a team to address the problem. The team must include at least one faculty member or laboratory administrator to help facilitate the project. The team may include multiple residents, faculty, administrators and hospital staff. The team should hold formal meetings to describe the problem, identify tools for measurement of the problem, establish a timeline for data collection, describe the intervention and outcomes, and provide a narrative of analysis and conclusions with applications to current practice and future steps for continued improvement at the completion of the project.

**RESIDENT PBLI EVALUATION**

At the completion of the project, a standard PBLI evaluation form signed by the resident should be submitted to the Residency Program Director. This form is available in the residency Dropbox account.

**RECOMMENDED READING LIST:**

https://www.dropbox.com/home/PBLI


*Approved by REC, 6.25.2015*
**DIDACTIC SESSIONS AND CONFERENCES**

In addition to training through rotation experiences, residents are expected to participate in scheduled didactic sessions, scholarly activities, teaching rounds and educational slide conferences. Required didactic sessions and conferences in a given block are dependent upon the resident rotation, as illustrated in the block diagram below (shaded block indicates required attendance):

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**Outside rotations**, including community blood bank, Peds, molecular at CMH, and North Kansas

Only AP/CP conference is required

Attendance is tracked by the Chief Residents and the Residency/Fellowship Program Coordinator. The program expects residents to attend all required conferences (100%) for a rotation unless they are on leave or there is a critical patient care issue that cannot be delayed. Conference attendance is reviewed on a semi-annual basis by the Residency Program Director. Semi-annual attendance levels below 80% will result in disciplinary action such as loss of educational funds for the next academic year (the 80% threshold accounts for absences due to sick leave, vacation, or out of town rotations).

Outside rotations, including Community Blood Center, pediatric surgical pathology and molecular pathology at Children's Mercy Hospital, and North Kansas City: only the AP/CP conference (resident lunch) is required (one conference all month).

*Approved by REC, 6.25.2015*
RESIDENT SCHOLARLY ACTIVITY

Residents are encouraged to participate in scholarly activities including method development, clinical or basic research, or literature reviews. Publishing a peer-reviewed article is considered as important educational experience and all residents are expected to publish a minimum of one manuscript or present at least one abstract at a national meeting during their training.

Resident Travel Funds

Each resident may apply for funding to attend up to two regional/national scientific meetings during their four-year residency program contingent upon availability of funds as determined by the Chair of the Department. Funding will be contingent upon the resident being in good standing by the Residency Program Director. The following guidelines must be met for funding consideration:

- The resident must be the presenting author of a poster or talk related to work performed within the Departments of Pathology at KUMC or the Kansas City VA.
- Funding will be provided for travel expense, lodging, food, and meeting registration, not to exceed $1,500/meeting (receipts are required for reimbursement).
- The resident must arrange for time-off of the scheduled rotation during the time of the meeting. If the resident is on a rotation that requires resident service, it is the responsibility of the resident to find coverage for the service (as approved by the Chief Residents and Residency Program Director).
- Funding for more than two meetings during the duration of the residency program will be considered on an individual basis. In such cases, the strength of the scientific project will be reviewed by the Residency Program Director and final approval will be determined by the Department Chair based upon availability of funds.
- Funding does not apply for expenses occurring after completion of the residency program.

Resident Educational Funds

- Educational funds are limited to program-related expenses. Each resident is allocated $1,000 for educational development per academic year. These funds do not carry over to the following year. Funding will be contingent upon the resident being in good standing (including adequate conference attendance) as deemed by the Residency Program Director. Accepted uses of allocated educational funds are listed in the following table. Allocated educational funds may not be used for travel reimbursement to rotational sites or American Board of Pathology (ABP) board examination fees.

- At the Residency Program Director and Department Chair’s discretion, residents may also accrue educational funds by covering Midwest Transplant Network (MTN) frozen sections after hours. PGY1 residents may not be on-call but are eligible for the funds if they receive an MTN case between the hours of 5 p.m. to 9:30 p.m. (routine scheduled shift extends to 9:30 p.m.) on their Day 1 frozen section day and take primary responsibility for the frozen procedure and case.
Resident educational funds will be credited $200 per call event for these cases outside of the hours of 8 a.m. to 5 p.m., Monday-Friday or anytime weekends and holidays. Resident must email the scanned PDF MTN form (copies in gross room) to Brian Donowa bdonawa@kumc.edu and Teal Shultz tshultz2@kumc.edu within one business day of performing the frozen to ensure MTN account is credited.

MTN funds will carry over from year to year. If not used by June 15th of the PGY4 year, they are forfeited. MTN fees may be used for educational fund expenses as well as travel reimbursement to rotational sites and ABP board examination fees.

- PGY2 and above residents must have a temporary Missouri medical license. Residents may use educational funds to apply for the temporary Missouri license. The resident is responsible for the cost of the first year of Missouri licensing. The department will pay the cost for renewal of the temporary license in years PGY3 and PGY4.

- Requests for reimbursement must include itemized receipts. Requests for reimbursement must be submitted by June 15th of each academic year for educational funds, and no later than June 15th of the PGY4 year for MTN Funds. Covered items are listed below, divided by tax status:

<table>
<thead>
<tr>
<th>Nontaxable Items</th>
<th>Taxable Items</th>
</tr>
</thead>
<tbody>
<tr>
<td>Professional journals and books</td>
<td>USMLE/COMLEX Step 3 exam fees</td>
</tr>
<tr>
<td>Expenses to attend pathology meetings</td>
<td>ABP Board examination permit fees <strong>(MTN only)</strong></td>
</tr>
<tr>
<td>Missouri temporary license</td>
<td></td>
</tr>
<tr>
<td>Board review courses/materials</td>
<td></td>
</tr>
<tr>
<td>Travel reimbursement to rotational sites <strong>(MTN only)</strong></td>
<td></td>
</tr>
</tbody>
</table>

**USMLE STEP 3 POLICY**

Residents must take the USMLE Step 3 exam before January 1 of their PGY1 year. Residents who are unsuccessful at passing USMLE Step 3 by that time will be placed on a remediation program and must re-take the test at the earliest opportunity. Residents must demonstrate successful completion of USMLE Step 3 by January 1 of their PGY2 year.

Residents who are unsuccessful in passing USMLE Step 3 by January 1 of their PGY2 year, will be placed on academic probation. Failure to complete the terms of academic probation may result in termination or non-renewal of their appointment contract.

*Approved by the REC, 10.23.2014*
PROFESSIONALISM

All residents are expected to demonstrate a commitment to carrying out the following professional responsibilities.

Licensing, Certification, Examinations, and Credentialing

- Resident completes and passes Step 3 of USMLE prior to January 1 of PGY2.
- Resident performs at expected level on in-service and other objective examinations.
- Resident maintains an up-to-date portfolio.

Honesty, Integrity, and Ethical Behavior

- Resident behaves truthfully and understand the concepts of ethical behavior.
- Resident seeks counsel when unethical behavior is suspected.
- Resident is truthful, acknowledges personal near misses and errors and puts the needs of patients first.
- Resident engages in ethical behavior.

Humanistic Behaviors of Respect, Compassion, and Empathy

- Resident understands and demonstrates the concepts of respect, compassion, and empathy.
- Resident models respect, compassion and empathy in complex situations.

Responsibility and Follow Through on Tasks

- Resident dependably completes assigned tasks in a timely manner, e.g., logging of duty hours.
- Resident assists team members when requested.
- Resident respects assigned schedules.
- Resident anticipates team needs and takes leadership role to independently implement solutions.

Giving and Receiving Feedback

- Resident accepts feedback constructively and modifies practice in response to feedback.
- Resident is able to provide constructive feedback
- Resident exemplifies giving and receiving constructive feedback
- Resident encourages and actively seeks feedback to improve performance.

Responsiveness to Each Patient’s Unique Characteristics and Needs

- Resident respects diversity, vulnerable populations, and patient autonomy.
- Resident embraces diversity and respects vulnerable populations.
- Resident is aware of potential for bias to affect clinical care.
- Resident demonstrates cultural competency.
- Resident identifies and avoids biases affecting clinical care.
Personal Responsibility to Maintain Emotional, Physical, and Mental Health

- Resident is aware of importance of emotional, physical, and mental health and issues related to fatigue/sleep deprivation.
- Resident manages emotional, physical, and mental health and issues related to fatigue/sleep deprivation, especially in stressful conditions.
- Resident exhibits basic professional responsibilities such as timely reporting for duty rested, ready to work, and is always appropriately dressed.
- Resident recognizes signs of impairment in self and others and facilitates seeking appropriate help when needed.
ELIGIBILITY AND SELECTION OF RESIDENTS

The following is the Department of Pathology & Laboratory Medicine’s policy regarding selection procedures for the appointment of residents to the program.

Initial Application Screening

The initial screening of applicants is done by the Residency/Fellowship Program Coordinator or Residency Program Director or Assistant Directors. As applications are received, the following are outlined for review.

1. **USMLE** or Comprehensive Osteopathic Medical Licensing Examination of the United States (COMLEX) scores. First attempt pass rate and scores greater than 200 are preferred.

2. **Year of Graduation.** If greater than 10 years, the type of work the candidate has been engaged in since graduation from medical school is noted.

3. **Experience following graduation.** Experience, either by education or work experience, in the field of pathology is noted.

4. **Personal Statement.** The applicant’s personal statement is evaluated on the following:
   - Command of the English language.
   - Stated genuine interest in pathology.
   - Overall quality of the statement.
   - Dean's letter.
   - Medical transcripts.
   - Letters of reference.
   - Any potential items for concern.

Additional Screening

If the Residency Program Director is unable, after the secondary screening to make a decision on whether or not to invite a candidate to interview, the application is sent to an Assistant Residency Program Director or another member of the REC for their review. After receiving feedback from the committee reviewer, the Residency Program Director will decide whether or not to extend an invitation to the candidate.

Each candidate that is selected for interview will be invited via email by the Residency/Fellowship Program Coordinator. Once scheduled, an email with an interview confirmation and instructions for the interview day is sent to the candidate.

Interview Process

Three interview dates are selected in November-January of each year with up to 16 candidates interviewed per interview day. Four faculty members and one Chief Resident interview all applicants. At the beginning of each interview day, an overview of the institution and program is presented and interviewers are given all application materials for each applicant to be interviewed.
Each interviewer is asked to complete a resident candidate evaluation form and also an individual ranking form for each candidate they interview. Interviewers are asked to assign each a quartile based on every applicant they have ever interviewed. In February, an annual ranking meeting is held with all faculty and resident interviewers and any other faculty who wish to attend. Each applicant is discussed in detail. After initial grouping into “Upper, Middle, or Lower Thirds,” the final rank list is determined by the committee.
GENERAL PROGRAM EVALUATIONS

RESIDENT EVALUATIONS

Near the end of each rotation block, an evaluation form is sent electronically (via MedHub) to each faculty member and selected technologists, pathology assistants and morticians (multi-source evaluations) on every resident with whom they have had educational interactions during the rotation block. A list of faculty members who will evaluate residents on each rotation has been developed and a tracking mechanism is used to ensure that all evaluations have been obtained. Evaluations are based on the ACGME six competencies. Faculty members are also encouraged to give immediate, verbal feedback to the residents.

Evaluation forms are placed in the resident's electronic file and are open for examination by the resident at any time. Resident evaluations are reviewed by the Residency Program Director and are summarized for the Clinical Competency Committee (CCC) meeting at least semi-annually. If a problem with performance is identified for any resident, the Residency Program Director or designee immediately meets with the resident to discuss the issues and develop a plan of action. If there are no problems with the performance, residents review and sign their evaluations at the time of evaluation release or at their semi-annual review meeting with the Residency Program Director.

The CCC may recommend resident-specific remediation and disciplinary measures to the Residency Program Director. Clinical faculty members meet at the end of the academic year to decide on promotion for each resident. All evaluations, performance on exams, attendance at conferences and overall performance are discussed with each resident at the semi-annual evaluation meetings with the Residency Program Director. At that time, residents are asked to write a self-assessment and goals for the next six months. Residents are asked at every six-month evaluation to provide suggestions for program improvement.

The Residency Program Director completes a summative evaluation for each resident finishing the program. The final evaluation summarizes all aspects of the resident's education and training, verifies that the resident is competent in the six general competencies and confirms that the resident has the ability to practice without direct supervision. The final evaluation letter is maintained in the resident's file.

PATHOLOGY MILESTONES

Resident progress is evaluated using pathology milestones developed by the ACGME and the ABP. These milestones encompass skills in both Anatomic Pathology (AP) and Clinical Pathology (CP).

There are a total of 27 milestones divided into six areas of core competency, which are summarized below. Each milestone has specific skills associated with various levels of achievement (Level 1 through 5). It is expected that residents will progress through these levels of achievement during their training. In general, the level of achievement corresponds to the skills associated with that year of training (i.e. a PGY-2 resident would achieve Level 2 skills in most milestones).

Milestones are assessed twice a year, both by the resident (self-assessment) and by the program (faculty members on the CCC). The results of these assessments are discussed by the Residency Program Director with the resident at every semi-annual evaluation.
MILESTONES

#1: Patient Care

1. Consultation: Analyzes, appraises, formulates, generates, and effectively reports consultations (AP/CP).
2. Interpretation and reporting: Analyzes data, appraises, formulates, and generates effective and timely reports (CP).
3. Interpretation and diagnosis: Demonstrates knowledge and practices interpretation and analysis to formulate diagnoses (AP).
4. Reporting: Analyzes data, appraises, formulates, and generates effective and timely reports (AP).
5. Surgical Pathology grossing: Demonstrates attitudes, knowledge, and practices that enables proficient performance of gross examination (AP).
7. Procedures: If training program teaches other procedures (e.g. bone marrow aspiration, apheresis, fine needle aspiration, ultrasound guided FNA, etc.) (AP/CP).

#2: Medical Knowledge

1. Diagnostic knowledge: Demonstrates attitudes, knowledge, and practices that incorporate evidence-based medicine and promote life-long learning (AP/CP).
2. Teaching: Demonstrates ability to interpret, synthesize, and summarize knowledge, teaches others (AP/CP).

#3: Interpersonal and Communication Skills

1. Intra-departmental interactions and development of leadership skills: Demonstrates attitudes, knowledge, and practices that promote safe patient care through team interactions and leadership skills within the laboratory (AP/CP).
2. Inter-departmental and healthcare clinical team interactions: Demonstrates attitudes, knowledge, and practices that promote safe patient care through interdisciplinary team interactions (AP/CP).

#4: Problem Based Learning and Improvement

1. Recognition of errors and discrepancies: Demonstrates attitudes, knowledge, and practices that permit improvement of patient care from study of errors and discrepancies (AP/CP).
2. Scholarly activity: Analyzes and appraises pertinent literature, applies scientific method to identify and interpret evidence-based medicine and apply it clinically (AP/CP).
#5: Evaluating Residents

1. Patient safety: Demonstrates attitudes, knowledge, and practices that contribute to patient safety (AP/CP).
2. Lab management: Regulatory and compliance: Explains, recognizes, summarizes, and is able to apply regulatory and compliance issues (AP/CP).
3. Lab management: Quality, risk management, and laboratory safety: Explains, recognizes, summarizes and is able to apply quality improvement, risk management, and safety issues (AP/CP).
4. Lab management - Test utilization: Explains, recognizes, summarizes and is able to apply test utilization (AP/CP).
5. Lab management - Technology assessment: Explains, recognizes, summarizes and is able to apply technology assessment (AP/CP).

#6: Professionalism

1. Licensing, certification, examinations, credentialing: Demonstrates attitudes and practices that ensures timely completion of required examinations and licensure (AP/CP).
2. Professionalism: Demonstrates honesty, integrity, and ethical behavior (AP/CP).
3. Professionalism: Demonstrates responsibility and follow through on tasks (AP/CP).
4. Professionalism: Giving and receiving feedback (AP/CP).
5. Professionalism: Demonstrates responsiveness to each patient’s unique characteristics and needs (AP/CP).
6. Professionalism: Demonstrates personal responsibility to maintain emotional, physical, and mental health (AP/CP).

RECOMMENDED READING LIST:


FACULTY EVALUATIONS

Near the end of each rotation block, every resident receives electronically a rotation evaluation form as well as a faculty evaluation form(s). Names of residents submitting such evaluations are suppressed. Faculty only have access to their evaluations at six month intervals in order to protect confidentiality.

The Residency Program Director has immediate access to all evaluations except their own. If problems are identified, they are discussed immediately with the appropriate faculty member by the Residency Program Director. Faculty evaluations are reviewed by the Residency Program Director every six months unless there are problems, which are addressed sooner.
Faculty members are given an overall assessment including the following:

- Clinical teaching ability.
- Commitment to educational program.
- Clinical knowledge.
- Professionalism.
- Scholarly activity.
- Faculty development.
- Resident evaluations.

The annual assessment is signed by the Department Chair and a copy is sent to the faculty member. The assessment is used by the Department Chair for annual faculty evaluations.

Approved by REC, 6.25.2015
ANNUAL PROGRAM EVALUATIONS

A formal resident training quality improvement program addresses individual resident performance improvement, faculty development and overall training program improvement as described below.

**Overall Goals of Resident Training Quality Improvement Program**

The Pathology Resident Training Quality Improvement Program provides a process for individual resident performance improvement as well as overall program improvement. Measurement tools are used to identify individual residents in need of remediation early in the program. A general remediation program has been developed and is tailored for the individual needs of each resident. Additionally, several measurement tools are used to monitor and identify areas of potential improvement within the overall training program. The Program Evaluation Committee (PEC), consisting of ten faculty members, the two Chief Residents, and the Residency Program Director are all responsible for ensuring the quality of resident education in anatomic and clinical pathology.

**Individual Resident Performance Improvement**

Individual resident performance is evaluated by several measurement tools including, but not limited to, the following:

- **Evaluations by Faculty** – Online evaluations based on the ACGME six competencies are completed by all faculty supervisors at the end of each rotational block.
- **360° Evaluations** – Online evaluation by select medical technology supervisors, pathology assistants, or other technical personnel that work directly with residents. Focuses on professionalism, system-based learning, patient care and interpersonal and communication skills and are completed at the end of each rotation block.
- **National Resident In-Service Exam (RISE)** – Resident performance is compared to peers at the national level in different subspecialty areas of anatomic and clinical pathology.
- **Departmental In-Service Exam** – A department-developed annual written exam covers subspecialty areas similar to the RISE.
- **Annual Practical Exam** – Oral exam for histological description, differential diagnosis and final diagnosis for unknown slides.
- **CAP Inspections** – Residents are evaluated by supervisors and faculty on performance of CAP self-inspection and mock inspections.
- **AP/CP Presentation Performance** – Presentations given by residents at this departmental conference are evaluated by all faculty at the end of the conference. The resident with the highest evaluation for an AP presentation and a CP presentation are given an award at the end of each year.
- **Interdisciplinary Conference Presentations** – Presentations made by residents at interdisciplinary conferences, including but not limited to, hematology conference, tumor board, ENT conference, breast conference, CPC and morbidity and mortality conferences are evaluated by attending faculty.
• **Journal Club Presentations** – Resident presentation and critical evaluation of current journal articles are evaluated by attending faculty.

• **Performance at Unknown Slide Conferences** – Participation in unknown slide conferences including surgical pathology, hematology, dermatopathology and neuropathology conferences are evaluated by attending faculty.

• **Conference Attendance** – Attendance at the required level for specific conferences is monitored. Failure to attend at the required level is considered a problem with competency in professionalism.

Specific areas of weakness identified by any of the above performance tools may result in repeating a rotation. More global unsatisfactory performance areas may result in placement on remediation as determined by the Resident Education Committee (REC).

### FACULTY DEVELOPMENT

- Assessment.
- Resident evaluation of faculty.
- Resident evaluation of rotations.
- RISE results for specific areas of training.
- Resident evaluation of core lecture series.
- Annual evaluation by the Residency Program Director for clinical teaching abilities, commitment to the educational program, clinical knowledge, professionalism, and scholarly activities.
- Faculty development improvement methods.
- Annual discussion of use of evaluations.
- Feedback on resident evaluation of faculty.
- Feedback on resident evaluation of rotations they teach.
- Feedback on RISE results in their area.

### Overall Training Program Quality Improvement Based on Outcome Measurements

The Resident Education Committee continuously monitors the quality of resident training in pathology. Examples of quality monitors and outcome measurement tools used to evaluate the quality of the training program include the following:

- **American Board of Pathology Specialty Exam** – Outcome data over a five-year period for different areas of pathology are provided to the program annually. For each category, the board reports if residents from the individual program performed in the upper, middle or lower third compared to all residents taking the boards. Additionally, the overall pass/fail rate over the past five years is provided for the program as compared to the national rate. This information is used to determine if the program may have specific areas of weakness in training. Program improvements will be investigated for areas in which the residents performed in the lower third.
• **National Resident In-Service Exam (RISE)** – RISE results are provided for several different areas of anatomic and clinical pathology. The overall program percentile performance compared to the national performance is used to identify areas of training in need of improvement.

• **Annual Program Evaluations by Residents and Faculty** – Anonymous electronic evaluations of the residency program by residents and by faculty are performed annually.

• **Departmental In-Service Exam** – Each year, the Pathology Department creates a multiple choice exam made up of 100 questions from the same subspecialty areas as the national RISE. This provides the department two, large objective exams for evaluation of residents. Performance variation within specific areas by the overall group of residents is used to identify potential areas for training improvement.

• **ACGME Survey Results** – Resident responses to strengths and weaknesses within the program are used to identify areas in need of improvement.

• **Senior Exit Program Evaluations** – Graduating residents are required to submit an evaluation of the training program prior to leaving the program. The evaluations are completely anonymous and are reviewed in detail by the REC.

• **Rotation Evaluations** – Each resident evaluates their rotation online at the end of each block. These evaluations are initially reviewed by the Residency Program Director on a biannual basis and then a summary is presented to the REC for use in program improvement.

• **Year-end Faculty Evaluations** – Each resident evaluates online, faculty they are in contact with monthly. The evaluations are reviewed by the Residency Program Director and if any problem is identified, it is reported to the Department Chair. The Chair may also receive a copy of de-identified evaluations for use in the annual faculty review.

• **Six-month Meetings with Residency Program Director** – As part of the six-month evaluation process, each resident is asked if they have any suggestions for program improvements.

• **Annual Curriculum Review** – In addition to monthly program review, a formal curriculum review is performed annually by the REC. An announcement is made to all faculty members that program improvement suggestions are welcomed.

• **Resident Monthly Meeting** – At each monthly meeting, which is attended by all residents, the Residency Program Director and Assistant Program Directors, the Chief Residents lead discussions on any program issues. Recommendations from this meeting are taken to the REC by the Residency Program Director.

• **Ad Hoc Projects and Task Forces** – Depending upon need, ad hoc task forces or subcommittees are formed to address specific program improvement questions.

• **Program Evaluation Committee** – An annual program evaluation is performed, with the committee meeting every summer. This annual review examines and summarizes any needs for improvement in program quality, resident performance, faculty development, or graduate performance.

**PROMOTION**

Residents are evaluated by their attending staff and at the end of each rotation. Evaluations are available for review on-line when completed. Residents being evaluated will receive an email notification when an evaluation is
completed. Residents are also evaluated by technologists, pathology assistants and autopsy assistants. These evaluations are to ensure that residents are progressing satisfactorily from rotation to rotation and that deficiencies relative to promotion to the next PGY level, if present, can be addressed as soon as possible.

In addition to rotation evaluations, information from other sources will be considered. These include attendance records for required academic sessions, results of written examinations, and informal reports. Residents are reviewed as to performance by the Residency Program Director at least twice, yearly. Residents are also reviewed by the clinical faculty at an annual meeting.

REMEDICATION, PROBATION, CORRECTIVE ACTION

Concerns regarding any aspects of a resident’s performance are brought before the department's Resident Education Committee (REC). One or two low satisfactory grades will result in informal counseling. A poor grade or unsatisfactory rotation evaluation will result in formal counseling, which may include development of a remediation plan, repetition of the rotation or probation. Consistently poor performance may suggest a need for adverse action. Very specific guidelines from the University of Kansas School of Medicine govern remediation, probation, and due process/grievance procedures pertaining to any such actions.

Whenever the Residency Program Director is informed of a significant concern regarding a resident’s performance, the resident involved will be contacted and given the opportunity to provide a response. The resident may provide this response by any or all of the following:

- In writing; or
- Verbal communication with the Residency Program Director; or
- Appearance before the REC.

The REC will subsequently review the facts and make a decision as to whether this information should be included in the resident's permanent file. If a decision is made to place the material in the resident's file, both criticism and response will be included. Supervising faculty may include any correspondence regarding concerns about resident's performance, with a proviso that same not be placed in the resident's file if difficulties are corrected within a given time frame.

GRIEVANCE PROCEDURE

Grievance matters are those relating to the interpretation of, application of, or compliance with the provisions of the GME Resident Agreement, the policies and procedures governing graduate medical education, and the general policies and procedures of the University of Kansas Medical Center (KUMC). Questions of capricious, arbitrary, punitive or retaliatory actions or interpretations of the policies governing graduate medical education on the part of any faculty member or officer of the Department of Pathology & Laboratory Medicine’s residency program are subject to the grievance process.

Complaints of illegal discrimination, including failure to provide reasonable accommodations, and sexual harassment, are processed in accordance with the Medical Center policies and procedures that are administered through the Equal Opportunity Office. Should a housestaff in the Department of Pathology & Laboratory Medicine have a grievance or be dissatisfied with any aspect of the program, he/she is encouraged to initially discuss the issue with his/her attending or the Chief Residents. If this is felt by the resident to be inappropriate or the issue is not satisfactorily resolved, timely discussion with the Residency Program Director is highly recommended.
Documentation of the issues and a statement of dissatisfaction by the aggrieved resident may be helpful, and is also encouraged, particularly when making an appeal to the Department's REC. In general, the resident will first discuss any grievance with the Chief Residents. If this fails to provide adequate closure to the grievance, then he/she is directed to speak with the Residency Program Director. Issues can best be resolved at this stage and every effort should be made to achieve a mutually agreeable solution. If the grievance is not resolved to the satisfaction of the resident after discussion with the Residency Program Director, the resident has the option to present the grievance, in writing, to the Office of Graduate Medical Education.

In situations where the grievance relates to the Chair or Residency Program Director, or where the resident believes that a fair resolution cannot be attained by presenting the grievance to those individuals, he/she may present the grievance in writing directly to the Office of Graduate Medical Education. The Associate Dean for Graduate Medical Education will meet with the resident, the Residency Program Director, the Chair and one or more of the program's Chief Residents to determine the cause and validity of the complaint and to determine the means of redress. Should the meeting with the Associate Dean fail to resolve the grievance to the satisfaction of the resident, the resident may request that he/she be heard by the Executive Dean. Any action(s) taken in good faith by the Executive Dean addressing the grievance will be final.
RESIDENT WORK ENVIRONMENT

WORK ENVIRONMENT

The Department of Pathology & Laboratory Medicine’s Residency Program and the University of Kansas School of Medicine are committed to promoting patient safety and resident well-being in a supportive educational environment. An appropriate ratio of education to service is ensured by providing a blend of supervised patient care responsibilities, clinical teaching, and didactic education. The program provides an educational and working environment in which residents may address concerns in a confidential and protected manner. Residents are integrated and actively participate in interdisciplinary clinical quality improvement and patient safety programs.

A culture of professionalism supports patient safety and personal responsibility. Appropriate educational resources are provided including medical information access, faculty supervision, and a wide variety and volume of both anatomic and clinical pathology cases. Residents are exposed to, and encouraged to participate in scholastic activities. Graded and progressive clinical responsibility within the supportive educational environment assures resident development of sufficient competence to enter practice without direct supervision upon completion of the program.

The Pathology Residency Program and the University of Kansas School of Medicine will:

a) Provide a stipend and benefits to the resident as stipulated in the applicable GME Resident Agreement;

b) Use its best efforts, within the limits of available resources, to provide an educational training program that meets the ACGME’s accreditation standards;

c) Use its best efforts, within the limits of available resources, to provide the resident with adequate and appropriate support staff and facilities in accordance with federal, state, local, and ACGME requirements;

d) Orient the resident to the facilities, philosophies, rules, regulations, procedures and policies of KUMC, School of Medicine, Department of Pathology & Laboratory Medicine, the Pathology Residency Program and to the ACGME’s and the RRC’s Institutional and Program Requirements;

e) Provide the resident with appropriate and adequate faculty and medical staff supervision and guidance for all educational and clinical activities commensurate with an individual resident’s level of advancement and responsibility;

f) Allow the resident to participate fully in the educational and scholarly activities of the program and KUMC and in any appropriate institutional medical staff activities, councils and committees, particularly those that affect Graduate Medical Education and the role of the resident staff in patient care subject to these policies and procedures;

g) Through the officers of the program and the attending medical staff, clearly communicate to the resident any expectations, instructions and directions regarding patient management and the resident's participation therein;
h) Maintain an environment conducive to the health and well-being of the resident;

i) Within limits of available resources, provide:
   i. Adequate and appropriate food service and sleeping quarters to the resident while on-call or otherwise engaged in clinical activities requiring the resident to remain in KUMC overnight.
   ii. Provide personal protective equipment including gloves, face/mouth/eye protection in the form of masks and eye shields, and gowns. The Occupational Safety and Health Administration (OSHA) and the Centers for Disease Control (CDC) assume that all direct contact with a patient's blood or other body substances are infectious. Therefore, the use of protective equipment to prevent parenteral, mucous membrane and non-intact skin exposures to a healthcare provider is recommended.
   iii. Patient and information support services.
   v. Uniform items, limited to scrub suits and white clinical jacket.

j) Through the Residency Program Director and program faculty, evaluate the educational and professional progress and achievement of the resident on a regular and periodic basis. The Residency Program Director shall present to and discuss with the resident a written summary of the evaluations at least semi-annually;

k) Provide a fair and consistent method for review of the resident’s concerns and/or grievances, without the fear of reprisal;

l) Provide residents with an educational and work environment in which residents may raise and resolve issues without fear of intimidation or retaliation including the following mechanisms:
   i. The GME office ensures that all programs provide their residents with regular, protected opportunities to communicate and exchange information on their educational and work environment, their programs, and other resident issues, with/without the involvement of faculty or attending. Such opportunities include, but are not limited to, confidential discussion with the Chief Residents, Residency Program Director, and Department Chair. Other intradepartmental avenues to confidentially discuss any resident concern or issue occur during the annual program evaluations completed by each resident and/or through discussion with the resident representative during the required annual program review;
   ii. The internal review process, during which residents in each program are afforded the opportunity to discuss their concerns about their programs with a resident from another program and have them presented confidentially to the GMEC;
   iii. An ombudsman, the Assistant Dean for GME Administration, or any other member of the GME staff, including the Executive Vice Chancellor, Senior Associate Dean and the Associate Dean, who are available for the residents to bring any issues raised in these protected resident meetings, or any other issues a resident may need to address;
   iv. Peer leadership and membership of the University of Kansas School of Medicine Resident’s Council, who are available to confidentially receive any resident concern and present their concerns to the Graduate Medical Education Committee and GME Staff;
v. Praise and concern comments can be sent through MedHub’s ‘Messaging’ directly and confidentially to the Residency Program Director, or to KUMC’s Designated Institutional Official (DIO). This can be accessed through any resident, MedHub home page.

vi. ACGME Resident Survey, administered directly to all residents in ACGME-accredited programs. This survey provides summary and anonymous feedback to program and GME leadership. For programs with less than four residents, the GME Resident Survey, which is a confidential, anonymous survey organized by the GME office, is administered annually;

vii. A grievance process, which provides the resident with a formal mechanism for addressing serious concerns within their programs, is outlined on page 37 of this manual;

viii. ACGME Department of Resident Services at residentservices@acgme.org or by phone (312) 755-7498 is available if the above described avenues have not satisfactorily addressed a specific resident issue. The ACGME Resident Services representative will work with the DIO to resolve issues and associated concerns. Valid complaints are processed by Resident Services and will require a response from the Residency Program Director and attestation to the response by the DIO, and review by the relevant review committee.

m) Upon satisfactory completion of the program and satisfaction of the program's requirements and the resident's responsibilities delineated herein, furnish to the resident a certificate of completion of the program;

n) Annually review and approve the number of residents and funding sources for each program and discuss these quotas and sources of funding with the Department Chair and Residency Program Director in a timely fashion so as to facilitate the recruitment and retention of residents;

o) Provide the agreed upon levels of financial support, subject to the terms of the resident contract; and

p) Exercise all rights and responsibilities expressed and implied by the “Institutional Requirements” of the ACGME.
SUPERVISION OF RESIDENTS

All work performed by residents is performed under supervision of attending faculty. All procedures performed in autopsy, surgical pathology and clinical laboratory medicine are performed under either direct or indirect supervision of an attending faculty member. All at-home call is supervised by faculty members. Resident responsibilities and progression of responsibility is described in each rotation description. More advanced residents are given increased responsibility which will include more time on each procedure or task being indirectly supervised (immediate availability) by the faculty member.

- In the clinical learning environment, each patient must have an identifiable, appropriately-credentialed and privileged attending physician (or licensed independent practitioner as approved by each review committee) who is ultimately responsible for that patient's care.

- This information should be available to residents, faculty members, and patients.
  - **Inpatient:** Patient information sheet included in the admission packet and listed on the “white board” in each patient room.
  - **Outpatient:** Provided during introduction verbally by residents and/or faculty.

- Residents and faculty members should inform patients of their respective roles in each patient's care.

- The program must demonstrate that the appropriate level of supervision is in place for all residents who care for patients.

Methods of Supervision

- Some activities require the physical presence of the supervising faculty member.
- For many aspects of patient care, the supervising physician may be a more advanced resident or fellow.
- Other portions of care provided by the resident can be adequately supervised by the immediate availability of the supervising faculty member or resident physician in his/her “final years of training”, either in the institution, or by means of telephonic and/or electronic modalities.
- In some circumstances, supervision may include post hoc review of resident-delivered care with feedback as to the appropriateness of that care.
- The privilege of progressive authority and responsibility, conditional independence, and a supervisory role in patient care delegated to each resident must be assigned by the Residency Program Director and faculty members.
- The Residency Program Director must evaluate each resident's abilities based on specific criteria and when available, should be guided by specific national, standards-based criteria.
- Faculty members functioning as supervising physicians should delegate portions of care to residents, based on the needs of the patient and the skills of the residents.
- “Residents in their final years of training,” or fellows should serve in a supervisory role of PGY 1 and “intermediate residents” in recognition of their progress toward independence based on the needs of each patient and the skills of the individual resident or fellow.

KUMC Pathology Residency Manual
Levels of Supervision Defined

To ensure oversight of resident supervision and graded authority and responsibility, the program must use the following classification of supervision established by the ACGME.

**Direct Supervision:**
This means the supervising physician is physically present with the resident and patient.

**Indirect Supervision A (with direct supervision immediately available):**
This means the supervising physician is physically within the hospital or other site of patient care, and is immediately available to provide Direct Supervision.

**Indirect Supervision B (with direct supervision available):**
This means the supervising physician is not physically present within the hospital or other site of patient care, but is immediately available by means of telephonic and/or electronic modalities, and is available to provide Direct Supervision.

**Oversight:**
This means the supervising physician is available to provide review of procedures/encounters with feedback provided after care is delivered.
<table>
<thead>
<tr>
<th><strong>RESIDENT REVIEW COMMITTEE-APPROVED LICENSED INDEPENDENT PRACTITIONER SUPERVISOR</strong> (PR VI.D.1)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Although pathology assistants are not licensed independent practitioners, they may be authorized by a department to provide supervision or oversight of dissection of surgical specimens and autopsies. The ultimate responsibility for a patient's care, however, lies with the attending physician, and cannot belong to a pathology assistant.</td>
</tr>
</tbody>
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<thead>
<tr>
<th><strong>OPTIMAL CLINICAL WORKLOAD</strong> (PR VI.E.)</th>
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</thead>
<tbody>
<tr>
<td>The clinical responsibilities for each resident must be based on PGY-level, patient safety, resident education, severity and complexity of patient illness/condition and available support services.</td>
</tr>
</tbody>
</table>

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<thead>
<tr>
<th><strong>MEMBERS OF THE INTERPROFESSIONAL TEAM</strong> (PR VI.F.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Residents must care for patients in an environment that maximizes effective communication. This must include the opportunity to work as a member of effective, inter-professional teams that are appropriate to the delivery of care in the specialty.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>COMPETENCIES TO ALLOW PGY1 RESIDENTS TO PROGRESS TO INDIRECT SUPERVISION</strong> (PR VI.D.5.a)(1)</th>
</tr>
</thead>
</table>
| Each PGY-1 resident must be directly supervised during performance of, at least, three initial procedures in the following areas:  
- Autopsies (complete or limited).  
- Gross dissection of surgical pathology specimens by organ system.  
- Frozen sections.  
- Apheresis.  
- Fine needle aspirations and interpretation of the aspirate.  
A PGY-3 or PGY-4 resident, pathology assistant or attending pathologist may directly supervise the gross dissection of surgical pathology specimens and/or autopsies.  
Blood banking/transfusion medicine fellows, PGY-3 or PGY-4 residents, or attending pathologists may directly supervise apheresis. |

<table>
<thead>
<tr>
<th><strong>DEFINING RESIDENT LEVELS “INTERMEDIATE LEVEL” &amp; “FINAL YEARS OF TRAINING” For establishing the minimum rest period between duty periods.</strong> (PR VI.G. b &amp; c)</th>
</tr>
</thead>
</table>
| PGY-2 residents are considered to be at the intermediate level.  
Residents in the final two years of the program (PGY-3 and PGY-4) are considered to be in the final years of education. |
The Resident Review Committee defines such circumstances as a required continuity of care for a severely ill or unstable patient, or a complex patient with whom the resident has been involved; events of exceptional educational value, or humanistic attention to the needs of a patient or family.

Intermediate residents and residents in the final years of education may stay on duty or return to the hospital to perform intra-operative consultations, apheresis, emergent autopsies (e.g., when a patient’s religion requires rapid burial), fine needle aspirations, immediate evaluation of cytology, transfusion medicine/blood banking emergencies or hematologic emergencies.

**CIRCUMSTANCES WHEN RESIDENTS IN THEIR FINAL YEARS OF EDUCATION MAY REMAIN OR RETURN IN < 8 HOURS**
(PR VI.G.5.c)(1)

<table>
<thead>
<tr>
<th>LEVEL of SUPERVISION</th>
<th>ACTIVITIES /PROCEDURES (as defined by RRC &amp; Program)</th>
</tr>
</thead>
<tbody>
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<td>DIRECT</td>
<td>Three initial procedures in the following areas:</td>
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<td></td>
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<td>- Frozen sections.</td>
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<td>- Apheresis.</td>
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<td></td>
<td>- Fine needle aspirations and interpretation of the aspirate.</td>
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<tr>
<td>INDIRECT A (with direct supervision immediately available)</td>
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</tr>
</tbody>
</table>

**DEFINED MAXIMUM NUMBER OF CONSECUTIVE WEEKS AND MAXIMUM NUMBER OF MONTHS PER YEAR OF IN-HOUSE NIGHT FLOAT**
(PR VI.G.6.)

Not Applicable.

**Program-specific guidelines for circumstances and events in which residents must communicate with appropriate supervising faculty**
(PR VI.D.5)

Not Applicable.

**Source of specific criteria and/or specific national standards-based criteria used to evaluate each resident’s abilities**
(PR VI.D.4.a)

Not Applicable.
<table>
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<tr>
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<tr>
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<td>(with direct supervision immediately available)</td>
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<tr>
<td>INDIRECT B</td>
<td>(with direct supervision available)</td>
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<tr>
<td>OVERSIGHT</td>
<td>(with direct supervision available)</td>
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</table>

**INTERMEDIATE LEVEL RESIDENTS (PGY 2)**

<table>
<thead>
<tr>
<th>LEVEL of SUPERVISION</th>
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<td>(with direct supervision available)</td>
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</table>

**RESIDENTS IN FINAL YEARS OF TRAINING (PGY 3 and 4)**

<table>
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<tr>
<th>LEVEL of SUPERVISION</th>
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<td>(with direct supervision available)</td>
</tr>
<tr>
<td>OVERSIGHT</td>
<td>(with direct supervision available)</td>
</tr>
</tbody>
</table>

Monthly schedules are posted online and provided to all residents and faculty members indicating all residents and faculty working in each anatomic and clinical pathology area. Likewise, on-call residents and faculty are posted online and distributed to all residents and faculty each month.

**Informing patient of resident role:** When residents have direct contact with patients (e.g. fine needle aspiration procedures or apheresis procedures) they must verbally introduce themselves to the patients, identify themselves as pathology residents, and describe their role in the procedure.
TRANSFER OF CARE (HANDOFF PROTOCOL)

To provide safe and effective patient care in pathology, transitions of care (anatomic and clinical pathology specimens/cases) will use effective and structured hand-off procedures.

To minimize patient care transition, residents are assigned to month-long rotations in which they manage individual cases from beginning to end. In certain circumstances, such as end of the month transition in surgical pathology, or when residents are contacted during at home, on-call, the following handoff policies must be followed.

End of Month Handoff in Surgical Pathology

- Departing and arriving residents **MUST** meet face to face to discuss incomplete cases and what each case needs to be signed out.
- The leaving Day 1 resident **MUST** correct the gross description on all cases prior to hand over.
- The status of each incomplete case **MUST** be indicated on the paperwork for the case (e.g. awaiting immunos or special stains, needs Q/A, history etc.).
- Day 2, 3 and 4 leaving residents: All incomplete cases **MUST** have corrected gross descriptions, topography, final diagnosis and tumor checklists entered as much as possible.
- The departing residents are to make themselves available for questions as needed during the first week of the next month. If they are at KUMC and on a light rotation, they are HIGHLY encouraged to finish up their incomplete cases themselves.

For on-call residents, if the resident is called in during the night

- Write the details regarding the call (e.g. patient information, physician information, results, etc.) on the "call details" white board located in the surgical pathology grossing room, **AND**
- Send an e-mail (prior to 8:30 a.m.) with call details to the pertinent resident and attending, both Chief Residents, and the pathology assistant.

For on-call residents, if the resident receives a phone call not requiring coming in

- Send an email (prior to 8:30 a.m.) with the call details to the pertinent resident and attending, both Chief Residents, and the pathology assistant.
- For clinical pathology calls, fill out the transfusion medicine call form the following day with the appropriate details.

In any situation **when clinical care exceeds a resident's ability (knowledge or volume)**, the resident should immediately contact the supervising faculty member and the Chief Resident. The faculty member will advise the Chief Resident if another resident needs to be immediately sought to help with the clinical tasks or if such duties may be delayed until additional resident help is available. All such incidents are recorded by the Chief Resident and reported to the Residency Program Director, who will investigate circumstances leading to the event. The need for intervention with the resident or for process changes with the clinical rotation will be evaluated.
RESIDENT DUTY HOURS AND CALL SCHEDULE

The Pathology Residency Program's resident duty hours are in compliance with the guidelines established by the Accreditation Council for Graduate Medical Education (ACGME) and Section 15, KU School of Medicine, Graduate Medical Education Policy and Procedure Manual, Section 15: Resident Duty Hours and Call Schedules.

Duty Hours

- Duty hours are defined as all clinical and academic activities related to the residency program; i.e., patient care (both inpatient and outpatient), administrative duties relative to patient care, the provision for transfer of patient care, time spent in-house during call activities, and scheduled activities such as conferences.
- Duty hours do not include reading and preparation time spent away from the duty site.
- Duty hours must be limited to 80 hours per week, averaged over a four-week period, inclusive of all in-house call and moonlighting activities.
  - Duty periods of PGY-1 residents must not exceed 16 hours in duration.
- Residents must be provided with one day in seven free from all educational and clinical responsibilities, averaged over a four-week period, inclusive of call.
  - One day is defined as one, continuous 24-hour period free from all clinical, educational, and administrative duties.
  - Adequate time for rest and personal activities must be provided. This should consist of a 10-hour time and MUST have an eight-hour time period provided between all daily duty periods for PGY-1 and intermediate-level (PGY-2) residents.
- At-home call (or pager call): The frequency of at-home call is not subject to the eight hours between duty periods rule. However, at-home call must not be so frequent as to preclude rest and reasonable personal time for each resident.
- Duty hours are monitored weekly by the Residency/Fellowship Program Coordinator. Any duty hour violation is immediately reported to the Residency Program Director, who then contacts the resident to investigate the violation. The Residency Program Director reviews duty hour documentation every six months.
- The Residency Program Director will submit to the Office of Graduate Medical Education, in partnership with the Budget, Reimbursement, Cost Accounting, and Revenue Cycle Office, duty hour reports for each resident in the program.
- The corrected call schedules and resident time records will be used to verify compliance with the duty and call policies, for invoicing affiliate institutions for resident services, and for documentation of the residents' activity reports that must be submitted to the Centers for Medicare and Medicaid Services.

Duty Hour Restrictions

Intermediate residents (PGY2) and residents in the final years of education (PGY3 and 4) may stay on duty or return to the hospital to perform intra-operative consultations, apheresis, emergent autopsies (e.g., when a patient's religion requires rapid burial), fine needle aspirations, immediate evaluation of cytology, transfusion medicine/blood banking emergencies, and hematologic emergencies. In cases where residents return to work in less than eight hours, the resident will be asked to verify the reason for the extended duty hours in MedHub.
The resident is expected to be rested and alert during duty hours, and the resident and resident's attending medical staff are collectively responsible for determining whether the resident is able to safely and effectively perform his/her duties.

If a scheduled duty assignment is inconsistent with the Resident Agreement or the Institutional Duty Hours and Call Policies, the involved resident shall bring that inconsistency first to the attention of the Residency Program Director for reconciliation or correction. If the Residency Program Director does not reconcile or correct the inconsistency, it shall be the obligation of the resident to notify the Department Chair or Associate Dean for Graduate Medical Education to take the necessary steps to reconcile or correct the raised inconsistency.

**On-Call and Resident Time Reporting**

**Note: Pathology residents do not take in-house call.**

At-home call (or pager call) is defined as a call taken from outside the assigned institution.

The frequency of at-home call is not subject to the every-third night or "24+4" limitations. At-home call, however, must not be so frequent as to preclude rest and reasonable personal time for each resident. Residents taking at-home call must be provided with one day in seven completely free from all educational and clinical responsibilities, averaged over a four-week period.

When residents are called-in to the hospital from home, the hours residents spent in-house are counted toward the 80-hour limit. Resident call-backs to the hospital while on home call do not initiate a new off-duty period (i.e., are not subject to the eight hour, between duty periods restriction).

The Residency Program Director and faculty monitor the demands of at-home call, and make scheduling adjustments as necessary to mitigate excessive service demands and/or fatigue. The call schedule and schedule of duty assignments will be published and made available for review by the residents on a monthly basis.
CALL ASSIGNMENTS

Call duties for pathology residents are pager call ("at-home call") only. Call duties include both AP (Anatomic Pathology) and CP (Clinical Pathology) services, and provides coverage for both services 24-hours a day while following call and duty hour limitations.

- PGY2-PGY4 Residents are eligible for all call periods.
- PGY1 Residents are only eligible for call on weekdays, 7:30 a.m. to 9:30 p.m.

**AP WEEKDAY CALL** (7:30 a.m. to 9:30 p.m., Monday-Friday): PGY1 - PGY4.
Covers surgical pathology services (frozen sections, MTN frozen sections, AP questions).

**COMBINED AP/CP WEEKNIGHT CALL** (9:30 p.m. to 7:30 a.m., Monday-Friday): PGY2 - PGY4.
Covers surgical pathology, Cytology, and any CP services (e.g. blood bank, microbiology, etc.).

**CP WEEKDAY CALL** (7:30 a.m. to 9:30 p.m., Monday-Friday): PGY1 - PGY4.
Covers CP service questions (e.g. blood bank, microbiology, etc.)

**COMBINED AP/CP WEEKEND CALL** (9:30 p.m. Friday to 7:30 a.m. Monday): PGY2 - PGY4.
Covers surgical pathology, cytology, and any CP services (e.g. blood bank, hematology, microbiology, etc.).

**AUTOPSY CALL** -- PGY1-PGY2 (PGY3-PGY4 eligible, if needed).
- Monday-Friday daytime schedule.
- Saturday schedule.
- Holiday schedule.

CALL SCHEDULES

The Residency Program Director has final approval over call schedules. While on-call, all residents are under the supervision of a faculty member who is available at all times via phone, pager, or in person. Residents must notify Chief Residents of vacation requests and away rotations as early as possible.

If conflicts arise in any of the call schedules, and the resident is not able to provide scheduled call for any reason, it is the responsibility of the resident to arrange coverage, and notify the Chief Residents and appropriate rotation and scheduling personnel right away.

**Combined AP/CP weeknight and combined AP/CP weekend call schedules**

The AP/CP weeknight and AP/CP weekend call schedules are made by the Chief Residents at the beginning of each academic year. Weeknight and weekend calls are divided among intermediate level (PGY-2) and senior level residents (PGY3-4) as evenly as possible. After March 31st, PGY4 residents are not required to take call unless scheduling emergencies arise.
Weekday AP and CP call schedules are made by the Chief Residents before each block. Weekday AP call schedule (7:30 a.m. to 9:30 p.m., Monday to Friday): PGY1 - PGY4.

1. The Day One resident assigned to Surgical Pathology at KUMC is on-call for weekday AP call that day.

Weekday CP call schedule (7:30 a.m. to 9:30 p.m., Monday to Friday): PGY1 - PGY4.

1. Residents assigned to CP rotations, including VA CP, will be given priority to cover CP Weekday call.

2. If there are not enough CP residents to cover call, residents on in-house electives, research electives or cytology are eligible for Weekday CP call.

3. Unless there are scheduling emergencies, residents assigned to surgical pathology at the VA or KUMC, offsite rotations, and KUMC hematopathology will be excluded from CP Weekday Call.

Autopsy Call

1. All autopsies are performed under the direct supervision of a faculty member.

**Autopsy Weekday Schedule:**

1. The Day Two resident assigned to surgical pathology at KU will perform scheduled autopsies that day.

2. As most autopsies are shared, an autopsy sharing schedule is made at the beginning of the academic year by the Chief Residents, with priority given to more senior residents who may need case numbers.

**Autopsy Weekend Schedule:**

1. Autopsies are not performed on weekends unless there are exceptional circumstances approved by the Autopsy Director, in which case the resident on AP/CP weekend call will perform the case with the on-call faculty member.

Holidays

Holiday call coverage is scheduled by the Chief Residents at the beginning of each block and approved by the Residency Program Director. All holiday call will follow the ACGME guidelines. Occasionally, a single resident may provide holiday call coverage for AP, CP and autopsy services.

*Approved by REC, 1.22.2015*
OUTSIDE ROTATIONS AND MOONLIGHTING

Opportunities for education and work outside of the KUMC campus are available. In some instances, these are encouraged for the education of program residents. At other times, the resident receives remuneration for professional services rendered (moonlighting). All of these instances must be requested, approved and authorized by the administration of both the department and of KUMC. For these reasons, the following guidelines must be met:

Moonlighting

Pathology & Laboratory Medicine residents working for pay on off hours at institutions or physicians’ offices outside of KUMC.

- PGY1 residents are not eligible for moonlighting.
- All moonlighting must be approved, prior to the onset, by the Department Chair, Residency Program Director and the Executive Dean of the School of Medicine.
- The description of the moonlighting functions must be on record in the office of the Department Chair.
- Moonlighting is restricted to more senior residents functioning in the specialty of pathology in areas of resident competence.
- It is the resident’s professional responsibility to appear for regular duty hours rested and fit. The moonlighting experience must, in no way, compromise the educational time or function of the resident in the Pathology Residency Program.
- Moonlighting opportunities must occur during "off" hours. If the resident’s performance is compromised, the Residency Program Director and/or Department Chair can suspend the resident’s moonlighting privileges. (See the Graduate Medical Education Policy and Procedure Manual)
- Because KUMC and program teaching faculty have no direct role in the supervision of the professional activities of residents engaged in moonlighting, the state self-insurance program does not cover moonlighting activities. The resident must obtain his/her own individual professional liability policy.
- A moonlighting resident may have a level of coverage that is different from his residency coverage. Such insurance may be purchased by the resident or may be arranged by another individual or agency (e.g. the entity engaging the resident’s services). Regardless of the means of obtaining insurance, a certificate of insurance documenting the existence of an in-force policy must be provided to the resident and a copy filed with the program.
- The resident must have a permanent license to practice medicine in the state in which the moonlighting is to take place.
- The hours spent moonlighting will be counted towards the 80 hour work week limit.

For information on moonlighting, please refer to the GME Policy and Procedure Manual: http://www.kumc.edu/Documents/gme/Web%20Ready%20Version%205.4%20%2012.2015.pdf
Educational rotations outside of KUMC

Department residents involved in either elective, or required (other than KCVAMC) rotations outside of KUMC.

- All such rotations must be approved, prior to their onset, by the Department Chair, Residency Program Director, the Executive Dean and the Executive Vice Chancellor.
- Approval must be obtained using a special form available from the Residency Program Director or Chair. This form must be submitted a minimum of three months in advance of the start of the rotation.
- A description of this experience must be on record in the residency program curriculum book in the Chair’s office.
- Such programs, whether elective or required, should be beneficial to the education of the resident.
- Terms of agreement include designation of the outside facility (university, hospital, etc.), effective period of the rotation, fiscal considerations, licensure, malpractice coverage, supervision and evaluation of the resident's performance.
- Licensure: The state and license number must be added to the rotation application.
- Malpractice insurance: Professional liability insurance coverage is provided by the University's self-insurance program.
- Supervision and evaluation. The resident must provide evidence of supervision, that the supervising staff agrees to be responsible for the supervision of the resident in all patient care and that an evaluation of the resident's performance be forwarded to the Residency Program Coordinator upon completion of the rotation.
RESIDENT FATIGUE

All residents must complete the required Fatigue Training during the institutional orientation at the beginning of each residency. All faculty members are also educated to recognize the signs of fatigue and sleep deprivation and must adopt and apply the following institutional policy to prevent and counteract its potential negative effects on patient care and learning.

Purpose

Symptoms of fatigue and/or stress are normal and expected to occur periodically in the resident population, just as it would in other professional settings. Not unexpectedly, residents may on occasion, experience some effects of inadequate sleep and/or stress. As an institution, the University of Kansas School of Medicine has adopted the following policy to address resident fatigue and/or stress.

Recognition of Resident Excess Fatigue and/or Stress

Signs and symptoms of resident fatigue and/or stress may include but are not limited to the following:

- Inattentiveness to details.
- Forgetfulness.
- Emotional instability.
- Mood swings.
- Increased conflicts with others.
- Lack or attention to proper attire or hygiene.
- Difficulty with novel tasks and multitasking.
- Awareness is impaired (fall back on rote memory).
- Lack of insight into impairment.

Response

The demonstration of resident excess fatigue and/or stress may occur in patient care settings or in non-patient care settings such as lectures and conferences. In patient care settings, patient safety, as well as the personal safety and well-being of the resident mandates implementation of an immediate and a proper response sequence. In non-patient care settings, responses may vary depending on the severity of, and the demeanor of the resident’s appearance and perceived condition. The following is intended as a general guideline for those recognizing or observing excessive resident fatigue and/or stress in either setting.

EXCESSIVE RESIDENT FATIGUE OR STRESS

Attending Faculty Responsibilities:

1. In the interest of patient and resident safety, the recognition that a resident is demonstrating evidence for excess fatigue and/or stress requires the attending faculty or supervising resident to consider immediate release of the resident from any further patient care responsibilities at the time of recognition.
2. The attending faculty or supervising resident should privately discuss his/her opinion with the resident, attempt to identify the reason for excess fatigue and/or stress, and estimate the amount of rest that will be required to alleviate the situation.

3. The attending faculty must attempt, in all circumstances without exception, to notify the Chief/supervising resident on-call, Residency Program Director and/or Department Chair, respectively, depending on the ability to contact these individuals, of the decision to release the resident from further patient care responsibilities at that time.

4. If excess fatigue is the issue, the attending faculty must advise the resident to rest for a period that is adequate to relieve the fatigue before operating a motorized vehicle. This may mean that the resident should first go to the on-call room for a sleep interval lasting no less than 30 minutes. The resident may also be advised to consider calling someone to provide transportation home.

5. If stress is the issue, the attending faculty, upon privately counseling the resident may opt to take immediate action to alleviate the stress. If, in the opinion of the attending faculty, the resident stress has the potential to negatively affect patient safety, the attending faculty must immediately release the resident from further patient care responsibilities at that time. In the event of a decision to release the resident from further patient care activity; notification of program and administrative personnel shall include the Chief/supervising resident on-call, the Residency Program Director and Department Chair, respectively, depending on the ability to contact these individuals.

6. A resident who has been released from further immediate patient care because of excess fatigue and/or stress cannot appeal the decision to the responding attending faculty.

7. A resident who has been released from patient care cannot resume patient care duties without permission of the Residency Program Director or Department Chair, when applicable.

Resident Responsibilities:

1. Residents who perceive that they are manifesting excess fatigue and/or stress have the professional responsibility to immediately notify the attending faculty, the Chief Resident, and/or the Residency Program Director without fear of reprisal.

2. Residents recognizing resident fatigue and/or stress in fellow residents should report their observations and concerns immediately to the attending faculty, the Chief Resident, and/or the Residency Program Director.

Residency Program Director Responsibilities:

1. Following removal of a resident from duty, in association with the Chief Resident, the Residency Program Director will determine the need for an immediate adjustment in duty assignments for remaining residents in the program.

2. Subsequently, the Residency Program Director will review the resident's call schedules, work hours, extent of patient care responsibilities, any known personal problems, and stresses contributing to this for the resident.

3. The Residency Program Director will notify the Department Chair and/or program director of the rotation in question to discuss methods to reduce resident fatigue.
4. In matters of resident stress, the Residency Program Director will meet with the resident personally as soon as can be arranged. If counseling by the Residency Program Director is judged to be insufficient, the Residency Program Director will refer the resident to the following possible services depending on the severity of the issue through contact with the GME Office (913) 588-7293.

5. If the problem is recurrent or not resolved in a timely and satisfactory manner according to program leadership and the GME office, the Residency Program Director has the authority to release the resident from patient care and educational duties pending evaluation according to the leave and probation terms as stated in the KUMC Graduate Medical Education Policy and Procedure Manual, Section 11.

6. The Residency Program Director will release the resident to resume patient care duties only after the resident has demonstrated no further impairment with fatigue or stress issues.

7. Training must be made up to meet RRC training guidelines.

Fatigue in Non-Patient Care Settings

If residents are observed to show signs of fatigue and/or stress in non-patient care settings, the Residency Program Director should follow the Residency Program Director procedure outlined above for the patient care setting.

In cases where the resident feels too fatigued to drive home safely following a nighttime on-call assignment, two options are available. A swing room is available for sleeping and a voucher system is available for taxi transportation home and back to work the following day (Residency/Fellowship Coordinator has vouchers).

RESOURCES:

- KUMC’s Student Counseling and Educational Support (913) 588-6580 offers psychological and education services at no cost to students, residents, and fellows: http://www.kumc.edu/student-services/counseling-and-educational-support-services.html.

- KUMC’s Department of Psychiatry and Behavioral Sciences (913) 588-1300 offers a full range of inpatient, outpatient, and emergency services for the diagnosis and treatment of personal problems: http://www.kumc.edu/school-of-medicine/psychiatry-and-behavioral-sciences/services-and-info.html.

- Kansas Department of Health and Environment’s Employee Assistance Program (EAP) provides information, short-term counseling, advice, and referrals from licensed professionals (888) 275-1205 (option 7) http://www.kdheks.gov/hcf/healthquest/eap.html.

- Lawrence campus: University of Kansas Counseling and Psychological Services (785) 864-CAPS (2277) https://caps.ku.edu/ or Psychological Clinic Counseling (785) 864-4121 http://psychclinic.ku.edu/.
RESIDENT LEAVE

For information on resident leave, please refer to the GME Policy and Procedure Manual: http://www.kumc.edu/Documents/gme/Web%20Ready%20Version%205.4%20%2012.2015.pdf

Generally, residents have the following types of leave available:

- Vacation.
- Sick Leave.
- Professional Leave.
- Family Medical Leave Act (FMLA) Leave.

Only a total of five working days of any type of leave may be taken in any given four-week rotation. Additional time off will have to be made up during elective time. Until the excess time off has been made up, the resident will not receive credit for that rotation. Residents must inform the attending on service of any type of leave being taken during the rotation and ensure that it is acceptable to the service.

All leave (with the exception of unanticipated sick leave) must be requested and approved in MedHub at least two weeks prior to the start of the rotation. Residents are responsible for arranging coverage during their leave if it overlaps with previously assigned service, conference or call duties, and notifying the Chief Residents and administrative staff of all changes.

When leaving town for any reason, residents must leave their complete, temporary address (inclusive of telephone and email) with the Residency/Fellowship Coordinator, kates2@kumc.edu.

The American Board of Pathology requires an average of 48 weeks of full-time service per year over a four-year period to be eligible for the AP/CP certification exam. Residents who cannot meet this standard will be required to extend their residency training.

Vacation

Each resident is entitled to fifteen days of vacation annually. Vacation blocks are scheduled by the Chief Residents prior to the start of the academic year. Residents may request changes to the assigned vacation blocks, if needed. However, any changes in the vacation schedule after the start of the year must be approved by the Chief Residents and the resident will be responsible for arranging service and/or call coverage as necessary.

In general, vacations are approved in a block of five days during the rotation, and must be approved by the attending staff on the rotation. At the discretion of the attending staff on the rotation, vacation days may be taken individually instead of as a contiguous block. All vacation days must be requested and approved at least two weeks prior to the start of the rotation. All vacation requests must be entered in MedHub and will be automatically documented with duty hours.

Vacation time must be used in the fiscal year (July thru June) in which it is earned.
The following additional restrictions on vacation time apply:

Scheduling of vacation is restricted to certain rotations. Residents may NOT take vacation time while in the following rotations:
- KU Surgical Pathology/Autopsy.
- KU Surgical Pathology Trainer/Elective.
- KU Hematology.
- VA Surgical Pathology.
- Children’s Mercy Hospital Surgical Pathology.
- Children’s Mercy Hospital Molecular Pathology.
- Autopsy, Jackson County.
- Community Blood Center.

**Note:** No vacation is permitted for any resident from June 15 to July 15.

**Sick Leave**

Residents have up to 10 workdays of sick leave per year, covered by the resident’s stipend, to cover personal illness or illness in the resident’s immediate family (spouse, parents or children). At the discretion of the Residency Program Director, a physician’s written statement may be required as a condition of approval for sick leave. The program also may require a certification that the resident is released to return to work following three or more consecutive days of absence resulting from the resident’s own illness.

Unscheduled (sick leave) absences must be reported by telephone to the service attending, Chief Residents and the Residency/Fellowship Coordinator as early as possible on the day of absence. All leave must be requested and approved in MedHub and will be automatically documented with duty hours. Sick leave must be used in the fiscal year (July thru June) in which it is earned.

**Professional Leave**

A resident may be entitled to up to five days of professional leave annually, subject to approval by the Residency Program Director. Requests for professional leave should be made and approved through MedHub at least two weeks in advance of planned leave. Professional leave may be used for conference attendance, taking medical board examinations, or interviews. Unused professional leave may not be carried over from year to year. Professional leave days are considered as service time for ABP training requirements.

**FMLA Leave**

Residents who have worked a minimum of one year and 1250 hours during that year at KUMC are eligible for FMLA leave. FMLA leave is utilized when there is an expectation of an extended period of absence due to one of the reasons listed below. FMLA leave protects the residents’ position, ensures continuation of benefits and allows paid vacation and sick leave to be used without additional approval. It also allows for additional unpaid leave during which other benefits of employment continue, at the resident’s expense.

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*KUMC Pathology Residency Manual*
Residents who anticipate the need for an extended period of absence must meet with the Residency Program Director as early as possible to develop an acceptable leave plan. Residents should complete the FMLA Leave Checklist from the GME Policy and Procedure Manual prior to meeting with the Residency Program Director (see example below). FMLA leave is scheduled with assistance from KUMC’s Human Resources Department.

A resident eligible for FMLA leave may request FMLA designation pursuant to the University's FMLA policy for up to twelve weeks of leave in a continuous 12-month period:

1. Because of the resident’s own serious health condition, including pregnancy, or a qualifying illness or injury;
2. To care for the resident’s immediate family member who has a serious health condition;
3. For the birth of a child or placement of a child with the resident for adoption or foster care;
4. For any “qualifying exigency” arising out of the fact that the resident’s spouse, son or daughter (of any age) or parent is on active duty or call to active duty status in support of a contingency operation as a member of the National Guard or Reserves.
5. A resident eligible for FMLA leave also may request up to 26 weeks of military caregiver leave to care for a spouse, son, daughter, parent or next of kin who is a covered service member in the regular armed forces, the National Guard or Reserves and who is undergoing medical treatment, recuperation or therapy, or who is otherwise on the temporary disability retired list, for a serious injury or illness relating to that covered service member’s military service.

Residents must first use all sick leave and vacation time while on FMLA. If the maximum number of vacation and sick leave days for the year have been used, the resident's remaining FMLA leave will be unpaid. Therefore, if it is known that a period of extended absence will be necessary (e.g. pregnancy), sick leave and vacation time for that year should not be used earlier to avoid having unpaid time off. If it is known that a period of leave will be necessary, it is advised to minimize the use of sick and vacation time to avoid extended unpaid time off.

Extended absence may affect the American Board of Pathology's training requirements and the resident's progress through the program. The American Board of Pathology requires an average of 48 weeks of full-time service per year over a four-year period to be eligible for the AP/CP certification exam. Residents who cannot meet this standard will be required to extend their residency training.

For more information on KUMC’s FMLA program, see http://www.kumc.edu/human-resources/benefitsrewards/family-and-medical-leave-act.html.

Approved by REC, 2.26.2015
OTHER RESIDENT POLICIES

Medical Student Teaching Responsibilities

Members of the resident staff participate in the teaching program as junior instructors. Residents are involved with teaching medical student histopathology labs in years R2-4. This is a valuable part of the resident experience and most residents enjoy the association with students.

Teaching responsibilities also include:
- The performance of autopsies with medical students.
- Substituting for senior staff in small group problem-based learning sessions.

Occasionally, a resident may be asked to give a lecture, if they have developed a special area of expertise, or express a desire to lecture.

Pagers

The Department of Pathology & Laboratory Medicine will purchase a pager for each resident. If the pager is lost or damaged, the resident is responsible for the cost of the replacement.

Procedures and Logbooks

The ACGME requires Pathology residents to list the following procedures on the ACGME web-based logbook:
- Autopsies.
- Bone marrow aspirates/biopsies.
- Fine needle aspirates.

It is the responsibility of each resident to maintain updated ACGME logs.

Hospital and Departmental Services

Consult the Chief Residents or Residency/Fellowship Coordinator regarding uniforms, laundry, and necessary keys. Keys, protocols, slides, sections, and blocks must be obtained from and returned to the appropriate departmental offices. Assignment of individual offices, microscopes, and other equipment will be made by the Chief Residents. Full day attendance for off-site rotations (VA and Children’s Mercy Hospital) is required. Exceptions include returning to KUMC for mandatory conferences and meetings (see Didactic Sessions and Conferences, page 23). Any other absence must be approved by the VA or Children’s Mercy Hospital faculty.

Note: The Graduate Medical Education Policy and Procedure Manual represents the institutional guidelines, policies and procedures governing the selection, appointment, evaluation and promotion of residents at the University of Kansas School of Medicine. While every effort has been made to ensure the accuracy and comprehensiveness of the information presented, the content of this manual is subject to change. See the following: http://www.kumc.edu/Documents/gme/Web%20Ready%20Version%205.4%20%2012.2015.pdf.
### Core Competency Teaching and Assessment Matrix

<table>
<thead>
<tr>
<th>Core Competency</th>
<th>TEACHING METHODS</th>
<th>EVALUATION METHODS</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Didactic lecture</td>
<td>Report Review</td>
</tr>
<tr>
<td></td>
<td>Faculty Seminar</td>
<td>Direct Observation</td>
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<td></td>
<td>Journal Club</td>
<td>Checklist</td>
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<td></td>
<td>Direct Supervised</td>
<td>Global Rating</td>
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<td>proc</td>
<td>Practical Skills</td>
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<td></td>
<td>Lab Inhouse</td>
<td>Staff Skills Rating</td>
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<tr>
<td></td>
<td>Disc Conf</td>
<td>Portfolios</td>
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<td></td>
<td>Online Tutorials</td>
<td>In-house</td>
</tr>
<tr>
<td></td>
<td>Unknown cases/etns</td>
<td>Written Exam</td>
</tr>
<tr>
<td></td>
<td>Project</td>
<td>Procedures or Case</td>
</tr>
</tbody>
</table>

#### PATIENT CARE
- Demonstrate diagnostic competencies
- Provide effective consultation
- Demonstrate technical skills

#### MEDICAL KNOWLEDGE
- Knowledge of evolving biomedical/clinical science
- Demonstrates an application of this knowledge to patient care

#### PRACTICE-BASED LEARNING & IMPROVEMENT
- Analyze own practice for needed improvements
- Application of research & statistical methods
- Use of information technology
- Facilitate learning of others

#### INTERPERSONAL & COMMUNICATION SKILLS
- Effective in information exchange
- Works well within a team
- Listening skills

#### PROFESSIONALISM
- Respectful, altruistic
- Ethically sound practice
- Sensitive to cultural, age, gender, disability issues

#### SYSTEMS-BASED PRACTICE
- Understand interaction of their practices with the larger system
- Knowledge of practice and delivery systems
- Practice cost-effective care
- Advocate for patients within the health system

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KUMC Pathology Residency Manual
SURGICAL PATHOLOGY

FACULTY

Ossama Tawfik, MD, PhD  Professor and Director – Surgical Pathology
Ivan Damjanov, MD, PhD  Professor
Fang Fan, MD, PhD  Professor
Wei Cui, MD  Associate Professor
Garth Fraga, MD  Associate Professor
Rashna Madan, MBBS  Associate Professor
Kathy Newell, MD  Associate Professor
Janet Woodroof, MD  Associate Professor
Da Zhang, MD, MSc  Associate Professor
Katie Dennis, MD  Assistant Professor
Elizabeth Friedman, MD  Assistant Professor

GOALS AND OBJECTIVES

The ultimate goal of the rotation in surgical pathology is that the resident becomes competent in the interpretation of surgical pathology material and to learn communicative and consultative skills that will aid the clinicians in the correct diagnosis and treatment of the patients. Goals are based on Skill Levels I and II.

Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Activity</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
<td>DL</td>
</tr>
<tr>
<td>Faculty sign-out</td>
<td>FSO</td>
</tr>
<tr>
<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
</tr>
<tr>
<td>Role modeling</td>
<td>RM</td>
</tr>
<tr>
<td>Lab inspections</td>
<td>LI</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
<td>IC</td>
</tr>
<tr>
<td>Online tools</td>
<td>OT</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
<td>USC</td>
</tr>
<tr>
<td>Project</td>
<td>P</td>
</tr>
</tbody>
</table>

Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Method</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
<td>RR</td>
</tr>
<tr>
<td>Direct observation</td>
<td>DO</td>
</tr>
<tr>
<td>Checklist</td>
<td>CL</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
</tr>
<tr>
<td>Standardized exam</td>
<td>SE</td>
</tr>
<tr>
<td>Practical slide exam</td>
<td>PSE</td>
</tr>
<tr>
<td>In-house written exam</td>
<td>IWE</td>
</tr>
<tr>
<td>360 multisource rating</td>
<td>360</td>
</tr>
<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
</tr>
</tbody>
</table>
## SURGICAL PATHOLOGY CORE COMPETENCY: PATIENT CARE

**Goal:**
*Skill Level I:* Residents must demonstrate competence in processing of basic types of patient specimens and evaluation of clinicopathological aspects of Surgical Pathology.

*Skill Level II:* Residents must demonstrate competence in processing all types of patient specimens, performing and interpreting frozen sections, utilizing ancillary studies, and generating clinically relevant diagnostic reports.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate competence in basic specimen processing skills (I).</td>
<td>DL, FSO, DSP</td>
<td>RR, DO, CL, GR/FE, SE, IWE, 360, PF</td>
</tr>
<tr>
<td>Demonstrate competence in selecting representative tissue samples for intraoperative frozen sections, preparing the same, and staining the sections. (I)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate proficiency in interpreting and reporting frozen sections within 20 minutes of receiving a specimen for that purpose in the pathology laboratory. (II)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the techniques for preparing intraoperative cytology smears. (II)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Be able to independently report the histopathologic aspects of routine and complex cases, including cases prepared by junior residents and/or pathology assistants with attention to organization of diagnostic format, development of differential diagnosis, and ordering of necessary special stains and other ancillary techniques. (II)</td>
<td>DL, FSO, DSP, RM, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE, PF</td>
</tr>
<tr>
<td>Demonstrate proficiency in digital imaging techniques. (II)</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

## SURGICAL PATHOLOGY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Skill Level I:* Residents must be able to evaluate normal histology and basic pathologic processes.

*Skill Level II:* Residents must demonstrate competence in histopathological diagnosis, grading and staging of tumors, and interpretation of ancillary studies.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Determine when a microscopic description and/or interpretation is necessary, and provide such information. (I)</td>
<td>FSO, RM, USC</td>
<td>RR, DO, GR/FE, PF</td>
</tr>
</tbody>
</table>
Be able to evaluate margins of tumor resection specimens using frozen sections and touch preparations. (I)  
Demonstrate knowledge of the common situations requiring expedited processing of a pathology specimen, and those that do not. (II)  
Demonstrate knowledge of the common indications for an intraoperative consultation. (II)  
Enumerate the indications and the limitations pertaining to intraoperative frozen section examinations. (II)  
Demonstrate knowledge of the common grading and staging systems applied to malignant neoplasms. (II)

**SURGICAL PATHOLOGY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**  
*Skill Level I:* Residents must be familiar with laboratory workflow, Surgical Pathology section of the Resident Manual, and laboratory safety practices.  
*Skill Level II:* Residents must be consistently fast and competent at signing out cases, including ordering of deeper sections, recuts, special stains, immunostains, generation of microscopic descriptions, and notification of clinicians.

**Objectives:**

<table>
<thead>
<tr>
<th>Activity</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to properly assign CPT codes to all specimens in anatomic pathology. (I)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Submit appropriate forms for billing pertaining to specimens. (I)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO</td>
</tr>
<tr>
<td>Know the procedures for the reporting of untoward incidents in the laboratory. (I)</td>
<td>DSP, RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of how and when to obtain external consultations in anatomic pathology and document the results appropriately. (II)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate an ability to manage workflow in the gross room, assist junior residents with gross dissection, provide accurate gross descriptions of routine and complex specimens, use the local anatomic pathology laboratory information system, and practice safety in the pathology laboratory. (II)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, 360, PF</td>
</tr>
<tr>
<td>Demonstrate knowledge of available procedures for locating a missing specimen and resolving questions of specimen identity. (II)</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Demonstrate knowledge of quality control pertaining to histologic sections and special stains, including trouble-shooting of mistakes in accessioning, labeling, & misidentification of specimens. (II) | FSO, DSP, RM | DO, GR/FE

Review consultation slides on referral cases with attention to pertinent clinical information, requests for additional slides or blocks if needed, and formatting of the final consultative report. (II) | FSO, DSP, RM | RR, DO, GR/FE

**SURGICAL PATHOLOGY CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS**

**Goal:**

*Skill Level I:* Residents must be familiar with the elements of a Surgical Pathology report and the information it conveys.

*Skill Level II:* Residents must be competent in constructing a comprehensive Surgical Pathology report and in written and verbal communication with other healthcare providers.

**Objectives:**

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
</table>
| Demonstrate knowledge of the common and basic elements of the surgical pathology report, including: (I)  
- Identifiers (patient and institution).  
- Input from the responsible pathologist.  
- Input from the responsible clinician.  
- Necessary dates and times that must be in the report.  
- Necessary clinical information.  
- Documentation of the specimens that were submitted.  
- Thorough and accurate gross description. | FSO, DSP, RM | RR, DO, GR/FE |
| Demonstrate the ability to effectively construct a complex surgical pathology report. (II) | FSO, DSP, RM | RR, DO, CL, GR/FE, PF |
| Be able to properly prepare synoptic surgical pathology reports for common malignancies. (II) | FSO, DSP, RM | RR, DO, CL, GR/FE, PF |
| Demonstrate the ability to dictate necessary amendments and/or addenda for surgical pathology reports. (II) | FSO, DSP, RM | RR, DO, GR/FE |
| Demonstrate the steps for preparation of consultation reports on outside slides and/or paraffin blocks, and transmittal of those reports to responsible clinicians and/or referring pathologists. (II) | FSO, DSP, RM | RR, DO, GR/FE |
### SURGICAL PATHOLOGY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Skill Levels I and II: Residents must demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to diverse populations within the healthcare environment.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>

### SURGICAL PATHOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Skill Levels I and II: Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Show a working knowledge of the basic principles of quality assurance, quality control, continuous quality improvement, and outcomes analysis, as they apply to anatomic pathology.</td>
<td>FSO, RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the standards (TJC, CAP) required for submitting surgical pathology specimens.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the basic recommendations/requirements (TJC, CAP, regional legal requirements) pertaining to retention of pathology specimens and records.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Task</td>
<td>Responsible Parties</td>
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<td>----------------------------------------------------------------------</td>
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<tr>
<td>Demonstrate knowledge of the basic principles of informatics in anatomic pathology, and ability to effectively utilize the local computer network.</td>
<td>RM, OT</td>
<td></td>
</tr>
<tr>
<td>Understand federal and state regulations with special application to anatomic pathology, and the general requirements for compliance in the reporting of professional activities for billing purposes.</td>
<td>FSO, RM, LI, OT</td>
<td></td>
</tr>
<tr>
<td>Demonstrate knowledge of web-based or organizations (CAP, ASCP, USCAP, etc.)-related learning and CME tools in anatomic pathology.</td>
<td>RM, OT</td>
<td></td>
</tr>
<tr>
<td>Demonstrate a familiarity with standards set forth by the CAP and TJC for laboratory certification in anatomic pathology, and participate in at least one internal (“mock”) inspection of the institutional anatomic pathology laboratory.</td>
<td>RM, LI, OT</td>
<td></td>
</tr>
<tr>
<td>Understand the principles applying to evaluation of the cost-effectiveness of laboratory procedures in anatomic pathology.</td>
<td>FSO, RM</td>
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</tr>
<tr>
<td>Demonstrate an ability to organize, perform, and analyze a quality control review project in surgical pathology for presentation to faculty.</td>
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<tr>
<td>Demonstrate knowledge of how to utilize risk-management resources in cases involving medicolegal liability.</td>
<td>DL, RM</td>
<td></td>
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<tr>
<td>Understand the basic legal aspects of medical malpractice lawsuits, and the potential roles of pathologists as defendants and consultants in such actions.</td>
<td>DL, RM</td>
<td></td>
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</tbody>
</table>
PGY SURGICAL PATHOLOGY-SPECIFIC GOALS

PGY 1 GOALS: By the end of the first year:

- The resident has documented direct supervision of grossing the first three specimens in the majority of specimens on the organ-based list of specimens.
- The resident can dictate informative gross dictations with proper cassette summaries and cut appropriate sections without direct supervision on all biopsies and simple routine specimens and most common cancer cases.
- The resident becomes efficient in managing cases, such that an appropriate around time is observed.
- The resident can independently perform frozen section procedures and can report frozen section results following joint faculty/resident interpretation of frozen sections.

PGY 2 GOALS: By the end of the second year:

- The resident has documented direct supervision of grossing the first three specimens for all specimens on the organ-based list of specimens.
- The resident demonstrates the ability to work up cases properly, including ordering appropriate histochemical and immunohistochemical stains.
- The resident demonstrates efficiency and professionalism in the handling of cases (turnaround time is kept to 48 hours, special stains, immunostains are ordered when the attending staff requests them, the resident does the follow-up on stains when they do not arrive when anticipated).
- The resident demonstrates an economy of sections that are adequate to provide all the necessary information, and minimizes the need to submit additional wet tissue.
- The resident demonstrates the ability to communicate appropriately to clinical colleagues, including impromptu drop-by visits and in CPC-type conferences.
- The resident is ready to start supervising (in the third year) junior residents in surgical pathology procedures.

PGY 3 GOALS: By the end of the third year:

- The resident must be able to compose a gross and microscopic surgical pathology report which is ready for electronic signature, with minimal, if any, correction.
- The resident should be comfortable performing independent intraoperative consultations.

PGY 4 GOALS:

- The resident should be able to supervise junior residents in all aspects of the practice of surgical pathology.
- The resident should have demonstrated increased medical knowledge through performance on study set examinations, conferences and conducting clinical conferences.
• The resident is practice ready for billing, Medicare compliance and accreditation issues.
• The resident should have completed subspecialty related rotations including dermatopathology and pediatric pathology at CMH.
• The resident must document minimally 200 intraoperative consultations/frozen sections.
• The resident must have reviewed minimally 2,000 surgical pathology cases that they have reviewed and signed out.
• The resident should be competent in all surgical pathology Skill Levels 1 and 2.

GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice--Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning found on page 20-21.

GENERAL ORGANIZATION

Approximately 30,000 cases are accessioned annually in the Division of Surgical Pathology, which primarily deals with tissues obtained while patients are in the University of Kansas Hospital, or are to be admitted to the hospital. Whether tissues have been excised for diagnosis or for therapy, surgical pathology is of direct relevance to both patients and treating physicians. The proper histopathologic documentation of the lesion constitutes an essential element in the work-up of the patient. The subsequent management of the patient is greatly influenced by the opinion expressed by the surgical pathologist.

The required time for training in surgical pathology is 12 blocks. Senior residents also devote a ½ block as junior resident trainers. This rotation is divided into four-week blocks but residents can be scheduled for several consecutive segments. Since this rotation requires resident (or fellow) coverage at all times, all absences must be excused by the Director of Anatomic Pathology and the Residency Program Director. It also requires pre-notification of the pertinent attending pathologists and arrangement of resident/fellow cross-coverage, if necessary. Punctuality and good attendance are critical in surgical pathology and reflect the professionalism and seriousness of the resident on this rotation.

Residents who are assigned to the rotation are expected to take an active role with the senior staff in completing the daily cases in a reasonable time. The residents are not only learning surgical pathology “on the job” but they are practicing physicians and members of a team collaborating closely with other physicians, often during performance of operations. This requires on the part of the residents a considerable degree of alertness, thoroughness, neatness, attention to details, as well as an understanding of clinical and surgical problems.

Therefore, the residents should have the sense of urgency in completing their daily assigned work with the senior staff members. They are also in an enviable position to utilize some of the material available to them in sophisticated studies involving tissue culture, immunology, electron microscopy, fluorescent microscopy, image analysis, nuclear morphometry, flow cytometry, chemical analysis, histochemistry, immunohistochemistry and molecular biology.
SURGICAL PATHOLOGY SCHEDULING

DAY 1 Resident:

- Gross all big and small specimens all day (PA/techs grosses biopsies and assists with placentas if needed).
- Perform all frozen sections (with assistance from PA and faculty) and the big specimens following the frozen sections (whether they come the same day or the next day).
- Follow up on biopsies from Day 4.
- New cases accessioned after 4:30 p.m. are assigned to the resident who is Day 1 on the next day. (PA does biopsies and placentas).
- On-call for afterhours frozen section specimens and STAT cytology specimens until 9:30 p.m.

DAY 2 Resident:

- Preview slides for big specimens for sign-out out and order all necessary special stains after consulting the assigned faculty.
- Sign-out big specimens.
- Finish cutting in additional specimen parts that are from cases logged during Day 1.
- Cover autopsy service.

DAY 3 Resident:

- Preview and sign-out any remaining big specimens.

DAY 4 Resident:

- Sign-out biopsy specimens with biopsy attending after reviewing the slides.
- Work-up and sign-out all remaining cases.
- Prepare for Day 1 by collecting appropriate patient history from O2 and pertinent previous material on planned surgeries for Day 1, and studying the gross manual for each type of specimen anticipated.

Junior Residents Responsibilities

- Frozen Section.
- Review operating room schedule before frozen section duty with senior resident/fellow and/or attending staff.
- Perform, under supervision, frozen sections, including description, cutting sections, staining and microscopic study.
- Arrange for all special procedures, saving tissue for special studies (e.g. electron microscopy, flow cytometry, microbiology, touch imprints, photography, immunohistochemistry, etc.).
• Complete all paperwork.
• Keep the area clean and ready for work.
• Grossing Specimens.
  o Read section on surgical pathology in the Residents’ Manual.
  o Read gross manual in Rosai’s *Textbook on Surgical Pathology.*
  o Learn how to use CoPath (computer application for reporting of pathology results in anatomic and
    some areas of clinical pathology) for specimen accession and gross dictation.
  o Call attending, senior resident/fellow, or pathology assistant for help and advice whenever needed.
  o Work smoothly with histotechnologists, pathology assistants and transcriptionists.
• Microscopic Sign-out.
• Collate and order all slides and paperwork.
• Obtain appropriate clinical history prior to sign-out.
• Write tentative diagnoses before review with the attending.
• Sign-out rush or STAT cases promptly with the attending.
• Cooperate with the staff to sign-out the cases as soon as possible.
• Notify clinicians with the diagnoses, if necessary.
• Return slides and paperwork promptly.
• Learn fundamentals and specifics of tissue codes and billing.
• Order and interpret special procedures when necessary, e.g., special stains, EM, immunohistochemistry.
• Evaluate controls for special stains and immunohistochemistry and understand the reasons for the
  appearance of commonly used special stains.
• Learn how to use Voicebrook system (speech to text transcription software).

Miscellaneous Duties

• Learn how to use the computer information systems (CoPath, SMS, etc.) to check for previous reports on a
  patient, and status/results of specimens in cytopathology and clinical pathology, radiology, etc.
• Learn how to take gross photography.
• Learn how to log cases for surgical pathology teaching files.
• Prepare for interdepartmental conferences.
• Attend appropriate conferences.

Senior Residents Responsibilities (the above plus…)

• Write succinct diagnoses on all cases before review with the attending.
• Write microscopic descriptions, comments, etc., as needed.
• Review outside slides with attending and write the reports.
• Review relevant previous surgical and cytological material as needed for comparison.
• Help in writing letters to outside consultants (if needed) to review difficult cases.
• Assist medical students and junior residents.
RECOMMENDED READING LIST:

SURGICAL PATHOLOGY RESIDENT DUTIES AND RESPONSIBILITIES

Division of Specimens

- The resident will be responsible for a minimum of one-third of the "big" specimens a day (approximately 3-6). The number depends on the resident experience level and the volume for the day. Big specimens include Whipple, hysterectomy for cancer, breast specimens, lung resections, colon resections, etc. The cases will be divided up the previous day when the OR schedule comes out and will be assigned by the surgical pathology fellow and the senior PA. If there is disagreement about the distribution of specimens, the final decision will be made by Dr. Katie Dennis, Residency Program Director.
  - **NOTE:** It is expected that the junior residents prepare for the specimens they receive (for example, read the grossing manual and become familiar with the staging for that tumor the night before).
- The remaining two-thirds of the big cases will be grossed by the PAs and PA students. When there is no PA student, the number of specimens that the resident is responsible for will increase. Also, if the PA is gone (sick, vacation, etc.) the resident responsibility will increase.
- The gross room staff will gross the subsequent parts of big cases that come late (e.g. lymph node dissections, staging biopsies, etc.) - the majority of which are accessioned the next day.
- The gross room staff (the PAs, the surgical pathology technicians, and the student PA) will gross the benign routine specimens after 1st year residents demonstrate competency (three specimens each part type).
- If the resident is done grossing their assigned specimens and there is no frozen section going on or leftover slides to sign-out with an attending, the resident will help with other bigs, additional parts and benign specimens.
- If the PA is not performing administrative duties and there is work to be done in the gross room, the PA will help with other bigs, additional parts, and/or benign specimens.
- The resident is expected to participate in the interpretation of frozen sections.
- As the resident becomes more experienced, they will transition into a more supervisory role by participating in frozen section selection.
- In blocks where there are residents on elective rotations, one relevant “big” specimen will be assigned to the elective resident per day (e.g., a mastectomy for the breast elective resident, a laryngectomy for the ENT elective resident, etc.).

Specimen Identification

- Proper identification of the specimen is of prime importance.
- Double check labels and requisition sheet to be certain the specimen and patient identification correspond. If there is doubt, call the clinician to identify the specimen before processing.
- Extreme care in labeling cassettes and meticulous work sheet records are essential to prevent serious mix-ups. **Mislabeling is the most serious problem in dealing with small specimens.**
Continuously monitor slide labels and be certain that the slides belong to the case. Records should help the resident determine how many slides and of what structure should exist. Mislabeling can occur in the Histology laboratory; if there is the slightest doubt, check labels and blocks.

If no specimen can be found in a container, call the requesting physician and staff before discarding.

Clinical Data

Although the pre-operative and post-operative diagnosis and pertinent history are required on all specimens submitted, these are often lacking. If there is any doubt about the nature of the specimen or what information is needed by the clinicians, clarify these matters before processing. Some complicated specimens need to be oriented by the surgeon.

Before the resident is called to the OR for frozen sections as well as before the case is signed-out, be aware of all pertinent data. This may require inquiry of the clinicians, review of chart, looking at x-rays (get these for review by attending when appropriate, e.g. bone tumors) pulling previous slides and reports. **Have all pertinent slides and reports on previous specimens ready at time of sign-out.**

Specimen Processing

Processing involves more than gross dictation and submitted fixed tissue blocks. The best time for collecting tissue for special studies is the fresh specimen and for some studies this is the only time.

Except for tissue culture and culture for microorganisms, absolute sterility is not requisite. If these studies are needed, use sterile instruments and gloves.

**Note: It is usually better for the surgeon to culture tissue for organisms in the Operating Room where sterile conditions prevail.** All cultures should be obtained in the OR.

As soon as the specimen is received, consider the need for the following, some of which must be done on fresh tissue.

- **Cultures.** Pathology will only obtain cultures if the surgeon has sent the specimen fresh and no other tissue is present in the OR. Any culture obtained in pathology is not ideal and this should be understood by the surgeon. Because of sterility requirements, get appropriate equipment before handling tissue. (Sterile, fresh tissue required).
- **Cytogenetics studies.** Fresh sterile tissue is required in some cases.
- **Extreme Drug Resistance Assay (Oncotech Studies).** Fresh sterile tissue is required.
- **Molecular Studies.** Many tumors require additional molecular testing. Residents should familiarize themselves with what tests are ordered on what tumor and what specimen and learn how to order such tests in CoPath. All are supposed to consult with their staff about ordering such tests to take care of patients appropriately.
- **Cell surface marker studies.** Fresh or fixed tissue acceptable.
- **Immunofluorescence.** Fresh tissue required.
• Analysis for hormones or other chemical studies. Fresh frozen tissue may be required.
• Photography. If in doubt, take a photograph before dissection. See Photography section below.
• X-ray. The Faxitron is particularly useful for identification of calcification. Better done before any dissection.
• Electron microscopy. Best done on very fresh tissue fixed immediately in glutaraldehyde.

The handling of specimens requires individual judgment in each case. In some instances, it is necessary or desirable to open or dissect the specimen immediately. In other circumstances, fixation before opening is preferable.

Gross Examination Only

With most specimens, microscopic examination is necessary to establish a diagnosis or provide adequate documentation. The microscopic slides and tissue blocks form a valuable, permanent record for patient care and investigation. However, microscopic study is likely to provide little information of value beyond what can be seen grossly in certain types of specimens. In these instances, adequate documentation can be provided by gross description. The following specimens may be examined grossly without microscopic study unless some unusual feature is present that warrants histologic examination:

• Foreign bodies (tissue around foreign bodies should be examined).
• Nasal cartilage for septal deviations.
• Cataracts.
• Products of Conception (POC) for therapeutic abortions unless tagged by clinical staff. All spontaneous abortions should be processed.

Note: Occasionally a clinician requests "Gross Only," for economy to a patient. Such requests may be considered within the guidelines noted above. The decision of whether sections should be submitted to establish a diagnosis rests with the pathologist.

Fixation

Good fixation is essential for good histology. The most useful general fixative is formalin, and the most rapid and effect fixation is accomplished by use of very thin tissue section. Other fixatives than formalin are useful, even necessary in many situations.

• Formalin. A buffered aqueous solution of formaldehyde. This fixative is relatively inexpensive, it penetrates tissues well, artifacts are minimal, and the tissue can remain in the fixative indefinitely. Nearly all special stains and immunoperoxidase studies can be done on Formalin fixed tissue. Formalin penetrates slowly; hence the need for thin section for rapid overnight processing usually required for surgical pathology. Formalin is the only fixative than can be used for long term storage. Use Formalin for any specimen for which no special fixative is prescribed. (See individual sites for techniques of fixative with Formalin.)

• Plus Fix™. B-Plus contains formaldehyde and zinc salts and other buffers. No dilution of the solution is necessary. This fixative is rapid and provides superior nuclear detail. B-Plus is used for lymph node protocol cases (not node dissections), and other specimens to be studied for lymphoid and hematopoietic
disorders. B-Plus is also good for testis biopsies, testicular neoplasms, and thymic lesions among others. Bone marrow biopsies should be fixed for two hours and other larger tissue like lymph nodes should be fixed at least three to four hours before processing. Bone Marrow tissue must be submitted in a white cassette. No rinsing of the blocks is needed after fixation. Note: Timing is critical, and tissues must be extremely thin. See Lymph Node for technique.

- **Zenker's.** In addition to mercuric chloride, Zenker's also contains potassium dichromate which gives the orange color. The only specific use for this fixative is to elicit the chromaffin reaction in pheochromocytomas and related lesions. Use same cautions as B-Plus.

- **Bouin's.** This fixative is based on picric acid and has a yellow color. It is excellent for renal and testicular biopsies and for certain special stain techniques especially granules of islet cell tumors and carcinoids. Since immunoperoxidase is now more specific and since B-plus can be substituted for testis, there is little need for Bouin's.

- **Hartmann's.** The addition of acetic acid to formalin increases penetration. Useful for dense specimens especially uterus to improve penetration of fixative, but it makes tissues "brittle" and lyses blood elements. In addition, Hartmann's fixative destroys eosinophils and severely compromises electron microscopic analysis.

- **Glutaraldehyde.** Used for electron microscopy, this solution is also an excellent general purpose fixative but is too expensive to use for this purpose routinely. Penetration is rather weak and tissue should be thin. For electron microscopy 1 mm cubes are used. (See special instructions for EM).

- **Absolute Alcohol.** Used for immunohistochemical stains for sarcoma and gout. These specimens need to be processed separately. Talk with the histologist.

**Dissection**

- Dissection may need to be individualized to demonstrate the salient features. Refer to specific organs for the recommended techniques, but be prepared to modify these to meet individual circumstances.

- Weights and measurements should be taken before cutting into a specimen, since the fluid contents may escape.

- Gentle handling of tissue is also important to avoid destructive artifacts.

- Avoid washing fresh tissue with tap water. Rinse bowel mucosa gently with formalin or saline.

- Use a sharp knife for big specimens, a sharp razor blade for small ones. **Slice;** don't chop. Change blades frequently. Scissors are useful for opening hollow viscera and removing sutures, but generally should be avoided for other cutting.

- **Remove all sutures and clips from tissue.** These can devastate microtome knives.

- Keep tissues wet at all times. Cover cut surfaces with a wet paper towel. **Put sections into formalin quickly.**

- Do not allow sections to dry by leaving tissues or cassettes out of fixative.

- Clean instruments well after each specimen and at the end of the day.

- Clean cutting station at the end of each day.
Decalcification

- Tissues needing decalcification must first be thoroughly fixed.
- Small specimens may be sent to the Histology laboratory in cassettes. The worksheet must be labeled "DECAL," and the specimens separated from those put onto the processing machines. The cassettes should be labeled "Decal" on the side.
- Large specimens are better handled by the prosector, who may need to partially decalcify a bulky specimen, then take sections for histology and finally complete the decalcifying process.
- It is the responsibility of the resident to check and trim decal specimens daily.

Photography

Photographs documents specimens and are an invaluable resource for teaching and research. A photograph must be properly exposed, but more importantly, correctly composed.

The following guides are useful in specimen photography, but before taking pictures, check the planned composition with an attending pathologist or the surgical pathology fellow.
- All specimens should be labeled, the label typed and with a metric ruler. The label and ruler should be at the edge of the picture, never on top of the tissue or in the middle of the picture.
- The picture should fill the frame as much as possible, since background is uninteresting.
- Background must be clean. Use the glass whenever possible (rarely a specimen is too big or too small). The color should be neutral, light blue for dark specimens, black for others. A black background is easily obtained by placing black plastic or paper underneath the glass the specimen is on. This is known as darkfield illumination.
- Avoid reflection from overhead lights (turn them off) and highlights (adjust light source).

Photo composition hints:
- Use anatomic orientation whenever possible.
- Show relationship of the lesion to normal tissue.
- The cut surface of a tumor is nearly always more useful than the outside, unless the outside shows relationships to normal structures.
- Avoid instruments, fingers, etc., in the picture (e.g., use a glass slide to hold larynx open).
- Papillary tumors often are best demonstrated when taken underwater.
- Partial (or complete) fixation may be useful in demonstrating some lesions.
- A few minutes in 70% alcohol helps restore contrast and color in fixed specimens.
- Digital photographs are archived on the hospital computer system in a secure server with relevant information including case number, patient’s name and diagnosis.

Note: Call pathology assistant, senior resident, surgical pathology fellow or attending staff for assistance with technical photography problems.
Submitting Tissue

The following guidelines should be observed in selecting and submitting tissue for microscopic study:

- Small biopsies that will fit in one cassette are generally totally submitted.
- Diagnostic biopsies of larger size may need to be entirely submitted, but there are exceptions. See specific organ instructions for sampling.
- **Excisional** biopsies containing a tumor should be blocked to show margins.
- India ink (or equivalent) can be used to mark margins. **Do not allow the ink to spread elsewhere!**
- By convention, sections will be cut from the SIDE FACING DOWN in the cassette. If there is any reason to orient the specimen another way, put instructions on work sheet (i.e. "on edge").
- Tissues must be **THIN** (2-3 mm or less than the thickness of the cassette) and must **not be crowded** into the cassette. Thick or crowded tissue cannot be processed properly and bad sections will result, especially tissue containing fat, such as breast.
- In general, fix large specimens, especially bowel resections, laryngectomies, lungs before cutting. Thinner, better anatomically oriented sections will result.

Assignment to Surgical Pathology

A schedule is provided for the residents who are assigned to the Division of Surgical Pathology. The rotation ensures exposure of each resident to specimens obtained from various services. The normal working hours are from 7:30 a.m. to 5 p.m., Monday through Friday. It is important that at least one resident be available during the evening and night hours and on weekends.

In the early part of the afternoon, daily, the residents are expected to complete their routine cases received on the preceding day with the attending pathologist. Specimens that arrive late Friday afternoon must be processed on Saturday morning. At that time, rush cases and delayed cases should also be completed with the senior staff. It is imperative that this schedule be followed, otherwise significant delay in providing the reports to the physicians will occur.

Frozen Sections

Frozen sections are an important method for rapid diagnosis when a surgeon has the greatest need to know the exact nature of the lesion. A frozen section should be technically excellent and it should not take more than 20 minutes to perform.

A frozen section diagnosis must be worded as clearly as possible. Before communicating the frozen section diagnosis with the surgeon, residents must request confirmation of the identity of the patient by utilizing **two identifiers** including medical record number and patient’s name. The diagnosis must be written on the surgical pathology request form prior to reporting the diagnosis to the surgeon.

The frozen section area should be kept in a state of optimal performance. Before leaving the frozen section area, the area must be tidy, clear of all evidence of prior work in order to enable the next person to work in the same area with maximum efficiency.
The Chief Residents will instruct the first year residents of how to do the frozen sections. By the time the first year residents rotate through surgical pathology, they should be ready to do the frozen sections with good quality on the first day.

CONFERENCES

There is a Surgical Pathology Conference from 8 - 9 a.m., Mondays (organ specific conferences) and Thursdays (unknowns) of each week. These are "working conferences" during which time all difficult and interesting cases are reviewed and discussed prior to final diagnosis. Participation in this conference by staff members assigned to these cases is mandatory. Residents are required to study these cases before coming to the conference.

During the conference, each resident is asked about the diagnosis and the reason behind it. By the time a conclusion is reached, residents should be able to realize what they have missed. With the multiple headed microscopes, it is easy for the students to follow the changes discussed.

Senior residents from 2nd year up will take turns presenting at the Tumor Board Conference, which is held at 7:30 a.m. on Fridays. Residents will review the selected cases with the staff, take the pictures and read about the cases. The resident on hematopathology (1st year and up) will present cases at the Hematopathology Conference (weekly on Thursdays at 8:20 a.m.).

The monthly Morbidity and Mortality Conferences will be presented by the resident (1st year and up) who signed out the case. Additionally, 4th year and other senior residents actively participate and present the pathology portion of the weekly Breast and ENT Conferences.

Other subspecialty conferences held weekly:

- Soft tissue and bone (monthly).
- Dermatopathology (weekly).
- Benign breast radiology/pathology correlation (weekly).
- Pulmonary (bi-weekly).
- Liver (weekly).
- GI (weekly).
- GYN (weekly).
- Colposcopy (bi-monthly).
- Renal pathology (weekly).
- Cytology.

These conferences are presented by pathology staff. Residents are encouraged to attend these conferences. The Clinico-Pathology Conference (monthly), which is presented by a faculty member, is a required conference for residents to attend.
RESIDENT EVALUATION

At the conclusion of each rotation month, the resident will be evaluated by the surgical pathology staff to assess:

- Overall performance of duties.
- Accomplishment of goals.
- Progress in understanding pathology and the specifics of surgical pathology.

To achieve an excellent rating, residents should strive for the following:

- High self-motivation.
- Show unusually good understanding of gross and microscopic pathology and its relationship to clinical findings.
- Literature review of difficult and interesting cases.
- Excellent interpersonal relationships.
- Conscientious, rapid, accurate conduct of duties with minimal supervision.
- Involvement in investigative studies and case reports for publication.
- Development of excellent teaching skills.
GOALS AND OBJECTIVES

The goal of the rotation in anatomic pathology at the Veteran's Affairs Medical Center is for the resident to become competent in the basic practice and principles of diagnostic surgical pathology. Goals are based on Skill Levels I and II.

<table>
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<tr>
<th>Legend for Learning Activities for Residents</th>
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<td>Didactic lecture</td>
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<td>Faculty sign-out</td>
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<td>Journal club</td>
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<td>Directly supervised procedure</td>
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<td>Role modeling</td>
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<td>Lab inspections</td>
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<td>Interdisciplinary conference</td>
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<td>Online tools</td>
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<td>Unknown slide conferences</td>
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<td>Project</td>
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<th>Legend for Evaluation Methods for Residents</th>
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<td>Report review</td>
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<td>Direct observation</td>
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<td>Checklist</td>
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<td>Global rating/faculty evaluation</td>
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<td>Standardized exam</td>
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<td>Practical slide exam</td>
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<td>In-house written exam</td>
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<tr>
<td>360 multisource rating</td>
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<tr>
<td>Portfolios</td>
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<tr>
<td>Procedures and case logs</td>
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KUMC Pathology Residency Manual
### SURGICAL PATHOLOGY VAMC CORE COMPETENCY: PATIENT CARE

**Goal:**
*Skill Level I:* Residents must demonstrate competence in processing of basic types of patient specimens and evaluation of clinicopathological aspects of Surgical Pathology.

*Skill Level II:* Residents must demonstrate competence in processing all types of patient specimens, frozen sections, utilizing ancillary studies, and generating clinically relevant diagnostic reports.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
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<tbody>
<tr>
<td>Demonstrate competence in basic specimen processing skills. (I)</td>
<td>DL, FSO, DSP</td>
<td>RR, DO, CL, GR/FE, SE, IWE, 360, PF</td>
</tr>
<tr>
<td>Demonstrate competence in selecting representative tissue samples for intraoperative frozen sections. (I)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate proficiency in interpreting and reporting frozen sections within 20 minutes of receiving a specimen for that purpose in the pathology laboratory. (II)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Be able to independently report the histopathologic aspects of routine and complex cases with attention to organization of diagnostic format, development of differential diagnosis, and ordering of necessary special stains and other ancillary techniques. (II)</td>
<td>DL, FSO, DSP, RM, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE, PF</td>
</tr>
</tbody>
</table>

### SURGICAL PATHOLOGY VAMC CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Skill Level I:* Residents must be able to evaluate normal histology and basic pathologic processes.

*Skill Level II:* Residents must demonstrate competence in histopathological diagnosis, grading and staging of tumors, and interpretation of ancillary studies.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Determine when a microscopic description and/or interpretation is necessary, and provide such information. (I)</td>
<td>FSO, RM, USC</td>
<td>RR, DO, GR/FE, PF</td>
</tr>
<tr>
<td>Be able to evaluate margins of tumor resection specimens using frozen sections. (I)</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, CL, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the common situations requiring expedited processing of a pathology specimen, and those that do not. (II)</td>
<td>FSO, RM</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
### Demonstrate knowledge of the common indications for an intraoperative consultation. (II)

| FSO, DSP, RM | DO, GR/FE |

### Enumerate the indications and the limitations pertaining to intraoperative frozen section examinations. (II)

| FSO, DSP, RM | DO, GR/FE |

### Demonstrate knowledge of the common grading and staging systems applied to malignant neoplasms. (II)

| DL, FSO, DSP, IC, OT, USC | RR, DO, CL, GR/FE, SE, IWE |

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**SURGICAL PATHOLOGY VAMC CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**

*Skill Level I: Residents must be familiar with laboratory workflow, and laboratory safety practices.*

*Skill Level II: Residents must be consistently fast and competent at signing out cases, generation of microscopic descriptions, and notification of clinicians.*

**Objectives:**

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
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<tbody>
<tr>
<td>Know the procedures for the reporting of untoward incidents in the laboratory. (I)</td>
<td>DSP, RM, LI</td>
</tr>
<tr>
<td>Demonstrate knowledge of how and when to obtain external consultations in anatomic pathology and document the results appropriately. (II)</td>
<td>FSO, DSP, RM</td>
</tr>
<tr>
<td>Demonstrate an ability to manage workflow in the gross room, assist junior residents with gross dissection, provide accurate gross descriptions of routine and complex specimens, use the local anatomic pathology laboratory information system, and practice safety in the pathology laboratory. (II)</td>
<td>FSO, DSP, RM</td>
</tr>
<tr>
<td>Demonstrate knowledge of available procedures for locating a missing specimen and resolving questions of specimen identity. (II)</td>
<td>FSO, DSP, RM</td>
</tr>
<tr>
<td>Demonstrate knowledge of quality control pertaining to histologic sections and special stains, including trouble-shooting of mistakes in accessioning, labeling, &amp; misidentification of specimens. (II)</td>
<td>FSO, DSP, RM</td>
</tr>
</tbody>
</table>
SURGICAL PATHOLOGY VAMC CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

Goal:
Skill Level I: Residents must be familiar with the elements of a Surgical Pathology report and the information it conveys.

Skill Level II: Residents must be competent in constructing a comprehensive Surgical Pathology report and in written and verbal communication with other healthcare providers.

Objectives:

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
</table>
| Demonstrate knowledge of the common and basic elements of the surgical pathology report, including: (I)  
  - Identifiers (patient and institution).  
  - Input from the responsible pathologist.  
  - Input from the responsible clinician.  
  - Necessary dates and times that must be in the report.  
  - Necessary clinical information.  
  - Documentation of the specimens that were submitted.  
  - Thorough and accurate gross description. | FSO, DSP, RM | RR, DO, GR/FE |
| Demonstrate the ability to effectively construct a complex surgical pathology report. (II) | FSO, DSP, RM | RR, DO, CL, GR/FE, PF |
| Be able to properly prepare synoptic surgical pathology reports for common malignancies. (II) | FSO, DSP, RM | RR, DO, CL, GR/FE, PF |

SURGICAL PATHOLOGY VAMC CORE COMPETENCY: PROFESSIONALISM

Goal:
Skill Levels I and II: Residents must demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to diverse populations within the healthcare environment.

Objectives:

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
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<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
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<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM, IC</td>
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<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>FSO, DSP, RM</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM, IC</td>
</tr>
</tbody>
</table>

84 KUMC Pathology Residency Manual
| Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research. | DL, OT | GR/FE, SE |
| Demonstrate a commitment to excellence and ongoing professional development. | RM | DO, GR/FE |
| Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team. | FSO, DSP, RM, IC | DO, GR/FE, 360 |

**SURGICAL PATHOLOGY VAMC CORE COMPETENCY: SYSTEM-BASED PRACTICE**

**Goal:**  
*Skill Levels I and II*: Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.

<table>
<thead>
<tr>
<th><strong>Objectives:</strong></th>
<th><strong>Learning Activities</strong></th>
<th><strong>Evaluation Activities</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>Show a working knowledge of the basic principles of quality assurance, quality control, continuous quality improvement, and outcomes analysis, as they apply to anatomic pathology.</td>
<td>FSO, RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the standards (TJC, CAP) required for submitting surgical pathology specimens.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the basic recommendations/requirements (TJC, CAP, regional legal requirements) pertaining to retention of pathology specimens and records.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the basic principles of informatics in anatomic pathology, and ability to effectively utilize the local computer network.</td>
<td>RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Understand federal and state regulations with special application to anatomic pathology, and the general requirements for compliance in the reporting of professional activities for billing purposes.</td>
<td>FSO, RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of web-based or organization (CAP, ASCP, USCAP, etc.)-related learning and CME tools in anatomic pathology.</td>
<td>RM, OT</td>
<td>DO</td>
</tr>
<tr>
<td>Demonstrate a familiarity with standards set forth by the CAP and TJC for laboratory certification in anatomic pathology, and participate in at least one internal (“mock”) inspection of the institutional anatomic pathology laboratory.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Understand the principles applying to evaluation of the cost-effectiveness of laboratory procedures in anatomic pathology.</td>
<td>FSO, RM</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
PGY SURGICAL PATHOLOGY VAMC-SPECIFIC GOALS

PGY 1 GOALS: By the end of the first year:

- The resident has documented direct supervision of grossing the first three specimens in the majority of specimens on the organ-based list of specimens.
- The resident can dictate informative gross dictations with proper cassette summaries and cut appropriate sections without direct supervision on all biopsies and simple routine specimens and most common cancer cases.
- The resident becomes efficient in managing cases, such that an appropriate around time is observed.

PGY 2 GOALS: By the end of the second year:

- The resident has documented direct supervision of grossing the first three specimens for all specimens on the organ-based list of specimens.
- The resident demonstrates the ability to work up cases properly, including ordering appropriate histochemical and immunohistochemical stains.
- The resident demonstrates efficiency and professionalism in the handling of cases (turnaround time is kept to 48 hours, special stains, immunostains are ordered when the attending staff requests them, the resident does the follow-up on stains when they do not arrive when anticipated).
- The resident demonstrates an economy of sections that are adequate to provide all the necessary information, and minimizes the need to submit additional wet tissue.
- The resident demonstrates the ability to communicate appropriately to clinical colleagues.
- The resident is ready to start supervising (in the third year) junior residents in surgical pathology procedures.

PGY 3 GOALS: By the end of the third year:

- The resident must be able to compose a gross and microscopic surgical pathology report which is ready for transcription, with minimal, if any, correction.
- The resident should be comfortable interpreting intraoperative consultations.

PGY 4 GOALS:

- The resident should be able to supervise junior residents in all aspects of the practice of surgical pathology.
- The resident should have demonstrated increased medical knowledge through study set examinations, conferences and conducting clinical conferences.
- The resident must have reviewed minimally 2,000 surgical pathology cases that they have reviewed and signed out.
- The resident should be competent in all surgical pathology Skill Levels 1 and 2.
GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice-Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning found on page 20-21.
SURGICAL PATHOLOGY VAMC RESIDENT DUTIES AND RESPONSIBILITIES

Surgical Pathology

- **Frozen sections**: Residents together with staff perform the gross processing of all frozen sections, except when previously arranged with the staff. Residents should always be immediately available by pager.
- **Gross**: Residents examine, dissect and dictate all gross surgical specimens, with direct or indirect supervision by the staff. Residents are expected to notify staff of any questions regarding gross processing, including specimen orientation, sectioning, special studies and photographs. Grossing methods for skin, lymph node, and prostate, etc. are different from KU. Seeking directions from senior residents or staff is recommended. As a courtesy to other residents and staff, grossing resident is required to thoroughly clean the grossing station at the end of each day.
- **Microscopic**: All specimens processed grossly by residents will be independently examined microscopically by the resident the following morning. They will proofread the previous day’s gross dictation, check the completion and correctness of the paperwork, thoroughly evaluate all histologic sections and write out their topographical statement and diagnostic evaluation. As arranged, the resident will then meet with staff and sign-out all cases.
- **Self-study**: The resident is expected to investigate the diagnostic possibilities and related areas of each case using the available resources (including textbooks, study set of prior surgical pathology cases, CAP-PIP cases, ASCP cytology cases, and online resources).
- **Duty hours**: Unless attending required morning conferences or arranged with staff, residents are expected to stay in service from 8 a.m. to at least 5 p.m.

**CONFERENCES**

Residents will attend VA pathology conferences and those KU pathology conferences that are required. Residents may attend other KU conferences if approved by VA staff. Presentation of VA surgical pathology cases at KU conferences is strongly encouraged. VA staff will assist in photographic preparations as needed.

**Investigation**

Residents are encouraged to develop investigative studies and prepare case reports for publication together with the staff.

**RESIDENT EVALUATION**

- The resident should demonstrate the personal qualities of a mature and proficient pathologist, such as motivation, integrity, realistic self-assessment, and good communication skills.
- The resident should show improvement in knowledge and practice of surgical pathology.
- The resident must be reliable and responsible for working up, and presenting pathology cases as indicated.
RECOMMENDED READING LIST:

PEDIATRIC SURGICAL PATHOLOGY - CHILDREN'S MERCY HOSPITAL

CMH SURGICAL PATHOLOGY FACULTY

Eugenio Taboada, MD  Director, Surgical Pathology Services
Robert Garola, MD  Clinical Associate Professor of Pediatric Pathology, UMKC SOM
Alexander Kats, MD  Director, Nephropathology
Lei Shao, MD  Director, Autopsy Services
Vivekanand Singh, M.D.  Director, Gastroenterology Pathology Services

GOALS AND OBJECTIVES

The goal of the rotation in pediatric surgical pathology at Children's Mercy Hospital is for the resident to become competent in the basic practice and principles of diagnostic pediatric pathology, primarily surgical pathology.

<table>
<thead>
<tr>
<th>Legend for Learning Activities for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
</tr>
<tr>
<td>Faculty sign-out</td>
</tr>
<tr>
<td>Journal club</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
</tr>
<tr>
<td>Role modeling</td>
</tr>
<tr>
<td>Lab inspections</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
</tr>
<tr>
<td>Online tools</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
</tr>
<tr>
<td>Project</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Legend for Evaluation Methods for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
</tr>
<tr>
<td>Direct observation</td>
</tr>
<tr>
<td>Checklist</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
</tr>
<tr>
<td>Standardized exam</td>
</tr>
<tr>
<td>Practical slide exam</td>
</tr>
<tr>
<td>In-house written exam</td>
</tr>
<tr>
<td>360 multisource rating</td>
</tr>
<tr>
<td>Portfolios</td>
</tr>
<tr>
<td>Procedures and case logs</td>
</tr>
</tbody>
</table>
# Pediatric Surgical Pathology CMH Core Competency: Patient Care

**Goal:**
Residents must demonstrate a satisfactory level of diagnostic competence in pediatric pathology.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Perform adequate gross processing and dissection of pediatric surgical specimens.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, PF</td>
</tr>
<tr>
<td>Demonstrate competency in diagnosis of pediatric pathology histologic sections</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, PF</td>
</tr>
</tbody>
</table>

# Pediatric Surgical Pathology CMH Core Competency: Medical Knowledge

**Goal:**
Demonstrate knowledge about established and evolving biomedical, clinical and cognitive sciences and the application of this knowledge to pediatric pathology.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of pediatric pathology disorders.</td>
<td>FSO, DSP</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE</td>
</tr>
<tr>
<td>Evaluate the significant clinicopathological aspects of pediatric surgical pathology cases.</td>
<td>FSO, DSP, IC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE</td>
</tr>
</tbody>
</table>

# Pediatric Surgical Pathology CMH Core Competency: Practice-Based Learning & Improvement

**Goal:**
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

<table>
<thead>
<tr>
<th>Objectives</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM, OT, P</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>FSO, JC, RM, OT, P</td>
<td>DO, CL, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM, USC</td>
<td>RR, DO, GR/FE, SE, PSE, IWE, 360</td>
</tr>
</tbody>
</table>
Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.

| PEDIATRIC SURGICAL PATHOLOGY CMH CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS |
|---|---|
| **Goal:** | Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families. |

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<thead>
<tr>
<th><strong>Objectives:</strong></th>
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<tr>
<td>Demonstrate the ability to write articulate, legible, and comprehensive yet concise reports and consultation notes. Provide a clear and informative report, including a precise diagnosis whenever possible, a differential diagnosis when appropriate, and recommended follow-up or additional studies as appropriate.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, 360</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>FSO, RM, IC, USC</td>
<td>RR, DO, GR/FE, PSE, 360</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC, USC</td>
<td>DO, GR/FE, PSE</td>
</tr>
</tbody>
</table>

| PEDIATRIC SURGICAL PATHOLOGY CMH CORE COMPETENCY: PROFESSIONALISM |
|---|---|
| **Goal:** | Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population. |

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<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
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Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter. | DL, FSO, DSP, RM, IC | DO, GR/FE, 360
---|---|---
Demonstrate a commitment to excellence and ongoing professional development. | RM | DO, GR/FE
Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team. | FSO, DSP, RM, IC | DO, GR/FE, 360

**PEDIATRIC SURGICAL PATHOLOGY CMH CORE COMPETENCY: SYSTEM-BASED PRACTICE**

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

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<tr>
<td>Demonstrate understanding of the role of the laboratory in the healthcare system.</td>
<td>DL, FSO, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Show a working knowledge of the basic principles of quality assurance, quality control, continuous quality improvement, and outcomes analysis, as they apply to pediatric pathology.</td>
<td>FSO, RM, LI</td>
<td>DO, GR/FE</td>
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<tr>
<td>Demonstrate a familiarity with standards set forth by the CAP and TJC for laboratory certification.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
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SURGICAL PATHOLOGY CMH RESIDENT DUTIES AND RESPONSIBILITIES

- **Gross**: Residents examine, dissect and dictate all gross surgical specimens, with the staff always immediately available. Residents are responsible for conducting gross examinations not routinely done by the PA and will include all large and tumor excisional biopsies. Residents are expected to notify staff of any questions regarding gross processing, including specimen orientation, sectioning, special studies and photographs.

- **Microscopic**: All specimens processed grossly by residents will be independently examined microscopically by the resident the following morning (or when sections are available). They will proofread the previous day's gross dictation, thoroughly evaluate all histologic sections and write out their topographical statement and diagnostic evaluation. As arranged, the resident will meet with the staff and sign-out all cases.

- **Self-study**: The resident is expected to investigate the diagnostic possibilities and related areas of each case using the available resources.

CONFERENCES

Residents will attend CMH pathology conferences and those KU pathology conferences that are required. The following is a listing of pediatric conditions that not uncommonly are biopsied or excised for pathologic evaluation. The resident should become familiar with these relatively common pediatric surgical specimens and their histologic features. The resident is expected to develop more in depth understanding and higher skill level assessing conditions that appear in upper case letters. Many of these conditions also occur in adults and may have similar or dissimilar features and manifestations in children. It is unlikely that the resident will see most of these specimens during a one month pediatric surgical rotation. However, they should supplement their exposure by reviewing the teaching collection and through readings and exposures to these conditions that occur on other surgical rotations as well.

Head and Neck:

- Cysts and developmental anomalies
- Mucocele
- Developmental
- Dermoid / epidermoid
- Branchial cleft derivatives
- Accessory tragi (auricles)
- Preauricular pits and skin tags
- Branchial cleft cysts and sinuses
- Odontogenic jaw (dentigerous, keratocysts)
- Fissural (nasopalatine and globulomaxillary)
- Nasal / paranasal encephalocele and nasal “glioma”
- Nasopharyngeal hairy polyp (hamartoma)
- Acquired conditions
- Allergic polyps, polyps associated with CF, Cholesteatoma
- Tumors
• Pyogenic granulomas
• Hemangiomas
• Congenital epulis (granular cell tumor)
• Cystic hygroma
• Pigmented neuroectodermal tumor of infancy
• Neurofibromas
• Rhabdomyosarcoma
• RETINOBLASTOMA
• Olfactory neuroblastoma
• Lymphomas
• LANGERHANS CELL HISTIOCYTOSIS
• Congenital Fibrosarcoma
• Fibromatoses (fibromatosis coli) and myofibromatosis

Respiratory System:

• Upper
  • Laryngeal webs, granulation tissue and laryngotracheal malacia
  • Juvenile laryngeal papillomas
  • Hemangiomas
  • Lower
  • Cysts and developmental anomalies
  • Bronchogenic cysts
  • PULMONARY SEQUESTRATION (extra and intra lobar)
  • Infantile lobar emphysema
  • CONGENITAL CYSTIC ADENOMATOID MALFORMATION
• Acquired conditions
  • Persistent Interstitial Pulmonary Emphysema
  • Pulmonary hemosiderosis
  • Pneumonias (opportunistic infections)
  • Tumors
  • Lung
  • Inflammatory pseudotumor
  • Pleuropulmonary blastoma
  • Adenocarcinoma
  • Metastatic tumors
  • Chest wall
  • PNET (Askin tumor)
  • RMS associated with CCAM
  • Chest Wall hamartoma (mesenchymoma of rib)
**Mediastinum**

- Anterior
  - Thymic epithelial cysts
  - Teratomas / YST / EC / Choriocarcinoma
  - LYMPHOMAS (LB and HD)
- Middle
  - Bronchogenic cysts
  - Pericardial cysts
  - Lymphoma
- Posterior
  - Neuroblastoma
  - Ganglioneuroma
  - RMS
  - Neurofibroma and neurolemmoma

**GI tract**

- Congenital
  - Duplication cysts and diverticula
  - Heterotopias (gastric, pancreatic)
  - Atresia and Congenital webs (e.g. duodenal)
  - HIRSCHSPRUNG’S DISEASE
  - Cystic fibrosis (meconium ileus and equivalent)
  - Microvillus inclusion disease
- Acquired
  - NECROTIZING ENTEROCOLITIS (NEC)
  - Reflux and allergic esophagitis
  - Intussusception
  - Volvulus;
  - Gastritis (Helicobacter pylori, Hypertrophic) and gastropathies
  - Inflammatory bowel disease
  - Celiac disease
  - Allergic gastroenteritis
  - Allergic proctitis
  - Giardiasis
  - Tumors
  - POLYPS (juvenile, Peutz-Jeghers hamartomatous, adenomatous, lymphoid hyperplasias)
  - B cell lymphomas (Burkitt's)
  - Adenocarcinoma
  - Anal Warts (HPV)
  - Mesenteric lymphangiomas
  - Hemangiomas
Liver and exocrine pancreas

- Congenital
- Annular pancreas
- Ectopic pancreas (stomach, duodenum, liver)
- BILIARY ATRESIA (extrahepatic and Intra hepatic)
- Choledochal Cysts
- Bile Duct Plate malformations
- Infantile AR Polycystic disease
- Congenital Hepatic Fibrosis
- Intrahepatic BD Cysts (Carolí’s disease)
- METABOLIC DISEASES
  - Amino acids [tyrosinemia, Urea cycle defects, cystinosis]
  - Bile Acids [PFIC, BA synthetic defects, Zellweger]
  - Carbohydrates [GSD, galactosemia, DM]
  - Glycoproteins [alpha-1-Antitrypsin deficiency]
  - Minerals [Wilson’s, Hemochromatosis, Indian Childhood cirrhosis]
  - GM1 and GM2
  - Lipids [MCAD, Gaucher’s, Niemann Pick, abetalipoproteinemia, gangliosidoses, others
  - Mucopolysaccharidoses {Hunter’s and Hurler’s disease}
  - Others [Cystic fibrosis]
  - Infection (CMV, Adenovirus, Herpes simplex, Toxo)
  - TPN and drug-related
  - Sepsis-related
  - Hypothyroidism and Hypopituitarism
  - Tumors
  - Hemangiomas, hemangioendotheliomas
  - Hepatoblastomas
  - Undifferentiated Sarcoma
  - Mesenchymal Hamartoma
  - Teratoma
  - Hepatocellular carcinoma
  - Focal Nodular Hyperplasia
  - Hepatic Adenomas
  - Rhabdomyosarcom
  - Pancreatoblastoma
  - Papillary cystic and solid tumor of pancreas

Urinary tract

- Congenital
- Renal ectopias (simple vs crossed)
- Fused horseshoe kidney
- Renal Agenesis (unilateral and bilateral (Potter’s Sequence)
- Collection system duplications
- Obstructions (UPJ, PUV)
- RENAL DYSPLASIA (bilateral / unilateral, multicystic, segmental, hypoplastic variants)
- POLYCYSTIC KIDNEY DISEASE (AR, AD)
- Glomerulocystic Disease (AD and sporadic)
- Medullary cystic disease (Medullary sponge and familial nephronophthisis-MCD complex)
- Umbilical urachal remnants
- Renal nephrogenic rests (Wilms’ tumor “seeds”)
- Acquired
- Glomerulonephritis
- Nephrotic syndrome: MCD, FSGS, Diffuse Mesangial Sclerosis, Membranous Nephritis
- Hematuria with normal renal function: IgA (Bergers), HSP, Alports, TMB disease
- Nephritic syndrome, Post infectious GN, HSP, HUS, MPGN, Lupus
- Renal Transplantation
- Acute cellular and humoral rejection
- Chronic transplant nephropathy
- BK virus and other opportunistic infections
- Tumors
- WILMS” (triphasic and anaplastic variants)
- Congenital Mesoblastic Nephroma (myofibroma-like)
- Clear Cell Sarcoma
- Rhabdoid
- Renal Cell CA
- Bladder / prostate Rhabdomyosarcoma

Female reproductive system

- Congenital
- Streak Ovary and ovotestis
- vaginal adenosis (DES exposure)
- Vaginal mesonephric duct remnants
- Acquired
- Ovarian Torsion
- Follicular and leuteinizing ovarian cysts
- Massive ovarian edema
- Lichen sclerosis et atrophicus
- Condylomata (genital warts)
- Tumors
- Germ cell tumors (EST, Mature and Immature Teratoma, gliomatosis peritonea, etc.)
- Gonadoblastoma
- Gonadal stromal tumors (GC and JGC tumor, Sertoli-Leydig cell, SCTAT, Lipoid, etc.)
- Lymphoma (Burkitt’s)
- Hemangiomas, lymphangiomas and neurofibromas
- Inflammatory and benign fibroepithelial vaginal polyps
- Rhabdomyosarcoma, botryoid variant
- Vaginal mesonephric papilloma
- Perineal aggressive angiomyxoma

**Male reproductive system and intersex anomalies**

- Congenital
- Ectopic Testis and Cryptorchism
- Testicular regression syndrome. No increased risk of malignancy. (+ Vas / epididymis; siderocalcific fibrous nodule; no testis. +/- female/ambiguous/micro penis / normal male if bilateral or unilateral and depending on early, mid, or late fetal onset of infarction.
- Mixed gonadal dysgenesis (46,XY/45X mosaic) - variable feminization. Increased risk if malignancy.
- Partial streak on one side having ovarian stroma with rudimentary cords and tubules and ipsilateral müllerian development (failed MDIF production).
- Cryptorchid testis with histologic disorganized tubules and suppressed müllerian development on opposite side.
- Male pseudohermaphrodite: Genotypic male phenotypically feminized caused by: androgen insensitivity syndromes; gonadotropin abnormalities; Leydig cell abnormalities; testicular regression syndrome; testicular steroid enzyme deficiencies; 5α reductase deficiency, 46,XY gonadal dysgenesis (SRY mutations).
- Failure to produce MDIF (early bilateral TRS [Swyer’s syndrome]
- Persistent Müllerian Duct syndrome - Genotypic and phenotypic male with uterus and upper vagina and normal testes.
- Androgen receptor insensitivity syndrome (also called “Testicular Feminization Syndrome)
- Complete: Female phenotype with cryptorchid testes that is normal before puberty but becomes dysplastic thereafter. X-linked recessive or X-linked dominant. Accounts for 80% or TFS. High risk of malignancy (>30% by 30 years).
- Incomplete: Female genitalia with some masculinization with further virilization at puberty. Testis near normal with maturation arrest at primary spermatocyte stage and normal interstitial and Sertoli cells. No increased risk of neoplasia.
- Acquired
- Torsion
- Infarction (TRS -late)
- Meconium periorchitis
- Tumors
- Epidermoid Cysts
- Adrenal rests
- Gonadal Stromal tumors
- Leydig cell tumors
- Nodular Leydig cell hyperplasia
- Testicular tumor of Adrenogenital syndrome
- JGC tumor
- Germ Cell Tumors
- PARATESTICULAR Rhabdomyosarcoma

**Exocrine System**

- Congenital
- Ectopic adrenal cortical tissue
- Congenital Adrenal Hyperplasia
- Adrenoleukodystrophy
- Ectopic thyroid tissue
- Thyroglossal duct remnants / cysts
- Ectopic parathyroid tissue
- Pancreatic nesidioblastosis
- Acquired Adrenal cortical hyperplasia
- Adrenal Medullary hyperplasia
- Dyshormonogenetic goiter
- Multinodular adenomatous thyroid hyperplasia
- Lymphocytic Thyroiditis
- Graves Disease
- Secondary hyperparathyroidism
- Primary hyperparathyroidism
- Tumors
- Pituitary adenomas
- Craniopharyngioma
- Langerhans cell histiocytosis
- Adrenal cortical adenoma
- Adrenal Cortical Carcinoma
- Pheochromocytoma
- NEUROBLASTOMA
- Papillary thyroid carcinoma
- True Follicular Thyroid carcinoma
- Medullary Thyroid carcinoma
- Pancreatic adenoma
- Hematopoietic system
- Congenital
- Ectopic thymus
- Diamond Blackfan Anemia vs TEC
- Kostmann’s Agranulocytosis
- Severe Combined Immunodeficiency Disease
- Bruton’s Agammaglobulinemia
- Thymic Aplasia / hypoplasia
- Acquired
• Infectious Lymph adenitis (granulomatous, cat scratch, toxo, EBV, etc.)
• Follicular hyperplasias
• Kikuchi disease
• Splenic changes in Hereditary Spherocytosis, ITP, Sickle cell anemia, Gaucher’s, Sea Blue histiocytosis and Niemann Pick disease
• Thymic cysts
• Tumors
• ACUTE LEUKEMIA (ALL and AML and subtypes)
• Transient myeloproliferative Disorder of Down’s Syndrome
• NON HODGKIN’S LYMPHOMAS (Burkitt, LB, Anaplastic LC)
• Hodgkin’s Lymphoma including NLPHD
• Histiocytoses (LCH, JXG, VAHS, RD disease)
• Metastatic tumors (Neuroblastoma, RMS, Ewings, others)
• Splenic epidermoid cyst, hemangiomas
• Thymomas

Bone and Soft tissue

• Soft tissues
• HEMANGIOMAS and variants (Common Infantile [CIH], Rapidly involuting congenital [RICH], Non involuting congenital [NICH], Cellular,
• Hemangiomatosis
• Lymphangioma
• Vascular malformations
• Fibrous Hamartoma of Infancy
• Infantile digital fibroma
• Fibromatosis coli,
• Infantile myofibromatosis
• Giant cell fibroblastoma
• Lipoblastoma and Lipoblastomatosis
• SACROCOCCYGEAL TERATOMA
• PERIPHERAL NEUROECTODERMAL TUMOR / EWINGS TUMOR FAMILY
• Bone
• Osteomyelitis
• Aneurysmal bone cyst
• Unicameral bone cyst
• Osteoid osteoma
• Eosinophilic granuloma
• Chondroblastoma
• Fibrous dysplasia
• Osteofibrous dysplasia
• Ewing sarcoma
• Osteosarcoma
RESIDENT EVALUATION

• The resident should demonstrate the personal qualities of a mature and proficient pathologist, such as motivation, integrity, realistic self-assessment, and good communication skills.
• The resident should show improvement in knowledge and practice of surgical pathology.
• The resident must be reliable and responsible for working up pediatric pathology cases, and presenting them to others as indicated.

RECOMMENDED READING LIST:

SURGICAL PATHOLOGY TRAINER

FACULTY

Ossama Tawfik, MD, PhD  Professor and Director – Surgical Pathology
Ivan Damjanov, MD, PhD  Professor
Fang Fan, MD, PhD  Professor
Garth Fraga, MD  Associate Professor
Rashna Madan, MBBS  Associate Professor
Kathy Newell, MD  Associate Professor
Da Zhang, MD, MSc  Associate Professor
Wei Cui, MD  Associate Professor
Janet Woodroof, MD  Associate Professor
Katie Dennis, MD  Assistant Professor
Elizabeth Friedman, MD  Assistant Professor

Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Activity</th>
<th>Code</th>
</tr>
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<tbody>
<tr>
<td>Didactic lecture</td>
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<td>Faculty sign-out</td>
<td>FSO</td>
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<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
</tr>
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<td>Unknown slide conferences</td>
<td>USC</td>
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<td>Project</td>
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Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Method</th>
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<tr>
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<td>Direct observation</td>
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<td>Global rating/faculty evaluation</td>
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<td>Practical slide exam</td>
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<td>In-house written exam</td>
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<td>Portfolios</td>
<td>PF</td>
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<td>Procedures and case logs</td>
<td>PCL</td>
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# Surgical Pathology Trainer Core Competency: Patient Care

**Goal:**
Senior residents will enhance their teaching skills by mentoring PGY1 residents on their first month of surgical pathology at KUMC.

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<tr>
<th>Objectives:</th>
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</thead>
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<tr>
<td>Train a PGY1 resident the skills necessary to independently function in the grossing and sign-out rooms.</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
<tr>
<td>Obtain graduated responsibility by overseeing the work of a PGY1 resident.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Train a PGY1 resident the skills necessary to perform an autopsy when requested.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
</tbody>
</table>

# Surgical Pathology Trainer Core Competency: Medical Knowledge

**Goal:**
Demonstrate knowledge about established surgical pathology procedures to enhance the training of a PGY1 during the first month of surgical pathology at KUMC and enhance diagnostic knowledge.

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<tr>
<th>Objectives:</th>
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<tbody>
<tr>
<td>Demonstrate knowledge of basic surgical pathology procedures for use in training of a PGY1 resident.</td>
<td>DL, FSO, JC, DSP, RM, IC, OT, USC, P</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE</td>
</tr>
<tr>
<td>Continue to develop diagnostic skills and knowledge with graduated responsibilities.</td>
<td>DL, FSO, JC, DSP, IC, OT, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE, PF, PCL</td>
</tr>
<tr>
<td>Develop additional knowledge of teaching and mentoring methods and styles.</td>
<td>DL, FSO, JC, DSP, RM, IC, OT, USC</td>
<td>RR, DO, GR/FE, SE, IWE, PF, PCL</td>
</tr>
</tbody>
</table>
### SURGICAL PATHOLOGY TRAINER CORE COMPETENCY:
**PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**
_Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices._

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</thead>
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<tr>
<td>Demonstrate the ability to mentor a PGY1 resident in how to critically assess the scientific literature.</td>
<td>JC, RM, OT, P</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice while mentoring a PGY1 resident.</td>
<td>FSO, JC, RM, OT, P</td>
<td>DO, CL, GR/FE</td>
</tr>
<tr>
<td>Demonstrate to a PGY1 resident how to use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### SURGICAL PATHOLOGY TRAINER CORE COMPETENCY:
**INTERPERSONAL & COMMUNICATION SKILLS**

**Goal:**
_Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families._

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<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to instruct a junior resident performance of procedures in surgical pathology.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
<tr>
<td>Demonstrate the ability to provide focused, clear, and concise instructions.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the anatomic pathology service role to junior residents to develop clinically advantageous and cost-effective strategies.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE, PSE, 360</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>FSO, RM, IC</td>
<td>RR, DO, GR/FE, PSE, 360</td>
</tr>
<tr>
<td>Demonstrate skills in educating colleagues and other healthcare professionals: demonstrate the ability to help other residents obtain proficiency in laboratory medicine.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE, PSE, 360</td>
</tr>
</tbody>
</table>

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### SURGICAL PATHOLOGY TRAINER CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Interact with others without discriminating on the basis of religious,</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>ethnic, gender identity, or educational differences.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability,</td>
<td>FSO, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>and professional appearance.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>supersedes self-interest.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>both during and outside of a patient encounter.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>human subjects in research.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>development.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>multidisciplinary healthcare team.</td>
<td></td>
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</tr>
</tbody>
</table>

### SURGICAL PATHOLOGY TRAINER CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the clinical laboratory in the</td>
<td>DL, FSO, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>healthcare system.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans</td>
<td>FSO, RM</td>
<td>RR, DO, GR/FE, SE,</td>
</tr>
<tr>
<td>based on knowledge of best practices in collaboration with other clinicians.</td>
<td></td>
<td>IWE, 360, PF</td>
</tr>
<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>
Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.

Understand and implement policies to continually improve patient safety as they relate to surgical pathology.

<table>
<thead>
<tr>
<th>TRAINER RESPONSIBILITIES</th>
</tr>
</thead>
<tbody>
<tr>
<td>Quickly learning and becoming competent in the tasks necessary during the surgical pathology rotation is a daunting challenge for new residents. Not only are residents required to do the grossing work necessary to the rotation, and learn microscopic and diagnostic skills, but they are also required to work at a pace that facilitates appropriate turnaround time for cases, as well as adhering to work hour limitation requirements.</td>
</tr>
</tbody>
</table>

Adequate rotation specific training is an essential part of a first year resident's training, and this rotation aims to make that training more consistent. During the first block of a PGY1 resident’s experience on KU surgical pathology, a senior resident will be assigned a concurrent block to help PGY1 residents become independently competent during their rotation. This senior resident will be specifically paired with the PGY1 resident and will be in addition to the four residents ordinarily assigned to the rotation.

The goal for the trainer resident is to teach the PGY1 resident the skills necessary to independently function in the grossing and sign-out rooms; but it is also an opportunity for the trainer to take on graduated responsibility, to enhance their own learning, to add to their teaching skills, and to act more independently in preparation for their own post-graduation career work.

<table>
<thead>
<tr>
<th>TRAINER EVALUATIONS</th>
</tr>
</thead>
<tbody>
<tr>
<td>The trainer will be evaluated by the PGY1 resident assigned, the pathology assistant, and a group evaluation by the surgical pathology staff. The trainer will evaluate the surgical pathology staff and the rotation at the end of the month.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>TRAINER CONFERENCES</th>
</tr>
</thead>
<tbody>
<tr>
<td>The trainer will attend surgical pathology conferences, Tumor Board and other monthly conferences required of AP residents. The trainer will lead at least one surgical pathology conference during the month with faculty approval. The trainer will also attend the first day of the month orientation meeting.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>TRAINER DUTIES</th>
</tr>
</thead>
<tbody>
<tr>
<td>The trainer will be expected to be a daily presence in the workflow of the PGY1 resident and immediately available for questions and help. The ultimate goal is to provide assistance to the PGY1, including grossing supervision and working up cases more heavily in the first two weeks, and eventually acting as an advisor in the last two weeks.</td>
</tr>
</tbody>
</table>

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trainer will provide direct supervision of grossing of organ-specific cases that require direct supervision. The trainer will initial the PGY1 worksheet to verify the supervision of the required specimens. By the last week, the PGY1 resident should be doing the bulk of the work on their own, with the trainer available for questions.

**TRAINER SUPERVISION**

The trainer will be under the direct supervision of the Residency Program Director/AP Director. Questions or problems that arise during the month should be directed to the Residency Program Director. The trainer and the PGY1 will each meet with the Residency Program Director prior to the rotation to go over the rotation objectives and expectations.

**TRAINER HOURS**

Trainers will be expected to work concurrently with the PGY1 residents during the first two weeks of their initial block of surgical pathology, keeping within stated work hour limitations.

**Prior to Day 1 rotation, the trainer will:**

**TOUR:** Show the PGY1 necessary items including but not limited to:

- Frozen stations/gross room.
- Supplies/stainer/tools/EM/RPMI location.
- Processor room/Formalin.
- Photography.
- IHC/Histology.
- PA and histology supervisor office.
- Cytogenetics.
- Flow Cytometry.
- Phone/pager lists/OR phone list.
- Grossing templates on G: drive.
- Transcription.
- Cytology lab-(For STAT BAL training to be done by cytology fellow).
- Location of cytology lab key and meal card.
- Slide/block filing.
- Paperwork filing.
- Frozen section slide filing.

The trainer will also go over the surgical pathology orientation checklist with the PGY1, sign and return it to the Residency Program Director. Ensure the PGY1 is given their SP notebook and point out additional books/manuals locations. Go over the PGY1 grossing checklist and tape it up in the gross room.

**People authorized to sign off on the grossing/training checklist:**

- SP Trainer
- SP Fellow
- Pathology Assistant
- Faculty
SURGICAL PATHOLOGY TRAINER SCHEDULING

DAY 1: Working in conjunction with the pathology assistant as necessary, the trainer will:

- Train the PGY1 on setting up a grossing station.
- Train the PGY1 on frozen section cutting/sign-out/calling results.
- Train the PGY1 on the dictation system/Voicebrook.
- Assist in grossing. The trainer will be expected to stand at the PGY1’s side during grossing to assist with grossing and answer questions until the PGY1 is competent to gross each type of specimen independently. The trainer may assist in grossing as the workload requires.
- Explain the resident role in the tissue banking process.
- Explain the resident role in Oncotech/Precision sampling.
- Assist PGY1 and staff with reading frozen sections.
- Eventually independently read frozen sections with staff approval.
- Train the PGY1 on accessioning/printing cassettes/Vantage system.
- Train the PGY1 on cleaning/end of the day procedure/loading processor.
- Help triage specimens for grossing with pathology assistant.
- Help triage specimens for specialty testing including flow cytometry and cytogenetics.
- Work with PGY1 on grossing days until grossing work is finished.

DAY 2-3: Working with the approval of the staff, in assisting the PGY1 in signing out big cases, the trainer will:

- Train the PGY1 to organize slides and paperwork/gather pertinent clinical information.
- Train the PGY1 to enter cases in the CoPath system including but not limited to:
  - Gross description proofreading/editing.
  - Final Diagnosis and Comment sections.
  - Clinical History section.
  - Accurate checking of accessioning.
  - Addendums.
  - Tumor checklists.
  - Ordering IHC/specials/etc.
  - Ordering prognostic markers.
- Train the PGY1 on critical values in surgical pathology.
- Train the PGY1 on filing slides and completed paperwork.
- Train the PGY1 on entering and filing histology/IHC QA sheets.
- Train the PGY1 on autopsy procedure on days when no pathology assistant is available.

DAY 4:

- Assist the PGY1 in signing out biopsies (see above recommendations for sign-out and CoPath training).
- Train the PGY1 on the policy of STAT cases (endomyocardial, liver etc. Train the PGY1 on routinely ordered special stains (liver, gastric etc).
- Assist in preparing leftover Day 2 cases for staff sign-out.
- Train the PGY1 to prepare for Day 1 by:
  - Checking/printing the OR schedule.
  - Looking up pertinent history in CoPath AND O2.
  - Pulling relevant slides as necessary.
  - Reviewing gross manuals as needed for anticipated specimens.

**Signing Out**

It is recommended that the trainer sits with the PGY1 and look at cases together, while the PGY1 enters information in CoPath. Once the PGY1 becomes competent in independent entering, the PGY1 will be encouraged to quickly look at slides first, enter as much in CoPath as possible, and then review them with the trainer before staff sign-out if time permits. The trainer will not be required to sit with the staff and PGY1 for staff sign-out unless requested by the staff.

Additionally, the trainer will:
- Order special stains before staff sign-out as necessary.
- QA new cancer cases with staff.
- Assist PGY1 in fielding clinical questions/phone calls on pending cases.
- Communicate with PA and PGY1 regarding grossing of leftover cases.
- Sign-out cases with staff without the PGY1 present as necessary.
- Train the PGY1 on microscopic photography/scanning slides as necessary.
MEDICAL RENAL/ELECTRON MICROSCOPY

FACULTY

Ivan Damjanov, MD, PhD  Professor
Timothy Fields, MD, PhD  Professor
Da Zhang, MD, MSc  Associate Professor

The rotation experience in medical renal pathology takes place during the last two weeks of the PGY4 year in the Surgical Pathology Trainer block. The experience is designed to allow a resident to obtain training in routine medical renal pathology and as an introduction to electron microscopy.

GOALS AND OBJECTIVES

The goal of the medical renal rotation is that the resident becomes competent in the interpretation of medical renal material, primarily biopsies, and to learn how to incorporate electron microscopic and immunofluorescent studies into a cohesive diagnostic picture. Communication and consultative skills will be emphasized, that will aid the clinicians in the correct diagnosis and treatment of patients.

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### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate an increased level above the general residency competency within the chosen specific area or subspecialty within anatomic pathology.*

<table>
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<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Develop diagnostic competency within medical renal biopsy interpretation.</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, GR/FE,</td>
</tr>
</tbody>
</table>

### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive sciences and the application of this knowledge to specific area of pathology.*

<table>
<thead>
<tr>
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</thead>
<tbody>
<tr>
<td>Demonstrate knowledge of grossing techniques, proper fixation, and ancillary studies including electron microscopy and immunofluorescence.</td>
<td>FSO, DSP</td>
<td>RR, DO, GR/FE,</td>
</tr>
</tbody>
</table>

### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
*Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>FSO, JC, RM, OT,</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM, USC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.*

<table>
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<tr>
<th>Objectives</th>
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</thead>
<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>FSO, RM, IC, USC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

<table>
<thead>
<tr>
<th>Objectives</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### MEDICAL RENAL/ELECTRON MICROSCOPY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the laboratory in the healthcare system.</td>
<td>DL, FSO, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Show a working knowledge of the basic principles of quality assurance, quality control, continuous quality improvement, and outcomes analysis as they apply to pathology.

<table>
<thead>
<tr>
<th>Demonstrate a familiarity with standards set forth by the CAP and TJC for laboratory certification.</th>
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<tbody>
<tr>
<td>FSO, RM, LI</td>
</tr>
</tbody>
</table>

**DUTIES AND RESPONSIBILITIES OF THE RESIDENT**

Residents will be responsible for assisting in grossing medical renal biopsies, answering clinician inquiries, writing preliminary reports for native and transplant renal biopsies, assisting in triaging specimen for ancillary studies, attending interdisciplinary conferences, and signing out with the faculty.

**RESIDENT EVALUATION**

The resident will be evaluated by the faculty mentor using the global rating faculty evaluation form.
Residents will spend four blocks on the cytology service during years 1–4.

<table>
<thead>
<tr>
<th>Legend for Learning Activities for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
</tr>
<tr>
<td>Faculty sign-out</td>
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<tr>
<td>Journal club</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
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<tr>
<td>Role modeling</td>
</tr>
<tr>
<td>Lab inspections</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
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<tr>
<td>Online tools</td>
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<tr>
<td>Unknown slide conferences</td>
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<tr>
<td>Project</td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>Legend for Evaluation Methods for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
</tr>
<tr>
<td>Direct observation</td>
</tr>
<tr>
<td>Checklist</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
</tr>
<tr>
<td>Standardized exam</td>
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<tr>
<td>Practical slide exam</td>
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<tr>
<td>In-house written exam</td>
</tr>
<tr>
<td>360 multisource rating</td>
</tr>
<tr>
<td>Portfolios</td>
</tr>
<tr>
<td>Procedures and case logs</td>
</tr>
</tbody>
</table>
**Goal:**
Residents will learn the utilization of cytopathology as a non-invasive modality of diagnosis. This will include hands-on experience of the collection of samples during fine needle aspirations of superficial masses and assistance during the sampling of deep lesions.

<table>
<thead>
<tr>
<th>Objectives</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate knowledge of how to evaluate common cytopathology specimens comprehensively.</td>
<td>DL, FSO, DSP</td>
<td>RR, DO, GR/FE, SE, IWE, PCL</td>
</tr>
<tr>
<td>Demonstrate knowledge of the application of ancillary techniques including image analysis, immunocytochemistry, flow cytometry, cytogenetics, electron microscopy, and molecular studies (FISH; PCR).</td>
<td>DL, FSO, DSP, IC</td>
<td>RR, DO, GR/FE, SE, PSE, IWE, PCL</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to rapidly evaluate common FNA biopsy specimens, including determination of specimen adequacy and the need for ancillary techniques, and the appropriate collection of materials for such techniques.</td>
<td>DL, FSO, DSP, RM</td>
<td>RR, DO, GR/FE, SE, PSE, IWE, PCL</td>
</tr>
<tr>
<td>Demonstrate working familiarity with the instruments and materials needed to perform FNA biopsies.</td>
<td>DL, FSO, DSP</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate correct performance of FNA, including preparation of smears and collection of diagnostic materials with proper handling for ancillary techniques on appropriate specimens at the surgical pathology gross cutting area.</td>
<td>DL, DSP</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate competency under supervision of staff cytopathologists in the performance of a clinical superficial FNA biopsy; appropriately taking history, correctly obtaining informed consent, competently examining the lesion to be biopsied, preparing the patient and biopsy instruments, physically procuring the specimen, and preparing and staining the smears with preliminary interpretation of the smears and appropriate after-care of the patient.</td>
<td>DL, DSP</td>
<td>RR, DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>
## CYTOPATHOLOGY CORE COMPETENCY: MEDICAL KNOWLEDGE

### Goal:
Pathology residents will gain proficiency in cytopathology as required to gain successful certification in Anatomic or Anatomic and Clinical Pathology.

### Objectives:

<table>
<thead>
<tr>
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<tbody>
<tr>
<td><strong>Complete both a pre- and post-program assessment test.</strong> Pre-test should be taken after the first month of the program and the post-test performed after the third required month.</td>
<td>DL, FSO, JC, DSP, RM</td>
</tr>
<tr>
<td>Residents are required through graded increase of responsibility and participation in sign-out, supplemental learning materials, conferences, cytopathology research and FNAs to achieve the goal of resident training in cytopathology.</td>
<td>DL, FSO, DSP, RM, IC, USC</td>
</tr>
<tr>
<td>Demonstrate knowledge of the current Bethesda System terminology for reporting on gynecologic cytopathology specimens, and of the principles and application of human papillomavirus probe analysis.</td>
<td>DL, FSO, DSP</td>
</tr>
<tr>
<td>Demonstrate knowledge of the elements of adequacy and the current laboratory reporting system (such as negative, inflammatory/reactive, atypical/suspicious, neoplastic or malignant) for FNA biopsy and exfoliative non-gynecologic cytopathology specimens from the various commonly sampled body sites.</td>
<td>DL, FSO, DSP</td>
</tr>
<tr>
<td>Demonstrate knowledge of the cytopathologic features of normal, reactive, infectious, dysplastic and neoplastic conditions as seen in common cytopathology specimens.</td>
<td>DL, FSO, DSP, USC</td>
</tr>
<tr>
<td>Demonstrate knowledge of how common cytopathology specimens are screened.</td>
<td>DL, FSO, DSP</td>
</tr>
<tr>
<td>Demonstrate knowledge of the content of training materials on correct performance of FNA biopsies.</td>
<td>DL, FSO, DSP, RM</td>
</tr>
<tr>
<td>Demonstrate familiarity with the principles of automated screening for gynecologic cytopathology specimens.</td>
<td>DL, FSO</td>
</tr>
</tbody>
</table>
## CYTOPATHOLOGY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
*Residents must gain awareness of laboratory management and knowledge of quality assurance measures and laboratory troubleshooting.*

<table>
<thead>
<tr>
<th>Objectives</th>
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</thead>
<tbody>
<tr>
<td>Be able to verify that cytopathology requisitions are completed correctly.</td>
<td>FSO, DSP</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate familiarity with the methods of collection, cytopreparatory processing, and turnaround times for common cytopathology specimens, in order to be able to answer clinicians' questions concerning expected results from the cytopathology laboratory.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to perform quality assurance, including the correlation of gynecologic and non-gynecologic cytopathology with surgical pathology, both in aggregate for quality assurance purposes and on a case-by-case basis for diagnostic purposes.</td>
<td>FSO, DSP, RM, LI, IC</td>
<td>DO, GR/FE, SE, IWE, 360</td>
</tr>
</tbody>
</table>

## CYTOPATHOLOGY CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.*

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<tbody>
<tr>
<td>Demonstrate knowledge of how clearly, concisely, and completely to compose a cytopathology report for specimens from various commonly sampled body sites based upon the final diagnostic findings, and of how appropriately to recommend clinical follow-up.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>

## CYTOPATHOLOGY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

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<tr>
<th>Objectives</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Observe patients' rights to consent, privacy and compassion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate respect, regard, and integrity; a responsiveness to the needs of patients and society that supersedes self-interest; assume responsibility and act responsibly; and demonstrate a commitment to excellence and on-going professional development.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>
Demonstrate a commitment to ethical principles pertaining to provision or withholding of clinical care, confidentiality of patient information, informed consent, and business practices.

<table>
<thead>
<tr>
<th>Objective</th>
<th>DL, FSO, DSP, RM</th>
<th>DO, GR/FE, 360</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate sensitivity and responsiveness to diversity, including awareness of their own, their patients’ and colleagues’ ethnic, culture, age, gender, and disabilities.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>

**CYTOPATHOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE**

**Goal:**
*Residents must gain knowledge of the ethical, socioeconomic and medical-legal issues in the practice of cytopathology.*

**Objectives:**

<table>
<thead>
<tr>
<th>Learning Activities</th>
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</tr>
</thead>
<tbody>
<tr>
<td>DL, FSO, DSP, RM, LI</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>

Demonstrate knowledge of how to apply concepts of quality control, quality improvement, risk management, and of regulatory compliance including correct coding as these pertain to the practice of cytopathology.

Use system resources to provide care that is of optimal value.

Understand how their patient care and other professional practices affect not only patients, but also other healthcare professionals, the hospital, the medical center, University and the larger society, and how these organizational components of the system affect their own practice.

Know how medical practices and delivery systems differ from one another, including methods of controlling healthcare costs and allocating resources.

Learn how to practice cost-effect, high quality healthcare and resource allocation.

**PGY 1 GOALS: By the end of the first year:**

- Learn the basic elements of cytology such as sample preparation, and criteria for malignancy.

**PGY 2 GOALS: By the end of the second year:**

- The resident demonstrates competency in recognizing inflammatory reactive repair, LGSIL, HGSIL and carcinoma on pap smears and is able to report them out with the Bethesda System 2001.
- The resident demonstrates an improvement in medical knowledge in cytology at sign-out.
- The resident is ready to start supervising (in the third year) junior residents in cytology.
PGY 3 GOALS: By the end of the third year:

- Residents should be reviewing and signing out all types of cytopathology with minimal correction by the attending staff.
- The resident should be able to perform most fine needle aspirations without direct supervision, and produce diagnostic aspirations that are well-preserved, well-stained and with adequate cell button for ancillary studies.
- The resident must develop professionalism and interpersonal and communication skills that are respectful and compassionate toward patients, demonstrating cultural competency.

PGY 4 GOALS:

- The resident must have reviewed a minimum of 1,500 cytologies (pap smears, nongynecologic exfoliates and fine needle aspirations).
- The resident should be able to supervise junior residents in all aspects of cytopathology.
- The resident is practice ready for billing, Medicare compliance and accreditation issues.
- The resident should have received training and be certified in ThinPrep.
- The resident should be competent in all Cytology Skill Levels 1 and 2.

GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice--Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on page 19-20.

RECOMMENDED READING LIST:

Graded Responsibilities

The first block rotation focuses on the basic cytopreparatory techniques and basic diagnostic skills. Residents spend one week in the laboratory to learn the techniques on specimen collection and staining. Residents then spend one week learning screening of pap smears with a senior cytotechnologist or a cytopathology fellow. Residents are expected to review cases everyday with the cytopathology fellow and sign-out with the attending cytopathologist on a daily basis. The first month should cover negative, atypical, dysplastic, carcinoma in-situ, and invasive carcinoma of gynecological origin.

In the second though fourth rotations, residents learn more non-gynecologic cytology, including the fine needle aspiration technique, adequacy check on radiologically guided fine needle aspirations, cytology of the thyroid, salivary gland, breast, urinary tract, lymph node, soft tissue, gastrointestinal tract, and miscellaneous fluid specimens. Residents are expected to review cases that have been pre-screened by cytotechnologists, formulate a diagnosis, obtain any necessary follow-up or clinical information on the case and sign these cases out with the fellow and/or attending cytopathologist.

Residents are encouraged to use supplemental learning materials including study sets and books, participate in conferences and participate in research activities in cytopathology.

Policies and procedures

Residents are expected to become familiar with the Department of Pathology & Laboratory Medicine, Resident Manual, as well as the Graduate Medical Education Policy and Procedure Manual.

GENERAL

The resident will work closely with the cytotechnologists to learn technical principles and early interpretation as well as some principles of organization. Whereas the cytopathologists will be involved in all aspects of the training, instruction at the multi-headed microscope emphasizing interpretation and follow up will be the most essential aspect of the rotation.

This is done according to the following schedule:

10 a.m. to 12 noon: Fellow/resident review of cytologic material.
1 to 4 p.m.: Sign-out of cytologic material.

Turnaround time: (i.e. the time from cytotechnologist review to pathologist review and approval).
DEPARTMENTAL TURNAROUND TIME POLICY

The Department of Pathology & Laboratory Medicine expects that over 80% of all specimens will meet the following expected parameters. **Note:** All times, days and hours, are working days from delivery of the specimen.

- Uncomplicated non-gynecologic specimens will be completed within 24 hours from delivery to the laboratory.

- Complicated non-gynecologic specimens will be completed within 48 hours.

- Fine needle aspirations should have a preliminary report within two hours and will be signed out the same day unless special stains, immunohistochemistry or review of the biopsy or previous material is required. In that case, they will be done within 24 hours and a report will be discussed with the clinician and documented in the chart.

- Gynecologic cases will be screened within one week of receipt and reviewed by the cytopathologist within two days of transmittal to the pathologist. The entire report will be finalized within 10 days total.
SPECIFIC CYTOPATHOLOGY PROCEDURES

Fine Needle Aspirations (FNA)

Principle

The primary reason for doing fine needle aspiration cytology is to rule out or confirm malignancy or to diagnose infectious diseases. The use of FNA will result in a decrease in the number of open biopsies, as well as a decrease in healthcare costs.

Equipment:

- Twenty-three to 27-gauge sterile needles in a variety of lengths, with "see through" plastic hubs, and 10-20 ml disposable (preferably "slip tip") plastic syringes.
- Frosted end plain glass slides.
- Ninety-five percent alcohol in a Coplin jar or bottle to immerse glass slides.
- Slide tray for air dried slides.
- Gloves of appropriate size, wood applicator sticks, gauze, alcohol swabs and bandages.
- RPMI-1640 collection fluid (cell culture medium) for cell block and/or flow cytometry, if necessary for presumed hematopoietic/lymphoreticular disorders.
- CytoLyte Collection fluid for ThinPrep (preferably not PreservCyt initially).
- Cytology requisition forms.
- Institutional/hospital patient consent forms.
- No. 2 lead pencil.
- Vial of glutaraldehyde for electron microscopy, if necessary.
- Microscope.
- Cart.

Note:

- The routine practice of rinsing the needle into either Cytolyte or cell culture medium allows for maximum retrieval of cytologic material and may allow the resident to prepare a cell block that can later be used for ancillary studies.
- Excessively bloody aspirates will likely not make readable or very cellular direct smears. The resident should probably put most if not all of such aspirates directly into Cytolyte, fixative (if Formalin or alcohol is used, make sure that the laboratory knows in advance), or cell culture medium. Repeat such aspirates using a smaller gauge needle.
- Residents may not perform unsupervised FNAs without the permission of the Cytology Director.
- Fellows and residents may not render final interpretations.
- Fellows may perform FNAs alone, perform adequacy checks on FNAs, and communicate their preliminary findings to clinicians after an appropriate period of training as determined by program faculty.
FNA Technique - The successful FNA of palpable masses requires:

- Slides are clean and appropriately labeled with the patient's name. Discard any and all slides that were taken from the box for an individual case, but were not used. If there is any doubt about whether a slide belongs to a particular patient or case, please notify the cytology faculty on-call. Do not assume anything.

- A palpable mass (i.e. something the resident can clearly feel). Indistinct or poorly defined masses, without image guided assistance, are as likely to result in a non-diagnostic FNA as they are to yield a diagnostic result.

- Proper palpation and immobilization of the lesion. The skin is swabbed with an alcohol pad (local anesthesia is usually not required). The mass is often best immobilized between the index and middle fingers of the gloved left hand (for right-handed aspirators) or the gloved right hand (for left-handed aspirators).

- The needle, usually a 25 swg 1 1/4 inches long securely connected to a 10 ml syringe (which is in turn attached to the Cameco Syringe Pistol), is inserted in the lesion, while it is immobilized. THEN the piston of the syringe is retracted to create a vacuum in the syringe, NOT before the needle is inserted.

- The needle is moved gently back and forth several times within the mass. NOTE: Changing directions of the needle while it is embedded deep within the lesion will tear tissue unnecessarily and produce more bleeding. If the resident must redirect, pull the needle towards the subcutaneous tissue, without exiting the completely, change directions and repeat back and forth movements. Continue to move the needle back and forth about 10 to 15 times or until a flash of blood appears in the hub of the needle.

- The needle is detached from the syringe very carefully and the piston is retracted to fill the syringe with air. Reattach the need and then forcibly expel the material onto a clean glass slide, near the frosted end.

- Skillful preparation of the smears. The quality of the resident’s interpretation is greatly dependent upon the quality of the cytologic smears. A high yield FNA can be rendered non-diagnostic if the smears are not skillfully prepared. The one step technique is the one that residents should use most often and works best with “thick and creamy” specimens, i.e. non-cystic and not overly blood specimens.

- Slide preparation using the one-step technique is described as follows:
  - Hold the slide with the aspirated material in the left hand with the label or frosted end between the thumb and index finger, using the middle, fourth, and fifth fingers to stabilize the glass slide along its long edge.
  - A clean slide is held in the right hand between the thumb and index finger perpendicular to the other slide, which holds the specimen.
  - The edge of the clean slide closest to the resident (the aspirator/smearer) is placed on the slide with the specimen at a 45-degree angle, so that the edge furthest from the resident (the aspirator/smearer) is over the specimen.
  - Lower the clean slide onto the specimen and it will spread without force slightly by capillary action, continue to spread without force by guiding the top slide with middle finger of the right had along the edge of the slide which holds the specimen.

- The “Dab” technique or “Touch and Divide” is a method of making multiple smears from a single FNA pass or aspirate. This also works best with the “thick and creamy” specimens that one obtains from lymph nodes.

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are solid tumors. This technique is desirable when the resident wants to avoid an excessively thick slide and/or when ancillary studies might be better performed on fresh cytologic preparations. The "Dab" technique or "Touch and Divide" is described below.

- Expel the cellular material onto the slide as described above.
- Lightly touch the specimen the clean slide, close the end "furthest" from the label and near the long edge of the slide furthest from the aspirator.
- The touch or dab can be repeated on a different part of the slide (closer to the label) one or more times.
- Smear the "dabbed off" samples onto separate clean slides as described above and several slides can result from a single specimen.

**Patient Consent**

A signed consent formed must be obtained from the patient. The rule of thumb is that, a physician should obtain such consent "any time the skin is broken". The signed consent form stays with the patient's chart.

**Procedure Note**

A procedure note must be written in the patient's chart following the FNA performed by the pathologist for inpatients. The procedure note can be included as a part of the cytology report for all out-patient procedures. The procedure note should follow the SOAP format, including what can be used as subjective data (or what the resident is seeing the patient for/chief complaint), objective data (what the resident found in the limited physical examination), resident assessment, and resident plan. It is important for billing purposes to document the procedures in this manner.

It is also important to document in the procedure note that two patient's identifiers are confirmed and a "time out" is called to identify the site of aspiration before procedure.

**Fine Needle Aspiration Consultation Service Procedure Note (EXAMPLE):**

The FNA procedure was performed by Dr.___________. The staff pathologist, Dr.___________ was present throughout the procedure. Prior to beginning the procedure, two of the patient's identifiers (medical record number, date of birth, or patient name) were confirmed in addition to the proposed site of the FNA. The FNA procedure was explained to the patient, and with his/her informed consent a limited physical examination and the FNA procedure were performed. #_____ FNAs were performed on an approximately #____ cm mass using separate, sterile, #_____ swg needles. The procedure was well tolerated and without complications.

**Preliminary interpretation (EXAMPLE):**

This preliminary result was reported to Dr.__________ at__________ on__________ by Dr.__________. Defer final diagnosis until all cytologic material has been stained and evaluated.

Final Diagnosis to follow. Thank you for this interesting consult.

Dr. Smith (Beeper Number 1234) Location

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Comment (EXAMPLE):

The FNA procedure was performed by Dr. ____________. The staff pathologist, Dr. ____________ was present throughout the procedure. Prior to beginning the procedure, two of the patient's identifiers (medical record number, date of birth, or patient name) were confirmed in addition to the proposed site of the FNA. The FNA procedure was explained to the patient, and with his/her informed consent a limited physical examination and the FNA procedure was performed. #______FNAs were performed on an approximately #___ cm mass using separate, sterile, #____ swg needles. The procedure was well tolerated and without complications.

Preliminary interpretation (EXAMPLE):

This preliminary result was reported to Dr. ____________ at ___ on _________ by Dr. ______________.

Stat Specimens

When there is a request for a STAT result, the pathology faculty member on service or on-call should be notified immediately. If the STAT request includes a request for PCP evaluation and it is after hours or on the weekend, it is up to the pathology faculty member on service or on-call to approve such requests. The fellow or resident should contact that faculty member on service or on-call.

Calling Physicians with Results

All new cancer diagnoses should be called to the physician and there should be documentation on the patient's cytology report of that call, (i.e. the name of the physician or healthcare provider who was given the results, the date and the time of the call). Anytime an interpretation is called to a physician, whether preliminary or final, there should be documentation of the telephone call, the name of the physician or healthcare provided given the results, the date and time of the call should be included in the report.

Technical Principles

Residents will be exposed to the methodology of collection, fixation, cytopreparation, staining and screening cytological samples. They will also participate in the processing of cytology specimens for E.M., immunocytochemistry and flow cytometry.

Interpretation and Reporting

Residents will be instructed in the principles involved in examining cytological preparations, applying diagnostic criteria and reporting cytopathologic findings. Incorporation in the report of ultrastructural, immunocytochemical and ploidy information will enable the resident to integrate concepts from various disciplines.

Utilization and Feedback

Residents will participate in the interface between cytopathology and the clinicians utilizing the laboratory in terms of reports, recommendations and submission of adequate samples, patient instruction, and other significant items. Review of pertinent X-rays, CT scans, scintigrams and other data from the patient charts will be strongly stressed.
Follow Up and Quality Control

Residents will actively participate in correlating a cytopathology diagnosis with histopathologic data derived from biopsies or surgical extirpations. Residents will also evaluate specimens as to the adequacy of cytopreparatory and staining methods and learn to trouble shoot deficient areas.

Quality Assurance and Standards of Care

Residents are expected to familiarize themselves with the current guidelines and standards of performance and care. Residents are required to have knowledge of issues related to Quality Assurance in cytopathology, by reviewing the Cytopathology Policies and Procedures Manual (located in 1601 Bell Hospital), attending division meetings, and through self-study www.cytopathology.org.

RESIDENT EVALUATION

Residents receive monthly evaluations via MedHub. The program faculty on a monthly basis will evaluate fellows. The Residency Program Director will provide formal written evaluations (formative and summative) on a semi-annual basis.

Fellows and residents are evaluated in the following areas:

- Technical skill.
- Morphologic skills.
- Clinical judgment.
- Teaching.
- Research efforts.
- Core competencies.

Additionally, fellows and residents have the opportunity to discuss their cytology training with the Residency Program Director and/or Department Chair on a monthly basis. Fellows and residents are required to evaluate the cytopathology program and faculty on a regular basis. All evaluations are reviewed and discussed with the Chair. Fellows are encouraged to write a formal evaluation of the program at its conclusion.
AUTOPSY SERVICE

FACULTY/STAFF

Katie Dennis, MD  Assistant Professor and Director – Autopsy Service
Ivan Damjanov, MD, PhD  Professor
Jim Fishback, MD  Professor
Elizabeth Friedman, MD  Assistant Professor
Mark Myers  Coordinator and Supervisor, Autopsy Service

The autopsy is a major key in the study of our discipline. It is an invaluable tool to assess and assure quality control of patient care. The autopsy is performed in a complex institutional, administrative, legal, and professional setting. It is important that the resident is aware of these elements.

The responsibility for determining when an autopsy will be performed is that of the staff consultant assigned to the autopsy service for that day. PGY1 residents must be directly supervised during their first three autopsies by faculty, PGY3 or PGY4 residents, or the designated pathology assistant. In general, autopsies are performed Monday through Friday. Saturday cases may take place at the discretion of the autopsy director in certain situations. Forensic autopsies are not performed at the University of Kansas Hospital. Residents attain forensic experience during the forensic rotation at the Jackson County Medical Examiner’s office.

<table>
<thead>
<tr>
<th>Legend for Learning Activities for Residents</th>
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<tbody>
<tr>
<td>Didactic lecture</td>
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<tr>
<td>Faculty sign-out</td>
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<tr>
<td>Journal club</td>
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<tr>
<td>Directly supervised procedure</td>
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<td>Role modeling</td>
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<td>Lab inspections</td>
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<tr>
<td>Interdisciplinary conference</td>
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<td>Online tools</td>
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<td>Unknown slide conferences</td>
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<td>Project</td>
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<tr>
<th>Legend for Evaluation Methods for Residents</th>
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<tbody>
<tr>
<td>Report review</td>
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<tr>
<td>Direct observation</td>
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<tr>
<td>Checklist</td>
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<tr>
<td>Global rating/faculty evaluation</td>
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<tr>
<td>Standardized exam</td>
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<tr>
<td>Practical slide exam</td>
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<tr>
<td>In-house written exam</td>
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<tr>
<td>360 multisource rating</td>
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<tr>
<td>Portfolios</td>
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<tr>
<td>Procedures and case logs</td>
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128
KUMC Pathology Residency Manual
## AUTOPSY SERVICE CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must become competent in performing medical autopsies.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate competent autopsy prosection using routine techniques,</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, CL, GR/FE, PF</td>
</tr>
<tr>
<td>completing gross examination in a period of three hours for uncomplicated</td>
<td></td>
<td></td>
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<tr>
<td>cases, or four hours for complicated cases.</td>
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<tr>
<td>Perform at least one adult and one pediatric autopsy under indirect</td>
<td>DL, FSO, DSP, RM,</td>
<td>RR, DO, GR/FE, SE, IWE</td>
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<tr>
<td>supervision (with the assistance of dieners and/or pathology assistants).</td>
<td>RM, IC</td>
<td></td>
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<tr>
<td>Demonstrate knowledge of modified autopsy techniques such as Letulle/Rokitansky-style organ removal and other en bloc dissections.</td>
<td>DL, FSO, JC, DSP,</td>
<td>RR, DO, CL, GR/FE, SE,</td>
</tr>
<tr>
<td></td>
<td>RM, OT, USC</td>
<td>PSE, IWE, PF, PCL</td>
</tr>
<tr>
<td>Demonstrate ability to remove the brain without causing injury to the</td>
<td>DL, FSO, JC, DSP,</td>
<td>RR, DO, GR/FE, SE,</td>
</tr>
<tr>
<td>structure.</td>
<td>RM, IC, OT</td>
<td>IWE, PF, PCL</td>
</tr>
<tr>
<td>Demonstrate ability to examine leg veins, bones and joints.</td>
<td>DL, FSO, JC, DSP,</td>
<td>DO, GR/FE, SE, IWE,</td>
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<tr>
<td></td>
<td>RM, IC, OT</td>
<td>PCL</td>
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<tr>
<td>Demonstrate ability to identify those cases for which blood samples and</td>
<td>DL, FSO, JC, RM,</td>
<td>DO, GR/FE, PCL</td>
</tr>
<tr>
<td>vitreous eye fluid are required for biochemical tests, and to collect those</td>
<td>IC, OT</td>
<td></td>
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<tr>
<td>samples in the proper fashion.</td>
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</table>

## AUTOPSY SERVICE CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Residents must demonstrate competency in basic skills in anatomic pathology.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Show the ability to correctly describe common abnormalities of diseased</td>
<td>DL, FSO, JC, DSP,</td>
<td>RR, DO, CL, GR/FE,</td>
</tr>
<tr>
<td>organs by gross and microscopic examination, including congenital,</td>
<td>RM, IC, OT, USC, P</td>
<td>SE, PSE, IWE</td>
</tr>
<tr>
<td>degenerative, inflammatory, neoplastic, and autoimmune disorders.</td>
<td></td>
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</tbody>
</table>
Demonstrate ability to describe those circumstances in which specimens (fluids or tissues) should be kept for toxicological studies, and knowledge of how to do so.

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>DL, FSO, JC, DSP, IC, OT, USC</td>
<td>RR, DO, CL, GR/FE, SE, PSE, IWE, PF, PCL</td>
</tr>
</tbody>
</table>

**AUTOPSY SERVICE CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**
*Residents must become competent in performing medical autopsies.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate an ability to compose a provisional anatomic diagnostic report of autopsy findings within 24 hours of completing the postmortem examination.</td>
<td>FSO, RM, DSP</td>
<td>RR, DO, GR/FE, PCL</td>
</tr>
<tr>
<td>Demonstrate an ability to compose a final autopsy report within 30 days of completing the postmortem examination.</td>
<td>FSO, RM, DSP</td>
<td>RR, DO, GR/FE, PCL</td>
</tr>
<tr>
<td>Demonstrate an ability to assist Autopsy I residents in the achievement of basic skills in anatomic pathology.</td>
<td>FSO, RM, DSP</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Take selective autopsy call in support of Autopsy I residents.</td>
<td>FSO, RM, DSP</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

**AUTOPSY SERVICE CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS**

**Goal:**
*Residents must become competent in writing autopsy reports and communicating results with medical staff and families.*

<table>
<thead>
<tr>
<th>Objectives:</th>
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</thead>
<tbody>
<tr>
<td>Demonstrate an ability to compose a provisional anatomic diagnostic report of autopsy findings within 24 hours of completing the postmortem examination.</td>
<td>FSO, DSP RM</td>
<td>RR, GR/FE, DO, PCL</td>
</tr>
<tr>
<td>Demonstrate an ability to compose a final autopsy report according to an approved format &amp; within 30 days of completing the postmortem examination, including accurate and complete anatomic diagnoses, thorough gross and microscopic descriptions, and pertinent clinical-pathologic correlations and mechanistic interpretations.</td>
<td>FSO, DSP RM</td>
<td>RR, GR/FE, DO, PCL</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC, USC</td>
<td>DO, GR/FE, PSE</td>
</tr>
</tbody>
</table>
## AUTOPSY SERVICE CORE COMPETENCY: PROFESSIONALISM

**Goal:**

_Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population._

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
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</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE, 360</td>
</tr>
</tbody>
</table>

## AUTOPSY SERVICE CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**

Residents must become familiar with all regulations associated with the autopsy service.

<table>
<thead>
<tr>
<th>Objective:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate familiarity with the laws regarding permission for autopsy and the classification of those autopsies requiring medico-legal status.</td>
<td>FSO, RM, DSP, IC</td>
<td>DO, GR/FE, IWE</td>
</tr>
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</table>

### PGY 1 GOALS: By the end of the first year:

- The resident has completed their first three autopsies under direct supervision.
- The resident demonstrates a strong knowledge in gross and microscopic anatomy.
- The resident understands and applies the seven components of the autopsy, as appropriate, required by ACGME for participation credit.
- The resident demonstrates the ability to review and extract appropriate information from the clinical history prior to the autopsy.
- The resident recognizes indications for performing an autopsy and recognizes settings where the coroner should be contacted.
- The resident can independently perform a full autopsy, including removal of the brain, and be able to correlate gross and microscopic findings with clinical history.
PGY 2 GOALS: By the end of the second year:

- The resident should be able to perform autopsies independently and efficiently with minimal correction by the attending.
- The resident should be able to prepare and discuss the autopsy findings at morbidity and mortality conferences, including preparation of gross and microscopic photographs.
- The resident can properly dissect the brain for gross examination independently, identify the majority of gross and microscopic neuropathology.
- The resident is ready to start supervising (in third year) junior residents in autopsy procedures.

PGY 3 GOALS: By the end of the third year:

- The resident should have completed a rotation at the Jackson County Medical Examiner's Office and be competent in general forensic autopsy skills.
- The resident should have case logs of a minimum of 50 autopsies, of which all 50 autopsies can be shared with one other resident. All seven elements must be documented, except forensic cases where microscopic sections are taken only when deemed necessary.

PGY 4 GOALS:

- The resident should have completed all autopsy training, including the Jackson County Medical Examiner's Office rotation.
- The resident should have minimally fifty (50) autopsies with gross and microscopic examination (see Year 3 goals; microscopics are taken as indicated on forensic cases).
- The resident should be competent in all Autopsy Skill Levels 1 and 2.

GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice-Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on page 19-20.

RECOMMENDED READING LIST:

CASE ASSIGNMENT

- The Day 2 surgical pathology resident is the primary resident on the autopsy case and will be responsible for the autopsy report and the sign-out of the case.

- All autopsy cases will be shared based upon the autopsy sharing schedule (priority should be given to the resident with less than 50 autopsies).

- A senior resident (3rd or 4th year) will be pre-assigned as the supervising resident for all 1st years for their first three autopsies. The supervising resident can also be considered the sharing resident.

- Pathology assistants will be present for most cases and may also act as supervisors for 1st year residents.

SIGNING OF DEATH CERTIFICATE

The death certificate should be signed by the responsible pathologist or coroner; most are signed online through the state. **Under no circumstances should the resident prosector sign the death certificate.** Residents are encouraged to observe the attending pathologist's process in entering and signing the death certificate.

AUTOPSY CALL SCHEDULE

The autopsy call and sharing schedules are prepared by the Chief Residents and copies are given to all residents, staff, morticians, and office personnel. From 8 a.m. to 4 p.m. each weekday, Monday through Friday, autopsies are performed by prosectors as assigned. On Saturdays, the autopsy technician will contact the on-call resident if there is a scheduled case. Autopsies are not performed on Sundays.

AUTOPSY PERMITS

The right to authorize the performance of an autopsy is guided by the law of the State of Kansas. In case of doubt, contact KUMC’s Office of the Legal Counsel [http://www2.kumc.edu/directory/DepartmentDetails.aspx?Id=31](http://www2.kumc.edu/directory/DepartmentDetails.aspx?Id=31). Generally, the autopsy technician and senior staff will be able to resolve the issue.

The vital statistics of an autopsy permit are filled out by nursing personnel and is then signed by the legally responsible person and witnessed and countersigned by a hospital physician or nurse. A duplicate copy of this permit is made and subsequently filed with the departmental autopsy record. A complete autopsy will ordinarily include examination of the brain, organs of the neck, and contents of the thoracic, abdominal, and pelvic cavities. This should be stated or implied in the permit. The autopsy permit should be coextensive with the autopsy to be performed. It should also contain permission to retain any part or organ for future study. In requesting permission for autopsy, the nature and the extent of the autopsy must not be misrepresented.

An autopsy permit may legally limit the extent and specify the manner of performance of an autopsy in any way the person signing the permit demands. Under no circumstances should the resident exceed the limit of the permission. Residents must adhere strictly to wishes of the party who authorized the postmortem examination.
Be certain that the body is identified properly and that the resident is performing the autopsy on the body for which there is legal permission. After reviewing the permit for restrictions, and prior to starting the external exam, a "time-out" procedure is done whereby the resident and the technician ensure the personal identifiers on both the paperwork and the identification bracelet of the decedent match.

It is important that the face, hands, and other exposed areas not be mutilated. This should not restrict the performance of an autopsy or serve as an excuse for omitting necessary, even though unusual, procedures in a given case. Proper restorative measures, however, must be taken. Be sure that these measures can be accomplished before the resident makes any incisions.

**NOTIFICATION OF CLINICIANS OF AUTOPSIES**

The autopsy technician or resident will notify the attending clinicians, if requested on the consultation form, when the autopsy is to start or is available for review. Some clinicians may wish to observe the dissection of certain organs. Please accommodate them. In case the responsible attending physicians are unable to attend the autopsy, the resident prosector or pathology senior staff should notify them of the autopsy findings following completion of the gross dissection.

**NOTIFICATION OF NEUROPATHOLOGIST OF AUTOPSIES**

The Day 2 resident will notify the neuropathologist of every autopsy case prior to the procedure, and together with the attending pathologist, develop a plan for examination of the brain and other organs such as spinal cord, muscle, or peripheral nerves for each case.

**ATTENDANCE AT AUTOPSIES**

In general, members of the senior and housestaff, visiting physicians and medical and other allied health students are admitted in the autopsy room, with the permission of the attending pathologist. It is within the legal right of persons authorizing permission for an autopsy to limit the attendance at that autopsy and to restrict the conditions under which it can be performed. The attending pathologist can restrict the autopsy to a private autopsy if desired and specifically state who may or may not attend. These are binding commitments. No lay people will be permitted in the autopsy room except by special permission from the Director of Autopsy Service and only after clearance from hospital risk management.

**OUTSIDE INQUIRIES CONCERNING AUTOPSY FINDINGS**

Phone calls from relatives are to be referred to the staff physician. Inquiries by lawyers on matters relating to subpoenaed autopsy findings and reports in court or before a grand jury should be discussed first with the pathologist or the Director of Autopsy Service. Any inquiries from local newspapers should be referred to the media relations department. Any request for information regarding coroner’s cases must be referred to the respective coroner’s office.

**ORGAN AND TISSUE DONATION**

Midwest Transplant Network is notified of every hospital death and approaches the family if appropriate for any tissue or organ donation.
AUTOPSIES REQUESTED BY OUTSIDE AGENCIES

In general, outside autopsies (which personnel of this department may be called upon to perform) may be from the following sources:

- Private autopsies may be performed on special arrangement by the Director of Autopsy Service.
- Brain bank and special CNS-related autopsies under the auspices of Dr. Newell.
- All outside autopsies will be transported to and performed at the hospital. Transportation will be paid by the outside party or agency.

RESEARCH TISSUES FOR OTHER DEPARTMENTS

Other departments in the hospital may ask that certain tissue specimens be kept. Requesting departments must fill out a tissue resource request form, available from Dr. Kathy Newell, director of KUMC’s Brain and Tissue Bank. All requests will be reviewed and cleared by Dr. Newell.

DEPARTMENTAL AUTOPSY SERVICE CLERICAL POLICIES

The Provisional Anatomical Diagnosis (PAD) must be submitted to the autopsy secretary via email within 24 hours of the autopsy. The secretary will enter the PAD into CoPath for the attending pathologist to edit and sign.

FINAL AUTOPSY REPORT

The sharing resident is responsible for entering all external exam, gross exam findings, and microscopic slide key into CoPath. The Day 2 surgical pathology resident is responsible for entering the Final Autopsy Diagnosis (FAD), microscopic descriptions, and the clinical summary and opinion. The FAD should follow the same order as the PAD as much as possible, with additions or adjustments made depending on microscopic exam or other findings. The final report may be sent to the department transcriptionist for proofreading services before sign-out. The final report must be signed out by the attending pathologist in CoPath within 30 working days. In some cases, addendums may be added later.

LATE LIST

The “Autopsy Pending List” is distributed weekly via email. Autopsies not completed in 30 working days will be deemed late. Autopsies will be deleted from the list after final signature in CoPath by the attending pathologist.

CODING

Autopsy diagnoses should be SNOMED coded into CoPath.

USE OF “STOCK JAR” FOR AUTOPSY

The plastic containers with screw cap lids are used to store small, representative portions of autopsy tissues and organs for prolonged periods of time (three years is required by Kansas law for coroner cases). These representative samples of tissues are then available for review at a later date. At the time of the autopsy, place small representative portions of the organs and tissues into the plastic container. The pieces should generally not exceed two centimeters in greatest dimension. Tissue samples must be thin enough so that they will fix properly.
In most cases, samples of tissues to be kept include:

- Brain
- Heart-LV, RV, septum, conduction system if appropriate.
- Aorta
- Five lung lobes
- Kidneys
- Adrenals
- Lymph nodes
- Spleen
- Bone marrow if pertinent.
- Liver
- Gallbladder if pertinent.
- Appendix if pertinent.
- Pancreas
- Urinary bladder
- Sex organs-ovaries, testes, prostate, uterus, vagina, Fallopian tubes.
- Thyroid
- Pituitary
- Esophagus, stomach, small and large bowel if pertinent.
- AND REPRESENTATIVE PORTIONS OF ALL PATHOLOGY OR OTHER TISSUES DEEMED PERTINENT TO A PARTICULAR CASE.

GENERAL PLAN OF THE AUTOPSY STUDIES-OBJECT AND SCOPE OF THE AUTOPSY

The autopsy is a scientific examination of the body to determine the pathologic processes present and their relation to clinical phenomena and history, to determine the causes of the pathological processes, and to acquire information regarding the processes and nature of disease and injury. The more effectively these ends are accomplished, the greater will be the contribution of the autopsy to the sum of knowledge concerning the disease or injury from which the patient died and thereby to clinical medicine, to public health and to the interest of the family of the deceased.

AUTOPSIES ARE PERFORMED FOR:

- Teaching and training.
- Discovery of new diseases and pathogenetic mechanisms.
- Evaluation of treatment - medical and surgical.
- Family benefits.
- Public health.
- Socioeconomic.
- Vital statistics.
- Medicolegal reasons.
The autopsy should cover not only those structures which are at the seat of obvious alteration, but all organs of the body because the normality of certain viscera is often quite as significant as the disease of others. Additionally, organs that appear normal macroscopically are frequently abnormal microscopically. The gross examination is amplified by microscopic studies, and possibly bacteriological, viral, toxicological, molecular examinations and such other investigations as may be indicated.

*The autopsy record embodies the results of the only complete examination the patient ever had.* For this reason, findings which may have little significance in the last or main illness take on some importance and deserve to be recorded. The evidences of disease produced by an autopsy are direct and objective. The findings are those of the lesion itself and not only of some disturbance which results secondarily from the presence of a lesion.

Autopsy records are a much-used source of statistical data relating to diseases. Rarely are these statistical studies made by the author of the protocols. For this reason, clear, concise language and completeness of records is imperative. Typographical errors in the protocol become as confusing or misleading to the reader as misstatements of fact. Protocols should be completed using proper grammar and English, as though they were being prepared for publication.

**PROVISIONAL ANATOMICAL DIAGNOSIS (PAD)**

After the pathologist has examined the organs in the autopsy room with the prosector and discussed the provisional anatomical diagnosis, the resident will email the autopsy secretary the PAD to enter into CoPath. The staff will edit as necessary and sign the PAD in CoPath within 24 hours, which will then be available in the hospital electronic medical record.

**FORM OF THE REPORT**

- Anatomical diagnoses.
- Cause of death.
- Prosector's comments and opinion.
- Gross autopsy findings.
- Brain after fixation.
- Microscopic findings.
- Postmortem laboratory findings.
- Diagrams, if any.
- Gross photographs.
- Clinical summary.

**Anatomical Diagnosis**

The final report begins with a list of diagnoses. Where applicable include in the list of diagnoses "clinical history of__ _______________ " either as a separate item, or in parenthesis, after a pertinent anatomical finding. The anatomical diagnoses should be as complete as possible. List the lesion first and the structure next (example: adenocarcinoma, right main bronchus). Diagnostic terms should be as specific as possible and yet be general pathological terms (for example: "arteriosclerosis, cerebral arteries" rather than "cerebral sclerosis"). Specifying the exact site is very important.
There are many situations in which the whole course of a disease depends upon a relatively innocuous lesion being located in a particular site. For example, subcutaneous abscess is not always a particularly serious lesion. A subcutaneous abscess of the upper lip, however, carries considerably more danger because of its location.

**In the diagnoses the order should be as follows:**

1. Disease process (noun).
2. Organ, tissue or cells.
3. Modifier (e.g. acute, massive, etc.).

The list of diagnoses should be as complete as possible, but should not include abnormalities of no significance. Amputated phalanges, tattoos, old operative scars, absence of teeth, etc., which bear no relation to the case may not be mentioned in the diagnosis. They should, however, be described in the protocol.

With regard to the major diagnoses, the first diagnosis should always be the fundamental disease, and should be similar to the wording on the death certificate.

**First Diagnosis Example**

Acute gangrenous appendicitis, with:
- Appendiceal abscess.
- Thrombosis of appendiceal vein.
- Pylephlebitis.
- Multiple liver abscesses.

Other diagnoses should include any other concomitant conditions such as hyperplasia of prostate, arteriosclerosis, etc. These diagnoses should be arranged in order of importance. Each should be followed by conditions which may have been secondary to the diagnoses. This arrangement of specific terms and specific sites will give the reader at a glance a fairly good summary of the patient's illness and death. Do not include any descriptions in the anatomical diagnoses. Where there have been surgical pathology specimens, they should be cross referenced in the diagnosis, the accession number given, described in the autopsy protocol where applicable, and duplicate slides filed with the autopsy slides. Do not append the surgical pathology report.

The Provisional Anatomical Diagnosis will, in most cases, be modified considerably in the light of subsequent studies and should be re-worked thoroughly to result in the Final Anatomical Diagnosis before the case is completed.

**Opinion**

Use the opinion section of the report to synthesize the clinical and pathologic findings. The first paragraph should be composed of a brief clinical summary ending with the time and date of death. Since a diagnosis sheet is present elsewhere in the autopsy protocol, a re-listing of diagnoses is inadvisable. An opinion can be based on a clinical problem list, or better, one that the pathologist generates after reading the chart and performing the gross autopsy. This should lead to a concise (usually one page) opinion, which should be principally a clinical-pathological correlation, discussion of significance of the principal findings.
Surprises, fulfilled predictions, interesting or unusual conditions, and significant negative findings should be mentioned. The opinion should include a statement as to the manner (all hospital cases should be natural) and final cause of death. References may be included to support unusual findings. Remember that all statements made in the opinion can be used directly in court. The opinion is not the place to criticize the clinical care, yet the autopsy still remains the best instrument of quality control. Use good judgment and common sense.

**Gross and Microscopic Description**

Because data is obtained essentially at two different times, two separate descriptions are written. The first embodies the data obtained from the clinical history, the gross examination and such chemical, bacteriological or frozen section data as are available at the completion of gross dissection. The gross description should be entered into CoPath by the sharing resident shortly after the gross dissection. The most complete and accurate description is made at the time of dissection, not by relying on memory afterwards. The second is written after the microscopic examination and all chemical, bacteriological, viral, histochemical and other studies have been completed. The first description is made to preserve the detailed data of the clinical record and gross examination until the final report can be written, and is used in selecting case material and specimens for conferences, classes, etc. Pictures or diagrams may be used as an adjunct to the written description.

**Autopsy Photographs**

Digital photographs should be taken at the time of autopsy of the overall body, the facial features, injuries, and significant anatomic abnormalities. These are not only used to document lesions, but will be used for CPC’s and numerous teaching purposes. The autopsy number and a ruler should be included in each photo. Copies of photos needed for presentations may be requested from the autopsy technicians via email.

**Clinical Summary**

The clinical summary should generally be one to three paragraphs in length, depending on the complexity of the case and length of hospitalization and should include dates, laboratory data, opinions of various examiners, results of special examinations and any other relevant data. The clinical summary should be chronological and should include clinical opinions and diagnoses. It should also include certain calendar dates - date of onset of the important illness, date of admission to the hospital, date of operation (for that illness), and date and time of death.

Other temporal relationships should be established with these dates. Important incidents in the past history (before the present illness) may be referred to by date. Events in the last illness, however, should be tied together by a few reference points and liberal use of days, weeks, months, years, etc. Example: Third postoperative day, sixth week of hospitalization, tenth year of disease, etc. The resident need not catalog every symptom so that the reader views the patient’s story exactly as the clinician did. It is necessary to distinguish important from unimportant therapeutic measures.

Generally it is necessary to state if the patient was treated with antibiotics, with transfusions, antimetabolites, diuretics, corticosteroids, etc., inclusive of the compound involved and the dose and duration of treatment may be mentioned. A chronologic appendix may be useful in complex cases. Supportive treatment such as vitamins, sedatives or tranquilizers need not be listed specifically. The dose and extent of radiation therapy should be given.

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*KUMC Pathology Residency Manual*
FINAL REPORTS

- The FAD, clinical summary, opinion, and microscopic description should be entered into CoPath by the Day 3 surgical pathology resident.
- The following procedure should be used in the performance and completion of all autopsy reports within 30 working days.
  - Attending staff will edit and notify the department transcriptionist if proofreading is requested.
  - Attending staff will sign-out case in CoPath.
  - Neuropathology reports may be done as addendums if not complete at the time of signing.
  - Slides will be returned to autopsy technicians for filing.
    - Paperwork (copy of permit, gross and external exam forms) will be returned by the resident to Linda Riley for filing.
The four-week rotation at the Jackson County Medical Examiner’s Office comprises a working exposure to the operation of an urban medical examiner’s office. The pathology resident should be able to perform a competent medicolegal examination and document the findings with sufficient detail to determine the cause and manner of death.

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</table>
### Forensic Pathology Core Competency: Patient Care

**Goal:**
*Residents should become competent in performing forensic autopsies.*

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<thead>
<tr>
<th>Objectives</th>
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<tbody>
<tr>
<td>Perform a competent medicolegal autopsy.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Produce an autopsy report of the examination to determine the cause and manner of death.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
</tbody>
</table>

### Forensic Pathology Core Competency: Medical Knowledge

**Goal:**
*Residents must demonstrate competency in basic skills in forensic pathology.*

<table>
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<tr>
<th>Objectives</th>
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<tbody>
<tr>
<td>Recognize and describe postmortem changes.</td>
<td>FSO, DSP, RM, DL</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Explain the physical principals of trauma relating to blunt force injury and gunshot wounds.</td>
<td>FSO, DSP, RM, DL</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Distinguish various types of injuries due to blunt force trauma, sharp trauma, gunshot wounds and automobile collisions both in medic and in layman's terms.</td>
<td>FSO, DSP, RM, DL</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>List the types of asphyxia deaths and give examples of pertinent findings.</td>
<td>FSO, DSP, RM, DL</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to describe those circumstances in which specimens (fluids or tissues) should be collected for toxicological studies in drug abuse and overdose cases.</td>
<td>DL, FSO, RM, DSP</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Generate a sufficiently detailed differential diagnosis for the sudden unexpected death of an infant, child and adult.</td>
<td>FSO, DSP, RM, DL</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
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### Forensic Pathology Core Competency: Interpersonal & Communication Skills

**Goal:**
*Residents must become competent in writing autopsy reports and communicating results.*

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<tr>
<th>Objectives</th>
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<tbody>
<tr>
<td>Demonstrate an ability to compose a provisional anatomic diagnostic report of autopsy findings within 24 hours of completing the postmortem examination.</td>
<td>FSO, DSP, RM, DL</td>
<td>RR, DO, GR/FE</td>
</tr>
</tbody>
</table>
Demonstrate an ability to compose a final autopsy report according to an approved format and within 30 days of completing the postmortem examination, including accurate and complete anatomic diagnoses, thorough gross and microscopic descriptions, and pertinent clinical-pathologic correlations and mechanistic interpretations.

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<th>FORENSIC PATHOLOGY CORE COMPETENCY: PROFESSIONALISM</th>
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<td><strong>Goal:</strong> Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.</td>
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<td>Demonstrate compassion: be understanding and respectful of decedents, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM</td>
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<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>FSO, RM</td>
<td>DO, GR/FE</td>
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<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, FSO, DSP, RM</td>
<td>DO, GR/FE</td>
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<td>Demonstrate interpersonal skills in functioning as a member of a forensic medicolegal team.</td>
<td>FSO, DSP, RM</td>
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<th>FORENSIC PATHOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE</th>
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<td><strong>Goal:</strong> Residents must become familiar with all regulations associated with the forensic autopsy.</td>
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<tr>
<td>Demonstrate familiarity with the laws regarding permission for autopsy and the classification of those autopsies requiring medico-legal status.</td>
<td>FSO, DSP, RM, DL</td>
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**DUTIES AND RESPONSIBILITIES OF THE RESIDENT**

The resident shall be supervised by the medical examiner faculty in their participation in forensic autopsies. The resident will dictate autopsies in which they are the primary prosecutor within 24 hours. Reports will be reviewed and edited by the supervising forensic pathologist. The resident shall follow up on microscopic, radiographic and toxicological examinations, and observe court proceedings. This responsibility and accountability is further described in the “Resident Agreement with JCMEO.” The ultimate responsibility for the autopsy and autopsy reports shall be by the forensic pathologist.
**RESIDENT EVALUATION**

Residents shall at all times be personally supervised by the forensic pathologist. Evaluation will be standard evaluation form.

**FORENSIC PATHOLOGY EDUCATIONAL PROGRAM**

**Overall Educational Goals**

Competencies that are common to all rotations are outlined below. Competencies that are specific to individual rotations are included with each sub-discipline. Residents will be given graduated responsibilities and will be evaluated at two, general skill levels. Specific goals and learning objectives (Skill Levels) are described under each sub-discipline.

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Residency training in transfusion medicine prepares the resident to provide laboratory and clinical transfusion services at community and tertiary care hospitals as well as medical school based tertiary care centers. It also prepares the resident for additional training in careers specializing in clinical transfusion medicine, transfusion medicine research, blood center operations, histocompatibility hematopoietic stem cell transplants or organ banking.

After the training period, the resident will be knowledgeable and experienced with red cell antigens, compatibility testing, component therapy, adverse effects of transfusion, transfusion appropriateness review, blood donor evaluation and donation, therapeutic apheresis, peripheral blood stem cell collection, stem cell processing and infusion. Transfusion medicine clinical training and experience is provided at two major teaching sites.

The University of Kansas Medical Center is an academic medical center with major programs in organ and stem cell transplantation, oncology, cardiovascular surgery, obstetrics and neonatal ICU. The Community Blood Center of Kansas City is a state-of-the-art blood collection and processing center, which provides apheresis services and conducts both clinical and basic science research related to transfusion medicine. The resident will take a two-week bench-oriented course, as an introduction to the science of transfusion medicine as part of their training.

**Supervisory Guidelines for Patient Care**

The resident is directly supervised by a senior staff attending physician. The attending physician spends one to three hours each week day with the resident. The resident carries out the provision of clinical blood bank services but with close overview. The attending physician also sees therapeutic apheresis patients and inpatient consults and countersigns notes made by the resident in the patient's medical record (consults, stem cell infusions, therapeutic procedures).

Individual decisions to approve components not meeting guidelines and deviations from blood bank procedures (e.g. release incompatible blood), are reviewed by the attending physician not only for supervision and teaching purposes but also for attending physician billing documentation. In addition to daily mentoring by the attending physician, the resident receives scheduled didactic learning sessions (one-on-one) from the physician in charge of the apheresis/donor center, the blood bank laboratory and the cell therapy laboratory.
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### TRANSFUSION MEDICINE CORE COMPETENCY: PATIENT CARE

**Goal:**
Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of transfusion medicine services.

#### Objectives:

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<tr>
<td>Gathers essential and accurate information about patients using all relevant available modalities.</td>
<td><strong>DSP, RM, IC</strong>&lt;br&gt;<strong>DO, GR/FE</strong></td>
</tr>
<tr>
<td>Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.</td>
<td><strong>DL, FSO, DSP, RM, IC</strong>&lt;br&gt;<strong>DO, GR/FE, SE, IWE</strong></td>
</tr>
<tr>
<td>Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of transfusion medicine.</td>
<td><strong>DL, JC, DSP, RM</strong>&lt;br&gt;<strong>DO, GR/FE, SE, IWE</strong></td>
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Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity. Where clinically appropriate, consult on the use of laboratory-based therapeutics such as blood transfusion and other forms of cellular therapy.

Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.

Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.

**TRANSFUSION MEDICINE CORE COMPETENCY: MEDICAL KNOWLEDGE**

**Goal:**

*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to transfusion medicine.*

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<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in transfusion medicine.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.</td>
<td>DL, JC, RM</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
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<td>Recognize the unique aspects of transfusion medicine practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
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<td>Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations.</td>
<td>DL, RM, LI</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations.</td>
<td>DL, RM, LI, OT</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice.</td>
<td>JC, DSP, RM, P</td>
<td>DO, GR/FE, SE, IWE</td>
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Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting.

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<th>TRANSFUSION MEDICINE CORE COMPETENCY: PRACTICE-BASED LEARNING &amp; IMPROVEMENT</th>
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<tr>
<td><strong>Goal:</strong> <strong>Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.</strong></td>
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<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>RM, LI</td>
<td>DO, GR/FE</td>
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<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT</td>
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<td><strong>Goal:</strong> <strong>Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.</strong></td>
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<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the transfusion medicine service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
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</table>
Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate. | RM, IC | RR, DO, GR/FE

Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to transfusion medicine. | DSP, RM | DO, GR/FE

Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge. | JC, RM, IC | DO, GR/FE

| **TRANSFUSION MEDICINE CORE COMPETENCY: PROFESSIONALISM** |
| **Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.* |

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<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>RM</td>
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<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>DSP, RM</td>
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<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
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</tbody>
</table>

**TRANSFUSION MEDICINE CORE COMPETENCY: SYSTEM-BASED PRACTICE**

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of transfusion medicine in the healthcare system.</td>
<td>DL, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
<td>RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.</td>
<td>DL, FSO, LI, OT</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Understand and implement policies to continually improve patient safety as they relate to transfusion medicine.</td>
<td>FSO, RM, LI, P</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
ADDITIONAL OBJECTIVES SPECIFIC TO TRANSFUSION MEDICINE

[Pathology Milestones]
PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice
For expanded definitions, see pages 30-31.

General Transfusion Medicine Objectives:

Interacts with physicians to ensure that blood components are used appropriately balancing risk, benefit and availability. [PC, MK, PBL, ICS, PRO, SBP]

- Evaluate transfusions and transfusion requests that are out of established guidelines.
- Approve initial requests for HLA-matched donors or crossmatch-negative platelets.
- Advise about difficult crossmatches, antibody problems and selection of compatible red cells.
- When fully compatible red cells are unavailable, approves release of least-incompatible blood and assures that clinician is aware of risk for a risk-benefit analysis.
- Approves use of granulocyte transfusion and ensures availability of high-dose (G-CSF) granulocytes when needed.

Provide medical direction to ensure an adequate supply of blood components. [PC, MK, PBL, ICS, PRO, SBP]

- Make decisions for the donor center regarding blood donors with medical suitability conditions out of established guidelines.
- Provides medical management of donors experiencing adverse reactions and complications due to blood donation.
- Provide peripheral blood stem cell collections.
- Evaluate requests, suitability of patient, write orders.
- Manage blood shortages (ABO, Rh switches).
- Interacts with clinicians using stem cell components particularly if there are positive bacterial culture results or if cell dose is out of desired range.
- Approves directed platelet donors.
- Participates in College of American Pathologists and American Association of Blood Bank annual inspections.
Provide transfusion-related services directly to patients. [PC, MK, ICS, PRO]

- Provide therapeutic apheresis services (plasma exchange, cytapheresis, SPA column, photopheresis, red cell exchange).

**Immunohematology Objectives for Community Blood Center:**

The resident will be able to explain and perform basic tests, solve antibody problems and correlate laboratory findings with clinical significance in the following areas. [PC, MK, PBL, ICS]

**Blood Groups**
- ABO and other carbohydrate antigens (Lewis, P).
- Rh and other protein antigens (Kell, Duffy, Kidd, MNS).
- Inheritance.
- Clinical significance for transfusion or pregnancy.
- Resolution of typing problems.

**Pre-transfusion Testing**
- Type and screen.
- Compatibility tests including a complete crossmatch through the antiglobulin phase using both tube and gel technologies.
- IgM and IgG antibodies.
- Mechanisms of immune hemolysis.
- Direct and indirect antiglobulin testing.
- Alloantibody identification using panels.
- Enhancement media including low ionic strength (LISS), enzyme, albumin and polyethylene glycol (PEG).
- Testing before non-red cell transfusion.
- Role of the computer crossmatch and electronic crossmatch.

**Red Cell Serological Studies**
- Autoantibody evaluation, cold and warm.
- Autoimmune hemolytic anemias – treatment and transfusion.
- Drug-related antibodies – evaluation and mechanisms.
- Elution – purpose, methods, and interpretation.
- Complex antibody problems – i.e., multiple, high frequency antigens, selection of appropriate components.
- Prenatal screening and intra-partum monitoring.
- Evaluation of hemolytic diseases of the newborn.
- Fetal maternal hemorrhage, Rh immune globulin dose.
- Exchange transfusion and/or intrauterine transfusion.
Platelet Serology
- Platelet antigen groups.
- Drug-dependent platelet antibodies.
- Autoimmune thrombocytopenia.
- Neonatal allo-immune thrombocytopenia.
- Post transfusion purpura.
- Platelet antibody testing including platelet immunofluorescence and antigen capture assays (MAIPA).

Neutrophil Serology
- Neutrophil-specific antigens.
- Autoimmune neutropenia.
- Drug-dependent neutrophil antibodies.
- Neutrophil serology including immunofluorescence and agglutination assays.

Whole Blood Collection Objectives at Community Blood Center:

The resident will be able to provide medical assistance to a blood donation program. A basic understanding of the following topics will be obtained. [PC, MK, PBL, ICS, PRO, SBP]

Blood Donation
- Performance of donor interview and physical examination.
- Donor safety issues.
- Recipient safety issues.
- Phlebotomy process.
- Evaluation and treatment of donor reactions and injuries.
- Autologous donation – candidates, frequency, common problem.
- Directed donation – pros and cons of a directed donor program.

Blood Component Preparation and Testing
- Component preparation including labeling.
- Donor testing:
  - Blood groups – ABO, Rh, antibody screen.
  - Infectious disease marker testing: syphilis, HBsAg, anti-HIV, ALT, anti-HBc, anti-HTLV, anti-HCV.
  - Confirmatory testing including anti-HIV western blots, anti-HCV RIBA, and anti-HTLV western blot.
  - CMV antibody testing.
  - Nucleic acid amplification testing (NAT) for Hep C, HIV, and West Nile virus.
- Observe and understand Leukoreduction of red cells and apheresis platelets. Know the benefits of leukocyte reduction for the recipient.
**Blood Donor Counseling** [PC, MK, ICS, PRO, SBP]

- Donor notification of positive results.
- Confidentiality and requirements for reporting infectious disease markers to public health agencies.

**Clinical Transfusion Service Objectives (KUMC)**

The resident should be able to discuss standard laboratory and clinical practices, investigate problems and questions, and assess transfusion therapy in the following areas. [PC, MK, PBL, ICS, PRO, SBP]

**Blood Component Therapy**

- Blood administration – specimen and patient identification, thawing, pooling, issuing, transport time and temperature, filtration.
- Blood components – red blood cells, platelet concentrates, apheresis platelets, plasma, cryopoor plasma, cryoprecipitate, and granulocytes.
- Blood appropriateness review – audit requirements, methods, transfusion committee activities.
- Modified components – washed, irradiated, leukocyte-reduced, frozen/thawed deglycerolized red cells, HLA-matched platelets, crossmatched platelets.
- Support for solid organ and bone marrow transplant patients.

**Adverse Effects of Transfusion**

The resident will:

i. Recognize and discuss pathophysiology, treatment, and prevention of the following: febrile, allergic, acute hemolytic, delayed hemolytic, anaphylactic, septic, transfusion-associated graft-versus-host disease, transfusion-related acute lung injury, and circulatory overload. [PC, MK, SBP]

ii. Manage massive transfusion including the appropriate and timely ordering of laboratory tests for monitoring the patient and recognizing the metabolic and hemostatic consequences of massive transfusion. [PC, MK, SBP]

iii. Have a working knowledge of the effects of stored blood on recipients, including pediatric patients. [PC, MK, SBP]

iv. **Transfusion-Related Infections**

   - HIV, hepatitis B and C, HTLV, CMV, West Nile Virus, protozoa, and bacterial contaminants: [PC, MK, SBP]
   - Current risks of transfusion.
   - Procedures to minimize risk for recipients.

**Apheresis for Collection of Blood Components, Patient Therapy and Blood Progenitor Cell Collections:**

The resident will have an understanding of the basic concepts of apheresis and stem cell collection. The resident will have a working knowledge of the variety of therapeutic apheresis instruments, procedures, and the appropriate indications.
**Therapeutic Apheresis**
The resident will:
- Have a clear understanding of the appropriate indications for either cytapheresis or plasmapheresis. [PC, MK, SBP]
- Act as a consultant and evaluate the patient before, during and after apheresis. [PC, MK, ICS, PRO, SBP]
- Be able to design a treatment plan including and write orders for use of replacement solutions. [PC, MK, ICS, SBP]
- Understand and recognize the side effects and reactions related to therapeutic apheresis. [PC, MK]
- Understand the processing of the instrumentation with regard to the separation and collection of various blood components. [PC, MK]

**KUMC Cell Therapy Laboratory:**

**Hematopoietic Stem Cell Processing**
The resident will:
- Understand the preparation and cryo-preservation of bone marrow/stem cells. [PC, MK]
- Have an introductory knowledge of the quality control aspects of a marrow processing laboratory including cell culture, cell viability, and sterile technique. [PC, MK, PBL]
- Have a general understanding of purging malignant cells. [PC, MK]
- Have a conceptual understanding of the protocol to change blood types in patients undergoing allogeneic transplants. [PC, MK]
- Understand the concepts of engraftment and the use of cytokines (recombinant growth factors) to speed marrow engraftment. [PC, MK]
- Understand the overall theory and methods of T-cell depletion. [PC, MK]
- Understand methods to positively select CD34+ stem cells. [PC, MK, PBL, ICS, PRO, SBP]
- Be familiar with techniques for thawing and transfusing frozen marrow or PBSCs including the side effects or reactions and will participate in these transfusions. [PC, MK, PBL]

**Umbilical Cord Blood Banking and Processing**
The resident will:
- Understand the role of cord blood in hematopoietic cell transplantation. [PC, MK, SBP]
- Understand the structure and operation of a cord blood bank. [PC, MK, PBL, SBP]
- Understand the medical evaluation and other criteria used to determine the suitability of cord blood for banking. [PC, MK]
- Understand the ethical issues involved in cord blood banking including those relating to consent. [PC, MK, PBL, PRO, SBP]
- Be knowledgeable of the methods used to process and store cord blood. [PC, MK]
- Be familiar with techniques for thawing and transfusing cord blood. [PC, MK]
SPECIFIC LEARNING OBJECTIVES FOR TRANSFUSION MEDICINE (SKILL LEVELS)

Skill Level I

- Demonstrate knowledge of the principles of patient/ unit identification and pretransfusion testing, including ABO/Rh testing, RBC antibody screen, and antibody identification. [PC, MK, PBL, SBP]
- Recognize the symptoms and signs of hemolytic and nonhemolytic transfusion reactions and demonstrate knowledge of the pathophysiology, treatment, and prevention of these complications. [PC, MK, SBP]
- Identify the major infectious complications of blood transfusions and the current risk of these infections, and explain how these infections can be prevented. [PC, MK, SBP]
- Identify the major noninfectious complications of blood transfusions, including transfusion-related acute lung injury, the risk of these complications, and strategies to prevent them. [PC, MK, SBP]
- Choose appropriate blood components and derivatives based on a thorough knowledge of the indications for transfusion. [PC, MK]
- Demonstrate knowledge of the pathophysiology, prevention, and treatment of hemolytic disease of the newborn. Recognize those antibodies in pregnant patients that are clinically significant and make appropriate recommendations for blood products. [PC, MK, SBP]
- Demonstrate knowledge of the pathophysiology and treatment of neonatal alloimmune thrombocytopenia. [PC, MK]
- Demonstrate proficiency in the evaluation and appropriate transfusion therapy of thrombocytopenic patients (both adult and pediatric). [PC, MK]
- Apply the principles of a massive transfusion protocol. [PC, MK, SBP]
- Demonstrate a working knowledge of the principles of hemostasis and coagulation and proficiency in the initial treatment of patients with bleeding disorders (see also the Hematology section). [PC, MK]
- Demonstrate knowledge of the transfusion requirements of special patient populations (e.g., hematology/oncology, pediatrics, geriatrics, transplantation, and burn/trauma). [PC, MK, SBP]
- Demonstrate knowledge of landmark published studies in transfusion medicine. [PC, MK, PBL]
- Demonstrate proficiency in evaluating and presenting findings from recent peer-reviewed journal articles related to transfusion medicine. [PC, MK, PBL, ICS, PRO]

Skill Level II

- Identify clinically significant RBC antibodies from an antibody panel including multiple alloantibodies and mixtures of alloantibodies and autoantibodies; determine how difficult it will be to obtain blood for this patient, and effectively communicate these results to clinicians. [PC, MK, ICS, PRO, SBP]
- Demonstrate proficiency in evaluating and recommending treatment plans for complex transfusion reactions. [PC, MK, PBL, ICS, PRO, SBP]
- Demonstrate familiarity with the appropriate use of highly specialized blood products (e.g., granulocytes, donor lymphocyte infusions, HLA-matched platelets, and coagulation factor concentrates). [PC, MK, SBP]
- Demonstrate familiarity with the requirements of all applicable regulatory and accrediting agencies [e.g., TJC, CAP, American Association of Blood Banks (AABB), and US FDA]. [PBL, SBP]
- Compare and contrast the various means of performing blood utilization reviews. [PC, MK, PBL, ICS, SBP]
• Demonstrate competence in the management of blood inventory and the ability to communicate effectively the hospital's needs to the blood supplier. [PC, MK, PBL, ICS, PRO, SBP]

• Demonstrate knowledge of various methods of blood conservation, including pre-and perioperative autologous blood collection, and approaches to “bloodless” surgery. [PC, MK, PBL, SBP]

• Demonstrate proficiency in evaluating patients that are refractory to platelet transfusions. Outline the principles of histocompatibility testing and platelet cross-matching and apply this knowledge in selecting appropriate platelet products when indicated (see also the Immunology and Immunogenetics section). [PC, MK, SBP]

• Demonstrate proficiency in the evaluation of patients with immune-mediated and non-immune-mediated hemolytic anemia and in the appropriate transfusion management of these patients. [PC, MK, SBP]

Blood Collection/Blood Center/Cell Processing Responsibilities

Skill Level I

• Compare and contrast the eligibility requirements for allogeneic and autologous blood donations. [PC, MK, SBP]

• Demonstrate knowledge of the indications for therapeutic phlebotomy. [PC, MK, SBP]

• Demonstrate proficiency in evaluating and treating adverse reactions associated with blood donation/phlebotomy (whole blood and apheresis donations). [PC, MK, SBP]

• Outline the assay principles of required donor blood tests and the associated confirmatory testing and describe donor re-entry algorithms. [PC, MK]

• Demonstrate professionalism in interactions with prospective donors. [ICS, PRO]

• Summarize the steps in blood component and blood derivative preparation. [PC, MK]

• Describe the factors that influence the motivation of volunteers to donate blood. [PBL, ICS, SBP]

• Explain the operational logistics required in determining appropriate blood inventory for a geographic region and the process of meeting daily, weekly, and monthly collection goals. [PC, MK, PBL, SBP]

Skill Level II

• Outline the necessary steps in donor notification and counseling associated with positive infectious disease testing results, and the donor look-back process. [PC, MK, PBL, ICS, PRO, SBP]

• Demonstrate knowledge concerning the requirements of all applicable regulatory and accrediting agencies. [PBL, SBP]

• Demonstrate knowledge of the principles of hematopoietic stem cell transplantation, including collection, processing, and storage of these stem cell products, and the indications for use (e.g., bone marrow, peripheral blood, and cord blood). [PC, MK, SBP]

• Demonstrate understanding of the elements of current good manufacturing practices and current good tissue practices as they apply to the collection, processing, ex vivo manipulation, and storage of all cellular therapeutic products (e.g., pancreatic islet cells, negative/positive selection/purging of hematopoietic stem cells, gene manipulations, donor lymphocyte infusions, dendritic cell vaccines, and ex vivo expansion of progenitor cells). [PC, MK, PBL, SBP]

• Develop an understanding of emerging areas of cellular therapy, including hematopoietic graft engineering and cellular immunotherapeutics. [PC, MK, PBL, SBP]
Therapeutic Apheresis

Skill Level I

• Summarize the principles of apheresis technology, including centrifugation, filtration, and immunoadsorption. [PC, MK]
• Demonstrate knowledge of the indications for therapeutic apheresis and of the appropriate replacement fluids to be used in various situations. [PC, MK, SBP]
• Demonstrate proficiency in evaluating and preparing patients for therapeutic apheresis, including discussion with the patient of the risks and benefits associated with apheresis procedures. [PC, MK, ICS, PRO, SBP]
• Communicate effectively with clinicians and housestaff regarding emergent or scheduled therapeutic apheresis procedures through conversations and writing of consult notes. [PC, MK, ICS, PRO, SBP]

Skill Level II

• Demonstrate proficiency in evaluating and treating adverse reactions associated with therapeutic apheresis. [PC, MK, SBP]
• Demonstrate proficiency in the treatment of patients using specialized methods (e.g., photopheresis and immunoadsorption columns). [PC, MK, SBP]

ADDITIONAL COMPETENCIES SPECIFIC TO TRANSFUSION MEDICINE

Patient Care

• Correctly classify transfusion reactions and give appropriate treatment recommendations.
• Choose appropriate cross-matching methods for various patients (e.g., electronic, immediate spin, and antiglobulin).
• Recognize and appropriately refer serological evaluations that are beyond the scope of a hospital-based transfusion service/blood bank.
• Correctly choose (or recommend) the appropriate blood product for patients with special needs.
• Triage and screen requests for blood components appropriately during inventory shortages.
• Demonstrate the ability to perform blood utilization reviews.
• Perform a donor interview and exam.
• Evaluate and perform initial management of whole blood and apheresis donor reactions.
• Write physician orders for peripheral blood hematopoietic stem cell collections and therapeutic apheresis procedures.
• Appropriately manage reactions that occur during peripheral blood hematopoietic stem cell collections or therapeutic apheresis procedures.

Medical Knowledge

• Demonstrate understanding of and ability to interpret major regulations and guidelines that are applicable to collection, processing, storage, and release of blood and other cellular therapeutic product.
Practice-Based Learning and Improvement
  • Demonstrate the ability to develop new policies and procedures or change existing policies and procedures based on a review of the literature or issuance of new guidelines by regulatory agencies.

Interpersonal and Communication Skills
  • Demonstrate the ability to discuss the process of therapeutic apheresis with patients, and/or family members where appropriate; answer their questions and obtain informed consent.

PGY 1 GOALS: By the end of the first year:
  • The resident should achieve competency in Skill Level 1.

PGY 2 GOALS: By the end of the second year:
  • The resident should achieve competency in Skill Levels 1 and some 2.
  • The resident should have worked up patients for autologous donor and pheresis in a competent manner.
  • The resident should have completed the rotation at the Community Blood Center.

PGY 3 GOALS: By the end of the third year:
  • The resident must achieve competency in Skill Level 1.

PGY 4 GOALS:
  • The resident should be competent in all Transfusion Skill Levels 1 and 2.

GENERAL
Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice-Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on page 19-20.

RECOMMENDED READING LIST:

KUMC Transfusion Service:

The resident will have a basic understanding of blood bank laboratory procedures, component production, blood collection, component therapy and evaluation of transfusion reactions.

Opportunities to Function as Consultant to other physicians

The pager for the daily clinical service at KUMC will be carried by the resident who is the primary contact from 8 a.m. to 5 p.m. for all clinical transfusion problems and consultations. The most common calls from clinicians on a daily basis include several on the topic of requests for HLA or cross-matched platelets in patients that are refractory to transfusion and questions about management of therapeutic apheresis patients. Calls also are received via this pager several times a day from blood bank staff and usually result in the need for the resident to directly contact our clinical colleagues. During the three months at KUMC, specific time periods will be assigned to the resident for carrying this pager, depending on the resident's experience and capabilities. As part of the clinical service team, the resident will document consultations and other patient updates directly in the patient's record.

The resident often interacts by phone and directly on the patient care unit with the clinicians taking care of the patient. Daily, the resident is on the patient care units, usually re-evaluating the use of components that were issued outside of the guidelines set by the Transfusion Committee (TC). The daily situations whereby the resident correlates laboratory results with clinical findings to determine whether special platelets are needed or should continue to be provided, including platelet counts, platelet antibody and platelet cross-matching results, the patient's clinical state (i.e., bleeding) or has non-immune causes of platelet refractoriness.
Approximately twice a week, the resident will correlate patient signs and symptoms with laboratory data in evaluating transfusion reactions. In plasma exchange patients, the resident will follow the clinical status daily, correlating with laboratory tests to determine effectiveness of therapy, and decide further treatments or manage adverse reactions (e.g., hypocalcemia). The resident will correlate results from several laboratories in the care of pheresis patients (e.g., TTP patients) – following hematology laboratory results (e.g., platelet count) and chemistry laboratory results (e.g., creatinine, LDH) and special hematology laboratory results (e.g., peripheral smear for schistocytes).

Transfusion Reaction Workups

All suspected transfusion reactions (8 a.m. to 5 p.m., weekdays) are immediately referred to the resident (when carrying pager) for immediate evaluation regarding need for intervention or special testing. Within 24 hours of a suspected reaction, the laboratory will have preliminary data (DAT, serum color, gram stain if performed, etc.) and refer the case to the resident with the evaluation report form. The resident obtains a clinical history for the patient, reviews the laboratory data and subsequently shares the data and the resident's preliminary assessment with the blood bank attending physician. Together, the assessment and recommendations are documented on the report.

All reactions are handled in this manner either by the resident or attending physician.

Training in the Unique Computer Needs of a Transfusion Service

During the blood bank laboratory orientation and training, the resident learns how laboratory staff use computers to acquire components from the blood supplier (Community Blood Center), enter test results, track the unit, and release the unit to the patient care unit. The resident also learns the principles behind the electronic cross-match using a computer. The resident frequently uses the blood bank computer for retrieving patient and blood bank data. The resident has computer continuous access in the resident/fellow office for patient care and for Medline literature searches.

Training in Immunologic/Serologic Aspects of Blood Product Screening

The resident is trained regarding blood donor testing at the Community Blood Center, including red blood cell serology and infectious disease serologic testing. In addition, they spend time learning neutrophil serology and platelet serology and cross-matches.

On-Call Duties

For a two-month period during the rotation, the resident carries the KUMC on-call pager during working hours and provides first call consultation and blood bank problem solving for KUMC. During this period, the resident will on average have one out of every seven days free of hospital duties. The resident is directly supervised by the KUMC blood bank medical director and has access to additional help from blood bank medical faculty at KUMC and the American Red Cross. While on-call, residents are supervised by a faculty member, who is available at all times, either via their office phone, pager, or home phone. During the KUMC site rotation, all calls are reviewed at the weekday morning blood bank rounds, with the HCMC blood bank medical director. At KUMC, the resident is expected to be available to laboratory personnel, either in person or by pager, throughout the working day. No on-call duties outside of regular laboratory working hours are assigned to the resident involving coverage at these sites.
Call duties are constructed in the following fashion. The resident who is assigned to transfusion medicine for the month is responsible for clinical pathology call from Monday to Friday, 8 a.m. to 5 p.m. The remainder of the call time is divided between all residents who are on a clinical pathology rotation who have previously rotated through transfusion medicine. The Chief Residents will make out the clinical pathology call schedule and make sure no resident is on-call for more than six days in a row.

**Reliable Communication with on-duty Faculty**

Blood bank medical directors and teaching faculty members on service are physically present during standard operating hours (8 a.m. to 5 p.m.) and immediately available in person or by pager; specific faculty members when not physically present in the laboratory are available by phone or pager. At all times, a supervising faculty member is on-call for evening and weekend questions.

At KUMC, the attending physician meets daily (weekdays) with the resident to discuss direct patient care activities. All residents on clinical pathology rotations, except for the hematopathology resident are expected to attend the weekly CP Call Rounds to review transfusion medicine calls from the previous week as well as interesting cases from other aspects of clinical pathology. In addition, the resident usually has an informal lunch meeting with the blood bank medical director at least once a week and often three times a week (case discussions, mentoring).

**Education in Blood Bank Management**

Blood supply inventory management is discussed at regular Transfusion Committee (TC) meetings. The resident should attend these meetings regularly.

**Scholarly Activities/Research**

The resident evaluates and discusses research findings in the literature and receives feedback from faculty. The resident is urged to write up and publish interesting or unique transfusion problems encountered during the rotation. Blood bank faculty eagerly offers to mentor the resident through the process leading to a publication.

If a resident is interested in a research project and has sufficient time, the blood bank faculty will arrange this in one of the transfusion medicine research labs.

**RESIDENT EVALUATION**

If problems with not meeting expected knowledge and skills are observed during the rotation, the lead faculty member meets with the resident to evaluate the problem and develop a corrective action plan. Residents are evaluated on performance of daily activities (described previously), participation in required meetings and conferences, and presentations to the staff on assigned cases. The residents are provided with continuous feedback on their performance during the rotation. In general, only deficiencies are noted in writing.

Residents are evaluated on their demonstrated ability to provide informative consultation to the clinical service teams, their medical knowledge, their application of this knowledge to efficient/quality patient care, and their diagnostic, technical and observational skills. Residents are also evaluated on their interpersonal skills, professional attitudes, reliability, and ethics with members of the teaching faculty, peers, laboratory staff, and clinicians. They are
further evaluated on their initiative in fostering quality patient care and use of the medical literature, as it relates to their assigned cases. Their timely completion of assigned interpretive reports is another component of the evaluation. Residents on probation receive a written, mid-rotation evaluation.
The clinical chemistry service provides two rotations (eight weeks) for all Department of Pathology & Laboratory Medicine residents. While designed for residents, clinical chemistry post-doctoral fellows and incoming chemical pathology fellows may also take this rotation either together with, or separate from the residents. In either case, the residents and fellows share equally the daily duties and are considered peers. Interaction with faculty is intended to provide a perspective on laboratory organization/design and the practice of clinical chemistry as seen in a variety of different settings.

The clinical chemistry rotation is organized to both teach the fundamental principles of clinical chemistry and to provide extensive experience with the day-to-day clinical application of those fundamentals. The fundamental principles are taught through a structured list of teaching objectives that are arranged into three separate units. Each unit coordinator is responsible for arranging the day-to-day scheduling of their unit, how and when the laboratory exercises will be performed, and when the resident will meet with faculty members. Depending on the faculty and the unit, this interaction includes the following:

- Didactic sessions on specific topics with faculty members.
- Assigned reading of textbooks and/or journal articles.
- Case studies, chart reviews, case presentations.
- Laboratory demonstrations and exercises.

At least one week prior to the start of each unit, a resident should review the section of this manual that describes the requirements for the next scheduled unit. Residents should then contact the Residency/Fellowship Coordinator to receive information regarding appointments for the didactic sessions and to complete any necessary arrangements for laboratory demonstrations and exercises. Prior to each unit, the resident should review the specific learning objectives and review the required reading assignment listed in each unit. If familiar with the material, the resident/fellow need not read the reading assignment word-for-word. However, if the material is not familiar, the resident/fellow should make every attempt to master it before meeting with faculty for the didactic sessions.

This will allow for more efficient use of time during the didactic sessions and will minimize the possibility of not having a chance to discuss with faculty any section that is confusing or needs extraction of the most clinically relevant points. The additional/optimal readings are for the resident particularly interested in that area or having a case or clinical question posed as part of their consultative responsibilities during the rotation. There is no expectation that a resident will read all of these additional/optimal readings during the clinical chemistry rotation.

The clinical applications are taught primarily through performance of a variety of assigned clinical responsibilities that are intended to provide extensive contact with attending and housestaff physicians from clinical services. Laboratory exercises are designed not only to illustrate technical aspects of clinical laboratory testing, but also interesting clinical results and potential clinically important interferences.
In some cases, the laboratory exercises involve actual performance of an “experiment” by the resident. Other times they merely observe a laboratory test being performed to gain some familiarity on the analytical time and complexity of a variety of representative tests. However, we recognize that our residents possess a wide variety in previous laboratory experience. Thus, the faculty has agreed that no laboratory exercise is required. At the beginning of each unit, the resident should clearly state which experiments they plan to do or not do so that the technologists who assist with the laboratory exercises do not waste time preparing for something the resident does not intend to complete.

Additional experience with practical problems related to laboratory operations and management is gained through involvement with basic issues of laboratory management, such as:

- Evaluation of analytical instrumentation.
- Laboratory staffing.
- Cost account of a new analytical method relative to an older one.
- New method evaluation for technical performance.
- Quality assurance practices.
- Review of abnormal results report from the lab information system.

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<th>Legend for Learning Activities for Residents</th>
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<td>Didactic lecture</td>
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<td>Faculty sign-out</td>
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<td>Journal club</td>
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<td>Directly supervised procedure</td>
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<td>Unknown slide conferences</td>
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<td>Project</td>
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<th>Legend for Evaluation Methods for Residents</th>
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<td>Practical slide exam</td>
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<td>In-house written exam</td>
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<td>360 multisource rating</td>
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<td>Portfolios</td>
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<td>Procedures and case logs</td>
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</tbody>
</table>
## CLINICAL CHEMISTRY CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of chemistry services.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of chemistry.</td>
<td>DL, JC, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity. Where clinically appropriate, consult on the use of laboratory-based therapeutics such as blood transfusion and other forms of cellular therapy.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.</td>
<td>DL, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>

## CLINICAL CHEMISTRY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to chemistry.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in transfusion medicine.</td>
<td>DL, JC, DSP, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>

KUMC Pathology Residency Manual
Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.  

| Recognize the unique aspects of transfusion medicine practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice. |
|---|---|---|
| DL, JC, DSP, RM, IC | RR, DO, GR/FE, SE, IWE |

Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations.  

| Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations. |
|---|---|---|
| DL, RM, LI, OT | DO, GR/FE, SE, IWE |

Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice.  

| Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting. |
|---|---|---|
| DL, JC, DSP, RM, LI, P | DO, GR/FE, SE, IWE |

**CLINICAL CHEMISTRY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**  
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
**Goal:**
Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the chemistry service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to chemistry.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
<td>JC, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
### CLINICAL CHEMISTRY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### CLINICAL CHEMISTRY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the chemistry laboratory in the healthcare system.</td>
<td>DL, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
<td>RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.</td>
<td>DL, FSO, LI, OT</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Understand and implement policies to continually improve patient safety as they relate to the chemistry laboratory.</td>
<td>FSO, RM, LI, P</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

*KUMC Pathology Residency Manual*
ADDITIONAL OBJECTIVES SPECIFIC TO CLINICAL CHEMISTRY

[Pathology Milestones]
PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice
For expanded definitions, see pages 30-31.

Laboratory Management
- Introduction to Reporting of Laboratory Results.
  - Be familiar with chemistry specimen processing, data handling, record keeping, telephone answering and stat procedure system. [PC, MK, PBL, ICS, PRO, SBP]
  - Learn the core lab chemistry automation line including: [PC, MK]
    ▪ Robotic movement of specimens.
    ▪ Automatic centrifugation.
    ▪ Serum level determination.
    ▪ Automatic decapping of specimens.
    ▪ Bar code identification and bidirectional interface with the LIS.
    ▪ Aliquoting of tubes for tests not intended for automation line.
    ▪ Analysis of assays on the LXIs, DXI and non-automation equipment.
    ▪ Understanding of computerized specimen storage and add-on orders done robotically.
  - Be familiar with the reporting units (SI and conventional), and enzymatic units. [PC, MK]
  - Understand the following:
    ▪ Laboratory accreditation – CLIA '88, JCAHCO, CAP. [PBL, SBP]
    ▪ Internal quality control programs-materials available, methods. [PC, MK, PBL, SBP]
      ▪ What is the precision of the method?
      ▪ How are "action" and "warning" limit defined?
      ▪ How large is the analytical imprecision compared to the reference range?
      ▪ Would a more precise analytical method be useful clinically?
    ▪ External quality control programs (proficiency testing)-CAP, others. [PC, MK, PBL, SBP]
  - Understand the impact of CLIA '88 on hospital and physician office laboratories. [PBL, SBP]
  - Be able to describe the concept of a quality assurance program, as opposed to a quality control program, as defined by the JCAHCO. Understand the concept of a quality assurance "indicator" and understand how to develop, perform, and utilize these indicators as part of an on-going quality assurance program. Attend quality assurance monthly meetings. [PBL, SBP]
  - Know the general principles of the clinical laboratory administrative structure for optimum direction and management. [PBL, SBP]
  - Know the principles for laboratory design. [PBL, SBP]
• Understand the following personnel matters: [PBL, ICS, PRO, SBP]
  o Registration, certification, licensure of various levels of laboratory personnel (CLA, MLT-non-degree, MLT-degree, MT, clinical chemists, pathologists) and agencies involved (ASCP, NCA, ABCC, ABP).
  o Importance of position descriptions and performance evaluations.
  o Ways of handling personnel problems with unionized and non-unionized employees.
  o Productivity measurements (annual billable tests).
• Understand principles concerning: [PC, MK, PBL, ICS, PRO, SBP]
  o What test should be offered and when they should be made available.
  o What analytical methods should be used - analytic, economic and other considerations. Instruments selection-financing (lease, purchase, "free" for reagent use.) routine vs. STAT, sample size.
  o Instrument maintenance - in-house availability or vendor service, service contracts.
  o Reagent selection - kits, price, delivery, ordering, inventory, quality control in-house and by the vendor.
  o Laboratory safety including state and federal regulations.
• Understand the importance of good communications with other areas of the hospital (e.g., medical and nursing staff, hospital administration, medical records, etc.). [ICS, PRO]
• Attend nurse liaison monthly meetings and round with the laboratory pathologists and administrative staff on Thursday mornings. [PBL, ICS]
• Attend all CAP Virtual Management courses. [PBL, PRO, SBP]

RECOMMENDED READING LIST:


**General Chemistry**

• Describe functions of the principal components of and major differences between the following instruments using a block diagram: [PC, MK]
  o Simple single-beam spectrophotometer.
  o Fluorometer.
  o Nephelometer.
  o Several major automated instruments used in the clinical chemistry laboratories (e.g., Beckman Coulter in the main KU Lab, Ortho Vitros at KU Med West and Cancer Center labs).
  o Describe Beer's Law, stray light, bandwidth, and Allen's correction. [PC, MK]
  o Know the importance of a spectrophotometer bandwidth. [PC, MK]
Describe the main clinical use of fluorescence polarization spectroscopy. Why is it useful? [PC, MK]

Describe the principles of flame emission and atomic absorption spectroscopy and the clinical uses of each? [PC, MK]

RECOMMENDED READING LIST:


**Therapeutic Drug Monitoring - Kinetics**

- List five reasons for performing drug analysis. [PC, MK, SBP]
- Know the definition and Clinical utility of the following pharmacokinetic terms: [PC, MK]
  - Volume of distribution.
  - Half-life.
  - Clearance.
  - Loading dose.
  - Steady state.
  - Bioavailability.
  - Therapeutic Concentration.
  - Peak.
  - Trough.

- Review the pharmacokinetics of the following drugs through case review and problems: [PC, MK]
  - Theophylline.
  - Vancomycin.

- Know the principles that are involved in the measurement of therapeutic drugs by: [PC, MK]
  - Fluorescent Polarization Immunoassays (FPIA).
  - Enzyme Multiplied Immunoassays (EMIT).
  - Cloned Enzyme Donor Immunoassay (CEDIA).
  - Kinetic Interaction of Microparticles in solution (KIMS).
  - High performance Liquid Chromatography (HPLC).
  - Gas Chromatography (GC).
  - Know the advantages and disadvantages of measuring drug levels by immunoassay techniques compared to those that use HPLC or GC. [PC, MK]
  - Know the circumstances wherein determinations of serum free drug levels are necessary. [PC, MK, PBL, SBP]
  - Describe the mechanism of toxicity, relevant serum drug concentrations and treatment for the overdose of salicylate and acetaminophen. [PC, MK, SBP]
**RECOMMENDED READING LIST:**


**Electrolyte/Acid-Base Balance**

- Understand the general operation of the following: [PC, MK]
  - Flame photometer.
  - Atomic absorption.
  - Blood gas and pH instruments.
  - Osmometer.
  - Ion Specific Electrodes.

- Understand the theoretical basis and application of ion-selective electrodes; understand the analytical difference between direct and indirect potentiometry, ion-specific electrodes on different instruments. [PC, MK]

- Know the flame emission methods for Na+ and K+, the atomic absorption methods for Ca++ and Mg++, the major methods for Ca++, phosphorus, Cl-, CO2 and HCO3, the flame, atomic absorption and electrode methods for lithium. [PC, MK]

- Understand the analytical principles and the quality control for blood gas measurements. [PC, MK, PBL]

- Understand the pathophysiology of water regulation, extracellular fluid sodium content, ventilation and oxygenation. [PC, MK]

- Understand the pathophysiology of hyponatremia, hypernatremia, hypokalemia, hyperkalemia, hypercalcemia, hypophosphatemia, hypomagnesemia. [PC, MK]

- Understand the pathophysiology of acid-base disturbances. Be familiar with the tables and nomograms for acute and chronic disorders. [PC, MK]

- Know the causes of hypoxemia; learn to calculate the arterial/alveolar pO2 difference. [PC, MK]

- Become familiar with the kinetics and effects of CO on oxygen saturation of hemoglobin. Know the principles of measuring saturated hemoglobin and methemoglobin by oximeter. [PC, MK]

**RECOMMENDED READING LIST:**


**Renal Function**

- Know the major methods for BUN and creatinine measurements.
- Know the relationship between BUN and creatinine in prerenal and renal azotemia.
- Know the relationship between serum creatinine and creatinine clearance and how methods (Jaffe vs. enzymatic creatinine) will affect these results.
- Know uric acid metabolism.
- Know the usual causes of hyperuricemia and their management.
- Discuss the ways of distinguishing prerenal, renal and post-renal failure.
- Know how to evaluate polyuria.

**RECOMMENDED READING LIST:**


**Basic Enzymology, Liver Function Tests, and Gastrointestinal Disease**

- Residents should have a working knowledge of the following basics:
  - The Michaelis-Menten equation.
  - Zero- and first-order kinetic portions of this equation.
  - Principles used in the measurement of serum or urine enzyme activity.
  - Principles used in the measurement of an organic molecule's concentration using an enzymatic method.
- Know the factors involved in clinical interpretation of enzyme tests:
  - Tissue distribution, intracellular location, isozymic forms of the commonly measured serum enzymes.
  - Mechanisms of release and duration of release from damaged cells and tissues.
  - How rates of enzyme clearance from plasma affect serum enzyme concentrations.
  - Which enzymes rise primarily because of induced enzyme synthesis rather than release from damaged cells.
- Be familiar with the analytical methods used in the determination of enzyme activity, enzyme mass, and isoenzymes:
  - Electrophoretic.
  - Spectrophotometric.
  - Immunoenzymatic.
- Know the normal anatomy, physiology and biochemistry of the liver:
  - Micro and macroanatomy of the liver and biliary tract.
Metabolism of carbohydrates, lipids and proteins in the liver.
- Hormonal influences in the liver (e.g. insulin, glucagon).
- Synthesis of specific plasma proteins.
- Bile acid synthesis and excretion.
- Urea synthesis and excretion.
- Drug metabolism in the liver.
- Metabolism and excretion of bilirubin.

- Understand the etiology and diagnosis of the major types of jaundice:
  - Pre-hepatic (hemolysis, ineffective erythropoiesis).
  - Hepatic (pre-microsomal, microsomal, post-microsomal, intrahepatic obstruction).
  - Post-hepatic (gallstones, stricture, carcinoma of the pancreas or biliary tree).

- Be familiar with the basic liver function tests:
  - Hepatocellular serum enzyme indicators (serum AST, ALT).
  - Obstructive jaundice indicators (serum conjugated bilirubin, alkaline phosphatase, gamma-glutamyltransferase).

- Know the macro and microanatomy of the upper and lower gastrointestinal tract and the specialized functions of each region.
- Understand the pathogenesis, diagnosis and management of acute and chronic pancreatitis.
- Know the difference between malabsorption and maldigestion, and how the laboratory is used to distinguish them.
- Understand the pathogenesis of diarrhea, the techniques for evaluation and the interpretation of stool testing.

RECOMMENDED READING LIST:

Analytical Techniques and Instrumentation

Skill Level I

- Understand the principles and operational characteristics of analytical chemistry techniques, including photometric, electrochemical, enzymatic, electrophoretic, radiometric, chromatographic, mass spectrometric, and immunologic methods (see also the Immunology and Immunogenetics section). [PC, MK]
- Understand different types of random-access automated analyzers and the measurement principles employed in these systems, including spectrophotometric, ion-selective electrode, and electrochemical methods, as well as immunologic methods, including enzyme multiplied immunoassay technique, cloned enzyme donor immunoassay, fluorescence polarization immunoassay, microparticle enzyme immunoassay, electrochemiluminescence, ELISA, turbidimetry, and nephelometry. [PC, MK]
- Understand the basic biology of, and analytical methods for, determination of qualitative and quantitative changes in blood and fluid proteins and amino acids (enzymes, biomarkers, hormones, and cytokines), carbohydrates, lipids and lipoproteins, and clinically relevant small molecules (including metals, trace elements, and vitamins). [PC, MK, SBP]

Skill Level II

- Understand the principles of laboratory robotics and automation strategies. [PC, MK, PBL, ICS, PRO, SBP]
- Understand the general principles of assay calibration, QC, and the need for calibration verification. [PC, MK, PBL, ICS, PRO, SBP]
- Understand the causes of both positive and negative interferences as well as how to detect and avoid them. [PC, MK, PBL, ICS, PRO, SBP]
- Understand the techniques employed for specific extraction of analytes from biological fluids. [PC, MK, PBL, ICS, PRO, SBP]
- Identify factors influencing separation and resolution in electrophoresis and chromatography, including mechanism of separation and mobile/stationary phases. [PC, MK, PBL, ICS, PRO, SBP]
- For chromatography, understand the importance of internal standards, the relative retention time, carryover, and matrix effects. [PC, MK, PBL, ICS, PRO, SBP]
- For mass spectrometry, understand the pitfalls of ion suppression and the need for defining characteristic ion ratios for reliable compound identification. [PC, MK, PBL, ICS, PRO, SBP]

ORGAN-BASED BIOCHEMICAL PATHOPHYSIOLOGY

ASSESSMENT OF PULMONARY FUNCTION: BLOOD GASES AND OXYGEN SATURATION

Skill Level I

- Understand the principles of partial pressure of gases and the need for an O2 carrier. Be able to describe the alveolar-arterial O2 gradient and anion gap. [PC, MK, PBL, ICS, PRO, SBP]
- Know the pathophysiology of ketoacidosis and lactic acidosis. [PC, MK, PBL, ICS, PRO, SBP]
• Understand the significance of P50, O2 content, O2 capacity, and O2 saturation and be able to distinguish between O2 saturation and Po2. [PC, MK, PBL, ICS, PRO, SBP]
• Be able to describe the hemoglobin-oxygen dissociation curve and factors that affect the curve and P50. [PC, MK, PBL, ICS, PRO, SBP]
• Understand the principles of integrated blood gas, electrolyte, and CO-oximetry systems. [PC, MK, PBL, ICS, PRO, SBP]

ACID-BASE CHEMISTRY, ELECTROLYTES, AND RELEVANT DISORDERS

Skill Level I
• Know the differential diagnosis of common electrolyte disorders. [PC, MK, PBL, ICS, PRO, SBP]

ASSESSMENT OF RENAL FUNCTION

Skill Level I
• Know the basic physiology of renal function. Understand the basic categories of renal diseases (e.g., pre-renal azotemia, obstructive azotemia, glomerulonephritis, acute vs chronic renal failure, uremic syndrome) and be familiar with the National Kidney Foundation practice guidelines for these conditions. Know the laboratory analytical methods (e.g., Jaffe vs creatinase) for the assessment of renal function (creatinine, urea nitrogen, glomerular filtration rate) and proteinuria. [PC, MK]
• Understand the concept of creatinine clearance, how it can be used to estimate glomerular filtration rate, and the various methods employed to measure it. Understand renal handling of electrolytes and key metabolites and the interpretation of urinary electrolyte measurements. [PC, MK]
• Understand the definition of osmolality, molecules in serum that contribute to osmolality, and calculation of osmolar gap as well as the principle of the osmometer. Understand the common pitfalls and sources of error during estimation of the osmolar gap (e.g., hyperproteinemina, hyperlipidemia, hypermagnesemia). Understand the differential diagnosis of an unexplained, increased osmolar gap, including alcohol or glycol ingestion, alcoholic or diabetic ketosis or ketoacidosis, and osmotherapy (e.g., mannitol or glycerol administration), among others. [PC, MK]. Understand the principles of fluid balance. [PC, MK]

CARDIAC BIOMARKERS FOR THE ASSESSMENT OF CORONARY ARTERY DISEASES

Skill Level I
• Know the current definition of myocardial infarction by the European Society of Cardiology/American College of Cardiology guidelines and the New York Heart Association classifications and understand the interaction of diagnostic modalities in its definition (electrocardiogram, laboratory testing, and imaging). [PC, MK, PBL, SBP]
Know the diagnostic and prognostic significance as well as the limitations of current coronary artery disease biomarkers [troponins I and T, creatinine kinase (CK-MB index and isoforms), and myoglobin]. [PC, MK, SBP]

Know the pathophysiology and evaluation of congestive heart failure. Understand the markers of congestive heart failure [B-type natriuretic peptide (BNP) and N-terminal fragment of the BNP prohormone (NTproBNP)] and their biological and technical limitations. [PC, MK]

Understand the utility of markers of inflammation in the evaluation of cardiac risk (e.g., homocysteine and C-reactive protein). [PC, MK]

ASSESSMENT OF LIVER AND BILIARY TRACT STATUS

Skill Level I

Understand the dynamics and mechanisms of liver enzyme release and the clinical utility of measuring hepatic enzymes (e.g., aspartate aminotransferase, alanine aminotransferase, glutamytransferase, alkaline phosphatase, and lactate dehydrogenase). [PC, MK, SBP]

Know the biochemical assessment of liver function by nonenzyme analytes such as albumin, ammonia, bile acids, bilirubin, urea nitrogen, cholesterol, total protein, and triglycerides. [PC, MK]

Understand bilirubin metabolism, fractionation of bilirubin (conjugated, unconjugated, bilirubin, direct vs indirect) and unique aspects of neonatal bilirubin. Understand the conditions and genetic defects that affect bilirubin metabolism, transport and clearance (e.g., Gilbert disease and Dubin–Johnson syndrome). [PC, MK]

ASSESSMENT OF THYROID FUNCTION

Skill Level I

Understand the structure, biosynthesis, secretion, and metabolism of thyroid hormones [thyroxine (T4), triiodothyronine (T3), and reverse T3 (rT3)]. Know thyroid physiology and control of thyroid function [thyrotropin-releasing hormone (TRH) and thyrotropin (TSH)]. [PC, MK]

Know the common causes of hypothyroidism and hyperthyroidism. [PC, MK]

Know the laboratory tests for evaluation of thyroid disorders and be able to interpret these analytes in their clinical context with an appreciation for the euthyroid sick state. [PC, MK]

Be familiar with current analytical methodologies for thyroid testing (TSH methods: 1st -, 2nd -, and 3rd generation assays; isotopic and nonisotopic methods; T4; free T3 methods; T-uptake methods; TSH suppression and stimulation tests). [PC, MK]

ASSESSMENT OF PITUITARY FUNCTION

Skill Level II

Understand the physiological action, biochemistry, and regulation of anterior pituitary hormones [adrenocorticotropic hormone (ACTH), growth hormone (GH), prolactin (PRL), luteinizing hormone (LH), follicle-stimulating hormone (FSH)] and of posterior pituitary hormones [antidiuretic hormone (ADH) and oxytocin]. [PC, MK]
• Understand endocrine tests of hypothalamic-pituitary function (cosyntropin test/rapid ACTH stimulation test, insulin hypoglycemia test, metyrapone test, levo-dopa test, arginine infusion test, glucose-GH suppression test, TRH test, gonadotropin-releasing hormone (GnRH) test, clomiphene test, corticotropin-releasing hormone (CRH) test, gonadotropin-releasing hormone test, water deprivation test, saline infusion test, and water loading test). Understand the pathophysiology of disorders of the pituitary. [PC, MK]

ASSESSMENT OF ADRENAL FUNCTION

Skill Level I
• Understand the physiological action, biochemistry, biosynthesis, chemical structure, and metabolism of glucocorticoids and mineralocorticoids. [PC, MK]
• Understand the physiological regulation of the renin-angiotensin-aldosterone system. [PC, MK]
• Understand clinical conditions associated with excess and deficiency of adrenal cortex hormones. Understand testing of the functional status of the adrenal cortex [basal values vs stimulation tests and suppression tests, circadian rhythm of corticosteroids, morning ACTH, cortisol (urinary, random, and free), rapid ACTH cortisol stimulation test, multiday ACTH stimulation, metyrapone stimulation, CRH stimulation, and quantitative serum and urinary steroid hormone panels]. [PC, MK]
• Understand the synthesis and metabolism of biogenic amines, including catecholamines and serotonin. [PC, MK]
• Be familiar with the strengths and weaknesses of tests available for evaluation of disorders of the adrenal medulla, such as pheochromocytoma or neuroblastoma. [PC, MK]

ASSESSMENT OF REPRODUCTIVE FUNCTION, PREGNANCY, AND PRENATAL TESTING

Skill Level II
• Understand the role of sex hormones in reproduction and the evaluation of pregnancy and reproductive dysfunction, such as menstrual disorders and infertility. [PC, MK]
• Understand the importance of demographic data and biochemical assessment in prenatal testing for fetal defects. [PC, MK]

ASSESSMENT OF GASTRIC, PANCREATIC, AND INTESTINAL FUNCTION

Skill Level I
• Understand the clinical manifestations of gastric, pancreatic, and intestinal disease and diagnostic methodologies such as the breath tests for Helicobacter pylori, fecal occult blood, lipase, and amylase (e.g., fractionation of amylase; pancreatic vs salivary and amylase/creatinine clearance ratio). [PC, MK]
• Appreciate the role of gastrointestinal hormones and enzymes in digestion and the evaluation of malabsorption and diarrheal syndromes. [PC, MK]
ASSESSMENT OF GLUCOSE AND EVALUATION OF DIABETES MELLITUS

Skill Level I
- Understand the metabolism of carbohydrates (insulin, C-peptide, and other regulatory hormones) and be familiar with the American Diabetes Association (ADA) definitions of impaired fasting glucose, impaired glucose tolerance, type 1 and type 2 diabetes mellitus, criteria for diabetic ketoacidosis and hyperosmolar hyperglycemic state, as well as gestational diabetes. Understand the underlying pathophysiology of different forms of diabetes. [PC, MK]
- Understand the diagnosis and laboratory assessment of diabetes (blood glucose, oral glucose tolerance test, hemoglobin A1c, fructosamine, and urinary microalbumin) and its complications. [PC, MK]
- Understand the diagnosis and evaluation of hypoglycemia. [PC, MK]

ASSESSMENT OF MINERAL AND BONE METABOLISM

Skill Level I
- Understand the biochemistry and physiology of calcium, phosphate, and magnesium. [PC, MK]
- Know the hormones that regulate mineral metabolism [parathyroid hormone (PTH), calcitonin, and vitamin D] as well as parathyroid hormone-related protein (PTHrP). Understand various PTH assays, including biointact PTH and intraoperative PTH. [PC, MK]
- Know the pathophysiology of metabolic bone diseases such as osteoporosis, osteomalacia, and Paget disease. [PC, MK]

ASSESSMENT OF PORPHYRINS AND DISORDERS OF PORPHYRIN METABOLISM

Skill Level II
- Understand the biochemistry of heme and porphyrins. [PC, MK]
- Understand the porphyrias and be able to consult on the selection and interpretation of both screening and diagnostic tests for each disorder. [PC, MK]

TUMOR BIOMARKERS

Skill Level I
- Be familiar with the definition, classification, biochemistry, and distribution of tumor markers, both protein and carbohydrate, including, but not limited to, prostate-specific antigen, calcitonin, human chorionic gonadotropin, alpha-fetoprotein, carcinoembryonic antigen, CA 15-3, CA 125, and CA 19-9.
- Know the limitations of laboratory assessment of various tumor markers and the factors affecting the results of different analytical procedures. [PC, MK]
- Understand the conceptual basis of assays used to screen for malignancy, including Bayes theorem. [PC, MK]

Skill Level II
- Be familiar with ongoing efforts to identify proteomic patterns for cancer detection. [PC, MK]
ASSESSMENT OF FETAL LUNG MATURITY

Skill Level I
- Understand the physiology of respiratory distress syndrome. [PC, MK]
- Understand fetal lung maturity testing [lecithin/sphingomyelin (L/S) ratio, phosphatidyl glycerol (PG), foam stability index (FSI or shake test), fluorescence polarization, and counting of lamellar bodies]. Understand the biochemistry, physiology, and diagnostic performance of fetal fibronectin. [PC, MK]
- Understand the biochemistry, physiology, and diagnostic performance of fetal fibronectin. [PC, MK]

TRACE ELEMENT ASSESSMENT

Skill Level II
- Understand the biochemistry, physiology, and metabolism of trace elements (iron, magnesium, zinc, copper, selenium, cobalt, and fluoride). Know the biochemistry and clinical significance of metal-binding proteins such as transferrin, ferritin, and ceruloplasmin. [PC, MK]
- Know the clinical assessments of trace elements (serum iron, iron-binding capacity, transferrin, transferrin saturation, serum ferritin, zinc protoporphyrin, and serum ceruloplasmin). [PC, MK]

VITAMIN ASSESSMENT

Skill Level I
- Know the definition and classification of vitamins: fat-soluble vitamins (A, D, E, and K) and water-soluble vitamins [B1,B2,B6,B12 (cobalamin), C, niacin, nicotinamide, folic acid, biotin, and pantothenic acid]. [PC, MK]
- Understand the clinical disorders associated with the deficiency as well as toxicity of vitamins. [PC, MK]

CHOLESTEROL AND LIPID ASSESSMENT

Skill Level I
- Understand the chemical structures, biosynthesis, classification, function, and metabolism of lipids and lipoproteins. [PC, MK]
- Understand the Fredrickson classification and the National Cholesterol Education Program Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (ATP III) classification of hyperlipidemia. [PC, MK]
- Understand the pathophysiology of lipid disorders. [PC, MK]
- Know the principles of analytical techniques for laboratory assessment of lipids. [PC, MK]

SERUM AND FLUID PROTEIN AND AMINO ACID ASSESSMENT

Skill Level I
- Understand the principles of protein analysis in body fluids (e.g., Kjeldahl and Biuret methods, refractometry, and qualitative dipstick). [PC, MK]
• Know the principles of serum, urine, and cerebrospinal fluid (CSF) protein electrophoresis. Recognize key patterns of dysproteinemias and monoclonal gammopathies (see also the Immunology and Immunogenetics section). [PC, MK]
• Understand approaches for distinguishing transudates vs exudates in fluids. [PC, MK]
• Know the analytical methods involved in genetic and acquired aminoacidurias and the current guidelines for screening neonates for these disorders. [PC, MK]

Skill Level II
• Understand the emerging technology of proteomics and its potential applications in clinical diagnostics. [PC, MK, SBP]

CLINICAL ENZYME KINETICS

Skill Level II
• Understand the principles of enzyme kinetics (e.g., Michaelis–Menten equation, concepts of Km, Vmax, and zero-order and first-order kinetics) and clinical enzymology, including isoenzymes, isoforms, and tissue distribution. [PC, MK]  
• Be familiar with the principles of analytical enzymology and know the concepts of activity vs mass assays (e.g., CK vs CK-MB assays). [PC, MK]

PEDIATRIC BIOCHEMISTRY

Skill Level II
• Understand the differences and unique aspects of pediatric and neonatal chemistry, including reference ranges. [PC, MK]

THERAPEUTIC DRUG MONITORING AND TOXICOLOGY

PHARMACOKINETICS

Skill Level I
• Understand the concepts of drug absorption, bioavailability, volume of distribution, and distribution phases (multicompartment models) and be able to predict peak drug levels. [PC, MK]
• Understand the differences between first-and zero-order kinetics of drug metabolism/elimination. [PC, MK]
• Understand the concepts of drug clearance, half-life, and the exponential rate constant. Be able to calculate steady-state drug levels and estimate peak and trough drug levels throughout a dosing cycle. [PC, MK]
• Understand the origin and consequences of nonlinear or zero-order pharmacokinetics on drug pharmacokinetics. [PC, MK]
• Understand the differences between measurement of total, free, and protein-bound drug levels and be able to assess the consequences of altered protein binding on pharmacokinetics and therapeutic drug monitoring. [PC, MK]
DRUG METABOLISM

Skill Level I
- Understand the differences between phase I and phase II drug metabolism reactions. [PC, MK]
- Appreciate the various consequences of competing metabolic pathways to modulate both the efficacy and toxicity of administered medications. [PC, MK]
- Appreciate the frequent interindividual variability of drug-metabolizing enzymes and its impact on the variability of drug response. [PC, MK]

PHARMACODYNAMICS

Skill Level I
- Understand the general mechanisms of drug action, including drug–receptor interactions, modulation of metabolic pathways, and nucleic acid biochemistry. [PC, MK]
- Understand how reference ranges for therapeutic drug monitoring are established and understand the varying utility of trough, peak, or steady-state drug levels for monitoring both drug efficacy and toxicity. Understand the therapeutic index. [PC, MK]

THERAPEUTIC DRUG MONITORING OF SPECIFIC DRUG CLASSES

Skill Level I
- Understand the principles and practice of therapeutic drug monitoring of antidepressants, mood stabilizers, and antipsychotics; anticonvulsants; cardioactive drugs; bronchodilators; antibiotics; and immunosuppressants. [PC, MK]
- Understand the relative significance of peak and trough levels for monitoring of these drug classes. [PC, MK]

TOXICOLOGIC SYNDROMES

Skill Level II
- Understand the pathophysiological basis and be able to recognize the five major toxicologic syndromes (cholinergic, anticholinergic, sympathomimetic, opiate, and sedative-hypnotic). [PC, MK]
- Be able to formulate a toxicologic differential diagnosis and be able to design a clinical laboratory testing protocol for each of the syndromes. [PC, MK]
- Understand the basic therapeutic approach to each syndrome. [PC, MK]

LABORATORY EVALUATION AND MANAGEMENT OF OVERDOSED OR POISONED PATIENTS

Skill Level I
- Be familiar with the National Academy of Clinical Biochemistry guidelines for Emergency Toxicology. [PC, MK, PBL, SBP]
- Understand the important differences between urine and blood (including serum and plasma) for monitoring and detection of drugs/xenobiotics. [PC, MK]
• Understand how to design and implement standardized STAT panels of laboratory tests for evaluation of overdosed/poisoned patients. [PC, MK, PBL]

• Understand the limitations of drug “screening” protocols and be able to consult on the design of more extensive drug-testing protocols to supplement the standard STAT panels. [PC, MK, PBL]

Skill Level II

• Understand the toxicologic profiles of specific agents, including acetaminophen, salicylates, alcohols and glycols, barbiturates, tricyclic antidepressants, carbon monoxide, organophosphates and carbamate, digoxin, lead, iron, and cyanide. [PC, MK]

• Understand the general supportive measures, the role of alkalinization, the importance of specific antidotes, and the variable efficacy of exchange transfusion, hemodialysis, plasmapheresis, and charcoal hemoperfusion of blood in the management of specific agents. [PC, MK]

LABORATORY EVALUATION OF DRUGS OF ABUSE

Skill Level I

• Understand the generic methodology of the routine immunoassays for drugs-of-abuse testing. [PC, MK]

• Be familiar with the major drugs of abuse and their clinical manifestations. [PC, MK]

• Know the common methods for adulteration of urine and the techniques available in the laboratory to detect them. [PC, MK]

Skill Level II

• Know the specific reactivities of each immunoassay, the standard cutoffs for detection, and whether the assay is capable of detecting the parent drug, its metabolites, or both. Know which members of a drug class are poorly or well detected by a generic immunoassay (e.g., oxycodone detection by the opiate immunoassay) and know the common causes of false positives due to cross-reactivities. [PC, MK]

RECOMMENDED READING LIST:


POINT-OF-CARE TESTING

Point-of-care testing (POCT) occurs across all disciplines, but because there are important common issues in its clinical use, it has been made a separate, distinct part of the curriculum in this document. The POCT curriculum may be taught in a concentrated fashion in any of the subdisciplines, depending on what is most appropriate to the institution.

Skill Level I

- Understand definitions of POC and waived testing. [PC, MK, PBL, SBP]
- Understand the range of analytes available in devices used at the point of care. [PC, MK]
- Understand the impact of POCT on clinical care, in terms of volume of tests performed, turnaround time, and the utilization of common POC tests (e.g., bedside glucose, rapid strep, and activated clotting time). [PC, MK, SBP]
- Understand the differences in reference ranges and test performance characteristics between POCT and central laboratory assays. [PC, MK, SBP]
- Appreciate the difference between POCT and near-patient testing and the personnel resources that best accomplish quality testing in these distinct situations. [PC, MK, PBL, SBP]

Skill Level II

- Understand the principles of performance for common POC tests such as glucose, urine drugs of abuse, rapid microbial antigen, and activated clotting time. Understand the performance characteristics of the common POC devices used for these tests. Know the issues surrounding specimen collection and preparation and the limitations and interpretation of results. [PC, MK, PBL, SBP]
- Understand the quality principles of POCT, including QC of unit-use testing devices, and proficiency/competency assessment of testing with multiple sites and operators and diverse testing personnel. [PC, MK, SBP]
- Understand the regulatory, administrative, and operational context of POC, waived, and home testing. [PC, MK, PBL, SBP]
- Be able to assess economic, workflow, human resources, and clinical factors driving the decision to perform testing at the point of care vs the central laboratory. [PC, MK, PBL, SBP]
- Know the most common test systems used in POCT. [PC, MK]
- Develop an appreciation of emerging POCT technologies, including microelectrical mechanical systems (MEMS) and other biosensor techniques, and their potential clinical applicability. [PC, MK, PBL, SBP]
RECOMMENDED READING LIST:


PGY 1 GOALS: By the end of the first year:

- The resident should achieve competency in Skill Level 1.

PGY 2 GOALS: By the end of the second year:

- The resident must achieve competency in Chemistry Skill Level 1 and most of 2.
- Residents should demonstrate the ability to do method validation, reference intervals and test utilization.
- Residents should be able to demonstrate the ability to perform laboratory accreditation and prepare the laboratory for accreditation.

PGY 3 GOALS: By the end of the third year:

- The resident should achieve competency in Skill Level 1.

PGY 4 GOALS:

- The resident should be competent in all Chemistry Skill Levels 1 and 2

GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice-Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on pages 19-20.

Guidelines for Patient Care and Specimen Handling

Specimen handling in the laboratories is the direct responsibility of the laboratory technologists. Resident decision making in the laboratory is under the direct supervision of the teaching faculty at their assigned site. The on-service teaching faculty members are physically present during standard operating hours; faculty members not physically present are rapidly available by phone or pager. No diagnosis is communicated to the clinicians before a faculty member has evaluated the case.
Resident Opportunities to Function as Consultant to Other Physicians

Residents have the responsibility, under faculty supervision, of discussing the interpretive consultative reports and laboratory results with appropriate members of the clinical/surgical teams. Through their discussions with the clinical team members, the residents have the opportunity to directly impact patient care.

On-call Duties

During this rotation, residents will, on average have one out of every seven days free of hospital duties. Due to the at-home nature of call and the limited number of emergencies, the call duties are constructed in the following fashion. The resident who is assigned to transfusion medicine for the month, is responsible for clinical pathology call from Monday to Friday, 8 a.m. to 5 p.m.

The remainder of the call time is divided between all residents who are on a clinical pathology rotation, who have previously rotated through transfusion medicine. The Chief Residents will make out the clinical pathology call schedule and make sure no resident is on-call for more than six days in a row. While on-call, residents are supervised by a senior staff member, who is available at all times, either via their office phone, pager or home phone.

Scholarly Activities / Research Activities

Residents are provided with continuous access to literature-searching programs. The expectation is that residents will utilize the medical literature to find up-to-date information on their cases. It is further expected that residents will utilize the medical literature to help provide our clinical colleagues with up-to-date knowledge related to their cases. During sign-out and discussion of cases, the residents and teaching faculty discuss each case from a scholarly perspective.

The resident and faculty discuss both normal and abnormal physiology and the mechanisms potentially responsible for creating the clinical findings observed. It is hoped that these discussions will foster an interest in research and the development of new knowledge. Residents are not only encouraged to become involved in research but technical, logistic, and economic support for such activities is available.

RESIDENT EVALUATION

Residents will be evaluated primarily on the performance of daily activities, particularly on evidence of their assimilation of the material present and their ability to apply it in a practical situation, namely, case interpretation. Also, they will be evaluated on participation in required meetings, conferences, and their presentations. The residents are provided with continuous feedback on their performance during the rotation. In general, only deficiencies are noted in writing. Residents are evaluated on their demonstrated ability to provide informative consultation to the clinical service teams, their medical knowledge, their application of this knowledge to efficient/quality patient care, and their diagnostic, technical and observational skills.

Residents are also evaluated on their interpersonal skills, professional attitudes, reliability, and ethics with members of the teaching faculty, peers, laboratory staff, and clinicians. They are further evaluated on their initiative in fostering quality patient care and use of the medical literature, as it relates to their assigned cases. Their timely completion of assigned interpretive reports is another component of the evaluation. Residents on probation receive a written mid-rotation evaluation.
HEMATOPATHOLOGY

FACULTY

Mark Cunningham, MD  Associate Professor and Director of Hematology and Flow Cytometry
Wei Cui, MD  Associate Professor and Director of Clinical Flow Cytometry Laboratory
Fred Plapp, MD  Professor
Lowell Tilzer, MD, PhD  Professor
Janet Woodroof, MD  Associate Professor
Da Zhang, MD, MSc  Associate Professor

The hematopathology rotation (three blocks) is designed to provide residents the opportunity to acquire comprehensive knowledge about hematologic diseases, acquire morphologic skills necessary for diagnosing hematologic diseases and develop effective consultative skills that aid the clinician in disease diagnosis.

Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Activity</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
<td>DL</td>
</tr>
<tr>
<td>Faculty sign-out</td>
<td>FSO</td>
</tr>
<tr>
<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
</tr>
<tr>
<td>Role modeling</td>
<td>RM</td>
</tr>
<tr>
<td>Lab inspections</td>
<td>LI</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
<td>IC</td>
</tr>
<tr>
<td>Online tools</td>
<td>OT</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
<td>USC</td>
</tr>
<tr>
<td>Project</td>
<td>P</td>
</tr>
</tbody>
</table>

Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Method</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
<td>RR</td>
</tr>
<tr>
<td>Direct observation</td>
<td>DO</td>
</tr>
<tr>
<td>Checklist</td>
<td>CL</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
</tr>
<tr>
<td>Standardized exam</td>
<td>SE</td>
</tr>
<tr>
<td>Practical slide exam</td>
<td>PSE</td>
</tr>
<tr>
<td>In-house written exam</td>
<td>IWE</td>
</tr>
<tr>
<td>360 multisource rating</td>
<td>360</td>
</tr>
<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
</tr>
</tbody>
</table>

KUMC Pathology Residency Manual
HEMOPATHOLOGY CORE COMPETENCY: PATIENT CARE

**Goal:**
Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of hemopathology services.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of hemopathology.</td>
<td>FSO, DL, JC, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.</td>
<td>FSO, DL, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>

HEMOPATHOLOGY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to hemopathology.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>FSO, DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in hemopathology.</td>
<td>DL, JC, DSP, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC, FSO</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>

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KUMC Pathology Residency Manual
Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.  

| DL, JC, RM | DO, CL, GR/FE, SE, IWE |

Recognize the unique aspects of hemopathology practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice.  

| DL, JC, DSP, RM, IC | RR, DO, GR/FE, SE, IWE |

Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations.  

| DL, RM, LI | DO, GR/FE, SE, IWE |

Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations.  

| DL, RM, LI, OT | DO, GR/FE, SE, IWE |

Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice.  

| JC, DSP, RM, P | DO, GR/FE, SE, IWE |

Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting.  

| DL, JC, DSP, RM, LI, P | DO, GR/FE, SE, IWE |

HEMOPATHOLOGY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

<table>
<thead>
<tr>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Objectives:</strong></td>
<td></td>
</tr>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>JC, RM</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>RM, IC</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>RM, LI</td>
</tr>
<tr>
<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT</td>
</tr>
</tbody>
</table>
### HEMOPATHOLOGY CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the hemopathology service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to hemopathology.</td>
<td>DSP, RM</td>
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</tr>
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<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
<td>JC, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
HEMOPATHOLOGY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

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<th>Objectives:</th>
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<th>Evaluation Activities</th>
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<td>Demonstrate compassion: be understanding and respectful of patients,</td>
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HEMOPATHOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

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<th>Objectives:</th>
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<td>Demonstrate understanding of the role of the hemopathology laboratory in</td>
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<td>Demonstrate the ability to design resource-effective diagnostic plans</td>
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<td>based on knowledge of best practices in collaboration with other clinicians.</td>
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<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
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Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.

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<thead>
<tr>
<th>Skill Level</th>
<th>Specific Objectives</th>
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<tbody>
<tr>
<td>I</td>
<td>Understand the clinical indications for bone marrow evaluation. [PC, MK, SBP]</td>
</tr>
<tr>
<td></td>
<td>Understand the diagnostic limitations of bone marrow aspirate and biopsy sections. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Learn technical aspects of performing and analyzing bone marrow aspiration and biopsy. Encourage performance of bone marrow aspiration and biopsy. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Identify sites for the acquisition of bone marrow in children and adults. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Learn handling, preparation and interpretation of bone marrow specimens, including special stains (e.g., silver stain and Prussian blue). [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Correctly assess bone marrow cellularity and M-E ratio. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Recognize effects of chemotherapy and growth factor stimulation on blood and bone marrow. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Understand common drug effects leading to benign cytopenias. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Correctly identify storage iron and assess adequacy. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Understand hematopoiesis and distinguish the stages for cells in each hematopoietic cell series.</td>
</tr>
<tr>
<td></td>
<td>Know the major hematopoietic regulatory factors/cytokines. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Recognize normal WBC, RBC, and platelet maturation as well as cellular dysplasia. [PC, MK]</td>
</tr>
<tr>
<td></td>
<td>Understand diagnostic principles involved in distinguishing transient myeloproliferative syndromes (such as associated with Down syndrome), transient cytopenias, and transient lymphocytoses from clonal disorders. [PC, MK]</td>
</tr>
</tbody>
</table>

Understand and implement policies to continually improve patient safety as they relate to the hemopathology laboratory.
Skill Level II

- Understand the pathophysiology, clinical findings, etiology, and expected bone marrow morphology for vitamin deficiency anemias, hemoglobinopathies, thalassemias, aplastic anemia, red cell aplasia, leukemias, myeloproliferative disorders, myelodysplastic syndromes, plasma cell dyscrasias, and mast cell diseases. [PC, MK]
- Integrate morphology, cytochemistry, immunophenotype, and molecular and cytogenetics in the differential diagnosis of acute and chronic leukemia, lymphoma, and myeloproliferative and myelodysplastic diseases. [PC, MK]
- Integrate peripheral blood smear and bone marrow findings and render a preliminary diagnosis. [PC, MK]
- Know the post-therapy findings seen after treatment for leukemia and the temporal relationships to marrow regeneration post therapy. [PC, MK]
- Recognize the bone marrow manifestations of infections (e.g., viral, fungal, and hemophagocytic syndromes). [PC, MK]
- Recognize the bone marrow manifestations of noninfectious systemic diseases (e.g., alcoholism, collagen vascular disease, and nonhematologic malignancies). [PC, MK]

RECOMMENDED READING LIST:


**PGY 1 GOALS: By the end of the first year:**

- The resident demonstrates the ability to properly work up a bone marrow biopsy. This includes correct identification of cells with appropriate cell count, write up and differential diagnosis.
- Demonstrate basic skills with interpretation of ancillary data (flow cytometry, cytogenetics, molecular genetics, immunohistochemistry).

**PGY 2 GOALS: By the end of the second year:**

- Same skills as Year 1, but be competent in some Skill Level 2 hematology.
- The resident should demonstrate the ability to synthesize flow cytometry, cytogenetics, and molecular studies with hematology findings.
PGY 3 GOALS: By the end of the third year:

- The resident demonstrates the ability to work up properly a bone marrow biopsy and aspirate, including correct identification of cells, appropriate cell count, write up and differential diagnosis.
- Demonstrates basic skills with interpretation of ancillary data (flow cytometry, cytogenetics, molecular genetics, immunohistochemistry).

PGY 4 GOALS:

- The resident should be competent in all Hematology Skill Levels 1 and 2.

GENERAL

Throughout the entire duration of residency training, the resident must also demonstrate the specific skills for Professionalism, Practice--Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on pages 19-20.

TRAINEE RESPONSIBILITIES

Hematopathology (i.e. Bone Marrow Pathology):

- Daily sign-out of bone marrow aspirations and biopsies:
  - Sign-out case with Pathology staff.
  - If the service is very busy and the resident is having difficulty completing the work in a timely fashion, the resident may need to ask the attending for help. The following are possible guidelines for case distribution by PG year. Please note that these are only guidelines, and final discretion on case distribution is the prerogative of the service attending.
    - PGY1: 8 bone marrows/day.
    - PGY2: 10 bone marrow/day.
    - PGY3 and 4: 12 bone marrows/day.
  - Prior to sign-out:
    - Organize all slides for an individual case (blood smear, marrow aspirate smears, core biopsy, cell block, special stains) on a single slide tray dedicated for that case.
    - Obtain all pertinent information for each case, including previous histologic material, laboratory data, and clinical data.
    - Write-up each case and establish a diagnosis.
- Maintain an organized marrow sign-out room.
- Attend teaching conferences held by pathology staff (e.g. CP Core Conference, AP Core Conference, etc.).
- Function as a consultant to resolve questions asked by lab personnel and clinicians; be available at all times by pager.
TRAIREE EVALUATION CRITERIA

- The trainee must show active and effective participation in the laboratory.
- The trainee must demonstrate knowledge about the clinical, pathologic, and diagnostic features of hematologic disease.
- The trainee must demonstrate skill in the microscopic analysis of neoplastic and benign hematologic disease.
- The trainee must demonstrate knowledge about the performance and interpretation of hematology tests.
- The trainee must demonstrate skill as a laboratory-based consultant for clinicians and laboratory personnel.
- The trainee must demonstrate skill in verbal presentations at clinical conferences.

VACATION POLICY

- In general, vacation is discouraged during this rotation because resident services are essential for working up and signing out bone marrow biopsies.
- If vacation is absolutely needed, or if sick leave is needed, the resident must find someone to cover. The first approach is to ask the wet hematology resident, provided that the wet hematology resident has experience in performing bone marrow biopsies.
- If there is no wet hematology resident, then ask the pathology Chief Resident to assign someone to cover. This generally requires pulling someone temporarily from another rotation.
- Inform the pathologist and heme lab supervisor when the resident will be away and who is covering.
- The sign-out pathologist must sign the resident leave form.
- The resident should always try and return the favor to the resident who provided the coverage.
WET HEME/FLOW CYTOMETRY

FACULTY

Mark Cunningham, MD  Associate Professor and Director of Hematology and Flow Cytometry
Wei Cui, MD  Associate Professor and Director of Clinical Flow Cytometry Laboratory
Fred Plapp, MD  Professor
Lowell Tilzer, MD, PhD  Professor
Janet Woodroof, MD  Associate Professor
Da Zhang, MD, MSc  Associate Professor

The wet hematology rotation/flow cytometry rotation (two blocks) is designed to provide residents the opportunity to acquire comprehensive knowledge and diagnostic skills in wet hematology (non-bone marrow) and flow cytometry (FCM).

Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Activity</th>
<th>Abbreviation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
<td>DL</td>
</tr>
<tr>
<td>Faculty sign-out</td>
<td>FSO</td>
</tr>
<tr>
<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
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<tr>
<td>Role modeling</td>
<td>RM</td>
</tr>
<tr>
<td>Lab inspections</td>
<td>LI</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
<td>IC</td>
</tr>
<tr>
<td>Online tools</td>
<td>OT</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
<td>USC</td>
</tr>
<tr>
<td>Project</td>
<td>P</td>
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</tbody>
</table>

Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Method</th>
<th>Abbreviation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
<td>RR</td>
</tr>
<tr>
<td>Direct observation</td>
<td>DO</td>
</tr>
<tr>
<td>Checklist</td>
<td>CL</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
</tr>
<tr>
<td>Standardized exam</td>
<td>SE</td>
</tr>
<tr>
<td>Practical slide exam</td>
<td>PSE</td>
</tr>
<tr>
<td>In-house written exam</td>
<td>IWE</td>
</tr>
<tr>
<td>360 multisource rating</td>
<td>360</td>
</tr>
<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
</tr>
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</table>
**WET HEME/FLOW CORE COMPETENCY: PATIENT CARE**

**Goal:**
*Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of wet heme/FCM services.*

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<tr>
<th>Objectives:</th>
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<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of wet heme and FCM.</td>
<td>DL, JC, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.</td>
<td>DL, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>

**WET HEME/FLOW CORE COMPETENCY: MEDICAL KNOWLEDGE**

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to wet heme and FCM.*

<table>
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<tr>
<th>Objectives:</th>
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<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in wet heme and FCM.</td>
<td>DL, JC, DSP, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
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<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>
Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.  

| Recognition of the unique aspects of wet heme/FCM practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice. | DL, JC, DSP, RM, IC | RR, DO, GR/FE, SE, IWE |
| Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations. | DL, RM, LI | DO, GR/FE, SE, IWE |
| Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations. | DL, RM, LI, OT | DO, GR/FE, SE, IWE |
| Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice. | JC, DSP, RM, P | DO, GR/FE, SE, IWE |
| Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting. | DL, JC, DSP, RM, LI, P | DO, GR/FE, SE, IWE |

**WET HEME/FLOW CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

| Objectives: | Learning Activities | Evaluation Activities |
| Demonstrate the ability to critically assess the scientific literature. | JC, RM | DO, GR/FE |
| Demonstrate knowledge of evidence-based medicine and apply its principles in practice. | JC, RM | DO, GR/FE |
| Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions. | JC, RM, OT | DO, GR/FE |
| Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice. | DSP, RM | DO, GR/FE, SE, IWE |
| Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety. | RM, IC | DO, GR/FE |
| Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel. | RM, LI | DO, GR/FE |
| Use proficiency programs to improve laboratory practices. | RM, LI, OT | DO, GR/FE |
**WET HEME/FLOW CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS**

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients’ families.*

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<tr>
<th>Objectives</th>
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<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the wet heme and FCM service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
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<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>RR, DO, GR/FE</td>
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<td>Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to wet heme/FCM.</td>
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<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
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### WET HEME/FLOW CORE COMPETENCY: PROFESSIONALISM

**Goal:**
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

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<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
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<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
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<td>DO, GR/FE</td>
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<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
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<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
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<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
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### WET HEME/FLOW CORE COMPETENCY: SYSTEM-BASED PRACTICE

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<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
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Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.

| DL, FSO, LI, OT | DO, CL, GR/FE, SE, IWE |

Understand and implement policies to continually improve patient safety as they relate to the wet heme/FCM laboratory.

| FSO, RM, LI, P | DO, GR/FE |

ADDITIONAL SPECIFIC LEARNING OBJECTIVES

[Pathology Milestones]
PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice
For expanded definitions, see pages 30-31.

Automated Hematology

Skill Level I
- Understand clinical indications for peripheral blood cell enumeration and differential analysis. [PC, MK]
  Know the components of a complete blood count and understand the information provided by each. [PC, MK]
- Understand the principles of automated cell counting, including red blood cell (RBC) indices and their derivation. [PC, MK]
- Understand how “absolute values” are determined and how they differ from “relative percent”. [PC, MK]
- Identify spurious white blood cell (WBC) RBC, hemoglobin, and platelet determinations and be able to propose a course of action to be followed for reporting results. [PC, MK]
- Understand appropriate WBC correction for the presence of nucleated RBCs. [PC, MK]
- Understand automated differential analysis and manual review criteria. [PC, MK, PBL]
- Understand the absolute neutrophil count and its clinical utility, as well as problems associated with band counts. [PC, MK, SBP]
- Understand QC procedures specific to cell counters, such as Rumke limits on differential cell counts and Bull analysis of indices. [PC, MK, PBL]
- Understand principles of automated and manual reticulocyte enumeration and their respective technical limitations. [PC, MK]
Skill Level II
- Interpret results of automated and manual cell counts and understand the relevant technical limitations. [PC, MK]
- Recommend appropriate steps for abnormal sample processing, analysis, and result reporting. [PC, MK, PBL]
- Review abnormal results and correlate results with peripheral blood smear findings and clinical history. [PC, MK]

Peripheral Blood Smear Analysis

Skill Level I
- Know proper preparation and handling of peripheral blood smears, including standard stains and special stains used to identify cellular structures and inclusions. [PC, MK]
- Understand normal RBC, WBC, and platelet morphology. [PC, MK]
- Be able to estimate WBC and platelet counts. [PC, MK]

Skill Level II
- Recognize abnormal RBC, WBC, and platelet morphology; formulate a differential diagnosis and suggest appropriate laboratory testing for follow-up. [PC, MK]
- Recognize technical artifacts in WBC, RBC, and platelet morphology. [PC, MK]
- Recognize infectious disorders that can be diagnosed by blood smear. [PC, MK]
- Recognize storage disorders and congenital disorders that have morphologic manifestations in the peripheral blood smear. [PC, MK]
- Correlate peripheral blood smear findings with bone marrow morphology. [PC, MK]

Body Fluid Analysis: CSF, Ascitic/Pleural Fluid, Joint Fluid

Skill Level I
- Understand clinical indications for body fluid analysis. [PC, MK]
- Understand manual hemocytometer cell counting. [PC, MK]
- Understand cytocentrifuge sample preparation and slide staining. [PC, MK]
- Identify blood and body fluid cell morphology. [PC, MK]

Skill Level II
- Interpret results of body fluid analyses in the appropriate clinical context. [PC, MK]
- Recognize malignant cells and recommend appropriate confirmatory tests. [PC, MK]
- Correlate abnormal body fluid cell morphology with cytology, flow cytometry, and other relevant diagnostic test results. [PC, MK]
- Identify body fluid crystals. Distinguish between urate and calcium pyrophosphate crystals, using polarized light. [PC, MK]
Manual Hematology Methods

Skill Level I
- Understand principles of microhematocrit determination and its technical limitations. [PC, MK]
- Understand the principles of erythrocyte sedimentation rate. [PC, MK]
- Understand the principle and utility of supravital stains, including reticulocyte stain, hemoglobin H preparation, and Heinz body preparation. [PC, MK]

RBC Disorders

Skill Level I
- Learn the clinical indications for laboratory tests involved in the assessment of intrinsic and extrinsic RBC defects/disorders. [PC, MK]
- Know the pathophysiology and characteristic laboratory findings of the major disorders causing normocytic, microcytic, and macrocytic anemia. [PC, MK]
- Describe iron metabolism and laboratory tests for iron depletion. [PC, MK]
- Understand hemoglobin synthesis and degradation. [PC, MK]
- Understand the principles of hemoglobin screening by HPLC and electrophoresis at acid and alkaline pH. [PC, MK]
- Understand the principle and clinical utility of screening tests for the presence of hemoglobin S. [PC, MK]
- Know the pathophysiology and laboratory features of intravascular and extravascular hemolysis. [PC, MK]
- Understand the principle and clinical utility of Kleihauer–Betke and/or flow cytometric analysis for fetal hemoglobin. [PC, MK]

Skill Level II
- Interpret hemoglobin electrophoretic patterns and ancillary tests for the diagnosis of: [PC, MK]
  - Major hemoglobinopathies.
  - Disorders related to enzyme defects.
  - Hereditary spherocytosis and other RBC membrane/cytoskeletal defects.
  - Paroxysmal nocturnal hemoglobinuria.
  - Hemolytic anemia.
  - Congenital dyserythropoietic anemias.

Platelet Disorders

Skill Level I
- Understand the pathophysiology of thrombocytopenia and thrombocytosis. [PC, MK]
- Differentiate between reactive and malignant processes. [PC, MK]
- Understand the pathophysiology of immune thrombocytopenia and thrombotic thrombocytopenic purpura. [PC, MK]
• Demonstrate competency in taking a bleeding history. [PC, MK]
• Understand the clinical utility of platelet function testing. [PC, MK]
• Understand general principles of platelet function testing. [PC, MK]
• Understand the pathophysiology of acquired and congenital platelet function disorders. [PC, MK]
• Understand the pathophysiology leading to major von Willebrand disease subtypes and expected laboratory results. [PC, MK]
• Recognize acquired platelet function abnormalities associated with antiplatelet therapy. [PC, MK]

Skill Level II
• Interpret platelet function studies, including screening tests, platelet aggregation, and platelet secretion studies. [PC, MK]
• Interpret studies performed for the evaluation of von Willebrand disease. [PC, MK]

Coagulation

Skill Level I
• Understand the clinical utility of coagulation and thrombosis testing. [PC, MK, SBP]
• Develop basic understanding of hemostatic and thrombotic disorders. [PC, MK]
• Understand the coagulopathy of liver disease. [PC, MK]
• Understand the pathophysiology of vitamin K deficiency and antagonism. [PC, MK]
• Understand the laboratory evaluation of disseminated intravascular coagulation. [PC, MK]
• Understand the pathophysiology of the hemophilia (A, B, and C). [PC, MK]
• Understand the pathophysiology of arterial and venous thrombosis. [PC, MK]
• Understand the general principles of screening coagulation tests (e.g., prothrombin time, activated partial thromboplastin time, fibrinogen, and thrombin time). [PC, MK]
• Understand the International Normalized Ratio derivation and its clinical significance. [PC, MK, SBP]
• Understand the effect of hematocrit and blood-drawing technique on anticoagulation of blood samples for coagulation testing. [PC, MK]
• Demonstrate competency in taking a bleeding and thrombosis history. [PC, MK]
• Understand results of mixing studies and factor assays to guide further coagulation testing. [PC, MK]
• Understand the principles of tests involved in the identification of lupus anticoagulant and antiphospholipid antibody syndromes. [PC, MK]
• Recognize the effect of circulating anticoagulants on coagulation testing. [PC, MK]
• Understand the monitoring of anticoagulation therapy. [PC, MK]
• Understand the method of action of direct thrombin inhibitors and their effect on coagulation testing. [PC, MK]
• Understand the principles of molecular analysis of thrombotic risk factors [e.g., factor V Leiden, prothrombin G20210A, and methylenetetrahydrofolate reductase (MTHFR)]. [PC, MK]
• Understand the principles of functional and antigenic assays for proteins of the anticoagulation and fibrinolytic systems. [PC, MK]
Skill Level II
- Interpret results of coagulation and hypercoagulability testing and recommend further studies as needed. [PC, MK]
- Summarize laboratory evidence for hemostatic and thrombotic disorders and be able to assess and explain bleeding and thrombosis risk. [PC, MK]
- Interpret results of Bethesda assays for factor inhibitors. [PC, MK]
- Interpret results of coagulation tests in the setting of fibrinolytic therapy. [PC, MK]
- Interpret results of heparin-induced thrombocytopenia testing (ELISA tests vs serotonin release assay/platelet aggregation studies) in the appropriate clinical context. [PC, MK]
- Understand monitoring and complications of biologics as drugs (e.g., recombinant activated protein C and recombinant F VIIa). [PC, MK]

Flow Cytometry

Skill Level I
- Understand clinical indications for flow cytometric evaluation of blood, marrow, solid tissue, or fluid cells. [PC, MK]
- Understand the physical components and operating principles of a flow cytometer. [PC, MK]
- Understand QC procedures unique to flow cytometry assays (e.g., nature of controls and accounting for all lymphocyte subsets in a blood sample). [PC, MK, PBL]
- Understand the principles of routine flow cytometry evaluation of leukocytes, including both surface and intracellular markers, and recognition of clonal abnormalities. [PC, MK]
- Understand principles of tests designed to evaluate DNA content (ploidy) and cell cycle, as used in the evaluation of products of conception and other tissues. [PC, MK]
- Understand platelet antibody testing by flow cytometry and its clinical applications. [PC, MK]
- Understand the diagnostic and prognostic information provided by flow cytometry. [PC, MK]
- Understand the principles of lymphocyte subset analysis: know the commonly used antigens to define T-cell subsets, natural killer, and B cells. [PC, MK]
- Appreciate the effect of age on lymphocyte subset normal ranges. [PC, MK]
- Observe/perform lymphoma/leukemia panel on blood and/or bone marrow. [PC, MK]
- Observe/perform lymphoma panel on lymph node or spleen specimens. [PC, MK]

Skill Level II
- Evaluate and interpret results of flow cytometry in conjunction with cytochemistry, immunocytochemistry, immunohistochemistry studies, and lymph node pathology as related to hematopoietic and lymphoproliferative diseases: –Understand the characteristic clinical, morphologic. [PC, MK, SBP]
- Immunophenotypic, cytochemical, and cytogenetic/ molecular features of acute myeloid leukemia, acute lymphoid leukemia, myelodysplastic syndromes, paroxysmal nocturnal hemoglobinemia, multiple myeloma, monoclonal gammopathy of undetermined significance, non-Hodgkin and Hodgkin lymphoma, neuroblastoma, chronic lymphoproliferative disorders, lymphomatoid granulomatosis, posttransplant lymphoproliferative disorder, polymorphic and lymphomatoid papulosis, and histiocytic disorders. [PC, MK SBP].

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• Interpret specific flow cytometric abnormalities associated with immunodeficiency syndromes. [PC, MK]
• Interpret CD34 counts for stem cell transplantation and for prognostication in myeloproliferative disorders. [PC, MK]
• Understand the principles and interpretation of reticulated platelet analysis. [PC, MK]
• Understand the principles of, and interpret analyses for, minimal residual disease. [PC, MK]

PGY SPECIFIC GOALS – See Hematopathology section where PGY specific goals are described for Hematopathology/wet hematology and flow cytometry.

TRAINEE RESPONSIBILITIES

• Sign-out of selected hematology tests with pathology staff:
  o Blood smears.
  o Body fluid cytospins.
  o Flow cytometry panels.
  o Hemoglobin electrophoresis.
  o Platelet aggregation.
  o Platelet function analyzer.

• Prior to sign-out:
  o Obtain all pertinent information about the case, including previous laboratory and clinical data.
  o Review the relevant pathology slides; perform a leukocyte differential count for blood smear and cytospin evaluations.
  o Write a narrative interpretation.
  o Review bone marrow aspirate smears to approve flow cytometry orders.
  o Review the procedure manual for all hematology tests performed in the laboratory; perform selected tests under technologist supervision.
  o Attend the weekly Internal Medicine Hematology Conference (each Thursday, 8:30 am).
  o Attend teaching conferences held by pathology staff (e.g. CP Core Conference, AP Core Conference, etc.).
  o Function as a consultant to resolve questions asked by lab personnel and clinicians; be available at all times by pager.
  o Provide training and cross-coverage for the bone marrow resident or post-sophomore fellow.

TRAINEE EVALUATION CRITERIA

• The trainee must show active and effective participation in the laboratory.
• The trainee must demonstrate knowledge about the clinical, pathologic, and diagnostic features of hematologic disease.
The trainee must demonstrate skill in the microscopic analysis of neoplastic and benign hematologic disease.

The trainee must demonstrate knowledge about the performance and interpretation of hematology tests.

The trainee must demonstrate skill as a laboratory-based consultant for clinicians and laboratory personnel.

The trainee must demonstrate skill in verbal presentations at clinical conferences.

RECOMMENDED READING LIST:

Also see the Hematopathology Reference Materials in previous section.

The three rotations through microbiology and immunology are designed to aid residents to become competent in the interpretation of clinical microbiology and immunology testing and to learn consultative skills that aid in the diagnosis of infectious disease and immune disorders.

### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of microbiology/immunology services.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>DSP, RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
</tbody>
</table>

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**Legend for Learning Activities for Residents**

- Didactic lecture (DL)
- Faculty sign-out (FSO)
- Journal club (JC)
- Directly supervised procedure (DSP)
- Role modeling (RM)
- Lab inspections (LI)
- Interdisciplinary conference (IC)
- Online tools (OT)
- Unknown slide conferences (USC)
- Project (P)

**Legend for Evaluation Methods for Residents**

- Report review (RR)
- Direct observation (DO)
- Checklist (CL)
- Global rating/faculty evaluation (GR/FE)
- Standardized exam (SE)
- Practical slide exam (PSE)
- In-house written exam (IWE)
- 360 multisource rating (360)
- Portfolios (PF)
- Procedures and case logs (PCL)
Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.

Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of microbiology/immunology.

Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity. Where clinically appropriate, consult on the use of laboratory-based therapeutics such as blood transfusion and other forms of cellular therapy.

Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.

Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.

### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to microbiology/immunology.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in microbiology/immunology.</td>
<td>DL, JC, DSP, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.</td>
<td>DL, JC, RM</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Recognize the unique aspects of microbiology/immunology practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>
Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations.  

![DL, RM, LI]  DO, GR/FE, SE, IWE

Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations.  

![DL, RM, LI, OT]  DO, GR/FE, SE, IWE

Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice.  

![JC, DSP, RM, P]  DO, GR/FE, SE, IWE

Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting.  

![DL, JC, DSP, RM, LI, P]  DO, GR/FE, SE, IWE

### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.*

<table>
<thead>
<tr>
<th>Objectives:</th>
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</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the microbiology/immunology service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
<td>JC, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

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### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: PROFESSIONALISM

**Goal:**
> Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### MICROBIOLOGY/IMMUNOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
> Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the microbiology/immunology laboratory in the healthcare system.</td>
<td>DL, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
<td>RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>
Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.

Understand and implement policies to continually improve patient safety as they relate to the microbiology/immunology laboratory.

ADDITIONAL SPECIFIC OBJECTIVES IN MICROBIOLOGY

[Pathology Milestones]
PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice
For expanded definitions, see pages 30-31.

Skill Level 1
- Be able to discuss and execute general requirements for laboratory safety. [MK, PBL, SBP]
- Know the principles of universal precautions. [MK, PBL, SBP]
- Understand preanalytic, analytic and postanalytic factors affecting interpretation of microbiological tests. [PC, MK, PBL]
- Know the commonly occurring clinical bacterial organisms and the appropriate laboratory methods employed for diagnosis. [PC, MK]
- Know what organisms constitute normal flora in various specimen types (sputum, urine, wound, tissue, etc.). [PC, MK]
- Recognize the typical Gram stain appearance, colony morphology and growth conditions for Staphylococcus, Streptococcus, E. coli, Proteus, Pseudomonas and Hemophilus. [PC, MK]
- Understand quality control/quality assurance principles for microbiology laboratory procedures (i.e.; media, reagents or kits and antimicrobial susceptibility testing). [PC, MK, PBL]
- Know the test methods (manual and automated) and principle of the following antimicrobial susceptibility testing method: [PC, MK]
  - Disk diffusion tests.
  - Broth dilution tests.
  - Agar diffusion (E test).
  - Automated systems (Vitek, Microscan, Phoenix systems).
- Describe the mechanisms that cause the following resistance in bacteria; methicillin resistance, extended spectrum beta lactamase, penicillin resistance in Streptococcus species. [PC, MK]
• Know the commonly occurring infections due to mycobacteria and the appropriate laboratory specimen needed for diagnosis. [PC, MK, SBP]

• Be able to describe the life cycles of intestinal, tissue and blood parasites. [PC, MK]

• Understand proper specimen collection, transportation and methods used to detect parasitic infections. [PC, MK]

• Be able to discuss appropriate laboratory methods used for diagnosis of viral diseases. [PC, MK]

• Describe the different methods used for serologic testing such as EIA and Western blot. [PC, MK]

• Be able to interpret results of antibody tests for hepatitis viruses, Epstein Barr virus panel, HIV Western blot and other viral pathogens. [PC, MK]

Skill Level 2

• Understand and be able to supervise performance of and interpret routine microbiologic staining procedures, including: [PC, MK]
  o Gram stains especially from CSF and sputum.
  o Acid-fast stains.
  o Wet mounts with and without Trichomonas.
  o Trichrome stains.
  o Malaria smears (pass the competency slide set of 10 various malaria smears).

• Know the common bacterial pathogens that infect various organs and their pathogenesis.

• Be knowledgeable about the laws and regulations governing select agents and how these agents should be identified and handled in a clinical laboratory. [PC, MK, PBL, SBP]

• Understand the principle and interpretation of: [PC, MK]
  o Conventional identification systems.
  o Commercially available identification systems (manual vs. automated).

• Know the commonly occurring infections due to yeasts and fungi. [PC, MK]

• Recognize the gross and microscopic culture characteristics of the following yeast/fungi: [PC, MK]
  o Cryptococcus neoformans.
  o Candida albicans.
  o Histoplasma capsulatum.
  o Coccidioides immitis.
  o Aspergillus fumigatus.
  o Rhizopus spp./Mucor spp.
  o Dematiaceous fungi.

• Understand the methods employed for the identification of yeasts and fungi. [PC, MK]

• Be able to describe specimen processing needed for mycobacterial cultures such as concentration and digestion. Know why and when this is needed. [PC, MK]

• Know the grouping of non-tuberculosis mycobacteria. [PC, MK]

• Understand the methodology and the limitations for direct detection of M. tuberculosis in clinical specimens. [PC, MK]

• Be acquainted with the NCCLS standards and guidelines for antimicrobial testing and be able to interpret different testing methods. [PC, MK, PBL, SBP]

• Understand which variables affect susceptibility testing. [PC, MK]
• Be able to recognize incongruent susceptibility results and have a plan for how to verify the results. [PC, MK]
• Describe the mechanisms of the common antiviral agents such as acyclovir, amantidine, oseltamivir, ATZ, gancyclovir and others. [PC, MK]
• Be able to interpret the listed serologic and molecular tests used for diagnosis of viral infections: [PC, MK]
  o HIV testing Western blot.
  o HIV viral load detected by PCR.
  o Understand mechanisms of transmission of nosocomial infections. [PC, MK]
  o Know the diseases that are required to report to the Kansas Department of Health and Environment. [PC, MK, SBP]

### ADDITIONAL SPECIFIC OBJECTIVES FOR IMMUNOLOGY

#### Skill Level 1

- Understand the performance and interpretation of tests for the detection of antigens and antibodies in various body fluids (including the various factors that might interfere with or produce error in the various immunoassay techniques). [PC, MK]
- Recognize the following reasons for a false positives and false negative test result in serological assays: [PC, MK]
  o Carry over effect.
  o Prozone effect (antibody and antigen excess).
  o Types of antibody reagents (monoclonal vs. polyclonal).
  o Stability of reagents (complement).
- Understand and be able to compare and contrast various immunodiagnostic method such as EIA, latex agglutination, nephelometry, immunodiffusion, electrophoresis (protein and nucleic acid). [PC, MK]
- Understand the performance and interpretation of clinically significant antibodies for diagnosis and evaluation of autoimmune disorders. [PC, MK]

#### Skill Level 2

- Recognize the various patterns associated with disease status when interpreting fluorescent ANA screens and titers. [PC, MK]
- Know the methods used to detect Rheumatoid factor (screen and titers). [PC, MK]
- Know the disease states associated the specific antibody epitopes (SSA, SSB, Sm, RNP, dsDNA, ssDNA, SCL70, histones) and be able to interpret various patterns. [PC, MK, SBP]
- Be able to interpret selected serologic tests and assays for diagnosis of infectious disease: [PC, MK]
  o Heterophile antibody detection.
  o Syphilis serology (non-treponemal and treponemal-specific).
  o Viral hepatitis.
**PGY 1 GOALS: By the end of the first year:**
- The resident should achieve competency in Skill Level 1.

**PGY 2 GOALS: By the end of the second year:**
- The resident should achieve competency in Skill Level 1 and some of Level 2.

**PGY 3 GOALS: By the end of the third year:**
- The resident should achieve competency in Skill Level 1 and most of Level 2.

**PGY 4 GOALS:**
- The resident should be competent in all microbiology and immunology Skill Levels 1 and 2.

**GENERAL**
Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionism, Practice-Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on pages 19-20.

**DUTIES AND RESPONSIBILITIES OF THE RESIDENT**
- Be present in the laboratory for significant periods of time to allow exposure to routine laboratory work.
- Attend in-patient rounds with infectious disease team.
- Meet the general objectives outlined above and complete any work on unknowns, reading assignments or projects assigned during the rotation.
- Attend the weekly teaching session on Wed at 8 AM.
- Attend or present one 30 minute case report at Clinical Pathology Conference (Boley conference room, 12 noon, last Tues each month).

**RESIDENT EVALUATION**
- The residents must show active participation in the laboratory.
- The resident must show improvement in knowledge of infectious diseases and laboratory skills.
- The resident must be reliable and responsible for clinical case presentations.
- The resident should assist the clinicians, to the extent possible for their level of training, on infectious disease rounds.
RECOMMENDED GENERAL MICROBIOLOGY READING LIST:


RECOMMENDED CLINICAL IMMUNOLOGY READING LIST:

Participation in management and informatics training and activities is an essential part of the resident's curriculum. Management and informatics activities are not limited to this single rotation, but should be considered to span the entire length of the residency. The activities are present in both the laboratory and administrative areas of the department. Participation in these activities from both perspectives will provide insight into the pathologist's role, and the administrative staff role in the operational aspects of running a department of laboratory informatics.

<table>
<thead>
<tr>
<th>Legend for Learning Activities for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
</tr>
<tr>
<td>Faculty sign-out</td>
</tr>
<tr>
<td>Journal club</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
</tr>
<tr>
<td>Role modeling</td>
</tr>
<tr>
<td>Lab inspections</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
</tr>
<tr>
<td>Online tools</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
</tr>
<tr>
<td>Project</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Legend for Evaluation Methods for Residents</th>
</tr>
</thead>
<tbody>
<tr>
<td>Report review</td>
</tr>
<tr>
<td>Direct observation</td>
</tr>
<tr>
<td>Checklist</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
</tr>
<tr>
<td>Standardized exam</td>
</tr>
<tr>
<td>Practical slide exam</td>
</tr>
<tr>
<td>In-house written exam</td>
</tr>
<tr>
<td>360 multisource rating</td>
</tr>
<tr>
<td>Portfolios</td>
</tr>
<tr>
<td>Procedures and case logs</td>
</tr>
</tbody>
</table>
### LABORATORY MANAGEMENT CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of management skills to provide appropriate and effective leadership in anatomic and laboratory medicine.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gain knowledge and skill in laboratory organizational management and leadership.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Gain knowledge and skills in laboratory finances.</td>
<td>DL, DSP RM, IC</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop skills in regulatory issues as they pertain to laboratory medicine.</td>
<td>DL, DSP RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>

### LABORATORY MANAGEMENT CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to pathology.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in anatomic and clinical pathology.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>DO, GR/FE, SE</td>
</tr>
</tbody>
</table>
| Understand the principles of human resources management, such as:  
  - Basic job interview skills.  
  - Determining staffing requirements and productivity.  
  - Ensuring competency of personnel.  
  - Strategies for employee recruitment, selection, and retention.  
  - Procedures for progressive discipline.  
  - The termination process.  
  - Legal protection of employees covering leave, benefits, retirement, discrimination, affirmative action, civil rights, and disabilities. | DL, JC, DSP, RM, IC | DO, GR/FE, SE |
Understand laboratory computer operations, including:

- Processes and content of information flow from order through result reporting.
- Laboratory design, layout, and instrument selection and their effect on productivity and cost.
- Effectiveness.
- Management of point of care testing.
- Use of policy and procedure manuals, NCCLS guidelines.
- Application of constructive conflict resolution and problem solving techniques in responding to.
- Concerns about department services.
- Principles and practice of quality management and performance improvement (CQI, PI), quality assurance and utilization management.

<table>
<thead>
<tr>
<th>Understand finance, accounting and reimbursement terms and their use:</th>
</tr>
</thead>
<tbody>
<tr>
<td>- The role of budgetary planning, managing, and controlling laboratory operations.</td>
</tr>
<tr>
<td>- Concepts of profit center, cost center, cost accounting, and capitation.</td>
</tr>
<tr>
<td>- Basic accounting including income, expenses, profit, as presented in an operating statement.</td>
</tr>
<tr>
<td>- Assets and liabilities as presented in a balance sheet.</td>
</tr>
<tr>
<td>- Use of benchmark tools for productivity and financial analysis.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required.</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations.</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations.</th>
</tr>
</thead>
</table>

| DL, JC, DSP, RM, IC | DO, GR/FE, SE |
| DL, JC, RM, IC | DO, GR/FE, SE, IWE |
| DL, JC, RM | DO, CL, GR/FE, SE, IWE |
| DL, RM, LI | DO, GR/FE, SE, IWE |
| DL, RM, LI, OT | DO, GR/FE, SE, IWE |
### LABORATORY MANAGEMENT CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
*Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>JC, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions.</td>
<td>JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel.</td>
<td>RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use proficiency programs to improve laboratory practices.</td>
<td>RM, LI, OT</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

### LABORATORY MANAGEMENT CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients’ families.*

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the laboratory service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Demonstrate skills in obtaining informed consent, including effective communication to patients about procedures, alternative approaches, and possible complications of laboratory-based patient care diagnostic and therapeutic activities, such as those related to chemistry.  

Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/ interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.

<table>
<thead>
<tr>
<th>LABORATORY MANAGEMENT CORE COMPETENCY: PROFESSIONALISM</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Goal:</strong> Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>DL, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research.</td>
<td>DL, OT</td>
<td>GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.

<table>
<thead>
<tr>
<th>LABORATORY MANAGEMENT CORE COMPETENCY: SYSTEM-BASED PRACTICE</th>
</tr>
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</table>

**Goal:**
Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.

<table>
<thead>
<tr>
<th>Objectives:</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate understanding of the role of the laboratory in the healthcare system.</td>
<td>DL, LI, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians.</td>
<td>RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of basic healthcare reimbursement methods.</td>
<td>DL, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME.</td>
<td>DL, LI, OT</td>
<td>DO, CL, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Understand and implement policies to continually improve patient safety as they relate to the clinical laboratory.</td>
<td>RM, LI, P</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

ADDITIONAL SPECIFIC OBJECTIVES IN LABORATORY MANAGEMENT

Pathology Milestones
PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice

For expanded definitions, see pages 30-31.

Management of Informatics Organizational and Leadership Skills

Skill Level I
- Understand the fundamental principles of human behavior in organizations, of management structure and function, and of organizational structures. Compare and contrast the structure of differing practice settings (e.g., hospital-based, specialty practice, independent laboratory, etc.). [PBL, ICS, PRO, SBP]

KUMC Pathology Residency Manual
- Develop the interpersonal skills required to effectively manage, lead, and motivate others, including professional peers. [PBL, ICS, PRO, SBP]
- Develop an understanding of the role of ethics in medical and managerial decision-making. [PBL, PRO, SBP]
- Appreciate the conflicting responsibilities and rewards of pathologists, administrators, and technologists, and even the competing interests within each group as necessary to the positive functioning of the laboratory. [PBL, PRO, SBP]
- Understand the nature of the relationships between pathologists, hospitals, and medical staffs, including a basic understanding of contracts, decision-making, and effective negotiation. [PBL, ICS, PRO, SBP]
- Develop skills to project an environment of patient-oriented and ethical service. [PC, MK, PBL, ICS, PRO, SBP]
- Understand the organization of the laboratory, including preanalytical sample acquisition, accessioning and processing, structure of analytical units, and postanalytical sample result. Recognize the different skill sets required of personnel in all of these areas. Be able to analyze work flow in the laboratory. [PC, MK, PBL, PRO, SBP]

**Skill Level II**
- Understand human resource systems, including effective processes for recruitment, retention, and performance management of technical and professional staff. [PBL, SBP]

**Financial Skills**

**Skill Level I**
- Understand the fundamentals of financial data collection and financial statement presentation and analysis. [PBL, SBP]
- Understand the role of the budget process for operational planning, managing, and control. [PBL, SBP]
- Understand how to properly assign Current Procedural Terminology (CPT) codes to procedures in the laboratory. [PC, MK, PBL]

**Skill Level II**
- Understand how to assess the need for new instrumentation as well as the process of financial justification of capital equipment investments such as these. [PBL, SBP]
- Understand the nature and behavior of costs in the laboratory, including test-cost accounting. [PBL, SBP]
- Understand the applicable forms and requirements of reimbursement, particularly Medicare reimbursement, for both clinical laboratories and pathologists. [PBL, SBP]
- Understand how to monitor utilization and become familiar with strategies to effectively manage utilization in a healthcare organization. [PBL, SBP]
Regulatory Skills

Skill Level I

• Become familiar with the accrediting agencies relevant to laboratory certification and licensure [e.g., CAP, AABB, Occupational Health and Safety Administration, CMS, Clinical Laboratory Improvement Amendments (CLIA), and TJC], and participate in at least one CAP “mock” or “self-inspection” of the laboratory. [PC, MK, PBL, ICS, PRO, SBP]

• Become familiar with the “test complexity” models under CLIA for clinical laboratory tests (i.e., high complexity, moderate complexity, waived, and physician-performed microscopy). [PC, MK, PBL, SBP]

• Understand the regulatory and compliance environment for laboratories, including CLIA and the Office of the Inspector General model compliance plan, and the implications that these have for the laboratory management team. [PC, MK, PBL, SBP]

• Become familiar with the patient privacy and data security requirements of the HIPAA, including the use of institutional review board (IRB) protocols for conducting clinical research. [PC, MK, PBL, SBP]

• Understand training, certification, licensing, and competency assessment standards for laboratory professionals, including medical technologists and medical laboratory technicians. [PBL, SBP]

• Understand the importance of a comprehensive laboratory safety policy and program. [PBL, SBP]

• Understand how Standard Operating Procedures (SOPs) are used in the routine operation of clinical laboratories. [PC, MK, PBL, SBP]

• Understand how SOPs are developed, authored, and reviewed and their importance in mandatory laboratory inspection by various accrediting agencies (e.g., CAP, TJC, and AABB). [PC, MK, PBL, ICS, SBP]

Skill Level II

• Understand the role of risk management in the laboratory and become familiar with the nature of medical malpractice, patient safety initiatives, institutional risk mitigation, and forensic testing. [PBL, PRO, SBP]

• Become familiar with the process of long-range planning and strategic management, and the implications that this process has for successful management. [PBL, SBP]

• Become familiar with the fundamental principles of marketing, sales, and a market-oriented service delivery strategy. [PBL, SBP]

• Become familiar with the process for creating and/or critically reviewing a business plan for a new or proposed service. [PBL, SBP]

• Become familiar with the different forms that practice relationships can take (e.g., sole proprietorship, partnership, and corporation), and the advantages and disadvantages of each. [PBL, SBP]

• Participate in the development and authorship, and/or review and revision of SOPs. [PC, MK, PBL, ICS, PRO, SBP]
Quality Assurance, Quality Control, Pre-and Postanalytic Management

Skill Level I

- Understand the role of quality assurance, quality management, and process improvement principles in laboratory operation and planning. [PBL, SBP]
- Understand the role of interlaboratory proficiency surveys, such as the CAP proficiency surveys. [PC, MK, PBL, SBP]
- Be able to develop templates for introduction of new analyte testing in the clinical laboratory, with defined responsibilities at each level of personnel functions. [PC, MK, PBL, ICS, SBP]
- Know fundamental statistical concepts for laboratory diagnostics, including descriptive methods, inference regarding population means, confidence intervals, parametric and nonparametric statistics, measures of variance and error, sources of analytical error, methodologic bias, ROC curves, Bayes theorem, reportable range, analytical range, and linearity. Utilize these methodologies to select and validate new diagnostic tests and analytical methods. [PC, MK, SBP]
- Understand principles of specimen collection (e.g., phlebotomy technique, safety, and specimen tubes) and specimen processing. [PC, MK, SBP]
- Recognize sources of preanalytical variation and the role of biological variability in laboratory assessment. [PC, MK, PBL, SBP]
- Know how to employ appropriate use of delta checks in detecting preanalytical, analytical, and postanalytical errors. [PC, MK, PBL, SBP]
- Understand the principles of postanalytical result processing and data delivery (see also the Informatics section). [PC, MK, PBL]

Skill Level II

- Understand the principles involved in determination of reference ranges and the limitations of reference range determinations. [PC, MK, PBL]
- Understand how to choose, use, and monitor the performance of reference laboratories. [PC, MK, PBL, SBP]

COMPETENCIES SPECIFIC TO LABORATORY MANAGEMENT

Medical Knowledge

- Understand the most common forms of clinical laboratory organizational structure.
- Understand management theory and the difference between leadership and management.
- Understand the general elements of an income statement and balance sheet.
- Understand the basic approach to creating a budget for the clinical laboratory.
- Be able to assign correct CPT codes for common pathology and laboratory medicine procedures.
- Understand the basic elements of the laboratory safety program.
- Understand the essential elements of choosing a reference laboratory.
- Understand the necessary elements of test cost accounting in the laboratory and be able to cost account a common laboratory procedure.
- Understand basic elements of billing and reimbursement alongside compliance law.
• Understand how to perform a new instrument evaluation and prepare a financial justification analysis.
• Be able to conduct a performance appraisal.
• Understand the necessary elements of a risk management program and be able to describe how to effectively manage an incident.
• Be able to conduct a management meeting within the laboratory.
• Know how to review external proficiency surveys and respond to identified problems or questions.
• Be able to design a program for test evaluation and validation.
• Be able to participate in a quality process improvement project.
• Understand how to seek and obtain IRB approval for clinical research studies.

Practice-Based Learning and Improvement

• Be able to perform a CAP self-inspection or mock inspection.
• Understand the basic elements of the model compliance plan for laboratories.
• Understand the basic elements of the strategic planning process.
• Be able to participate in a quality process improvement process.

Interpersonal and Communication Skills

• Understand how to conduct an interview for a new employee.

Systems-Based Practice

• Understand the differences between different forms of professional practice.
• Understand the essential elements of professional employment and practice group contracts.
• Understand how to develop a business plan, together with a marketing and sales plan for hospital laboratory outreach program.

RECOMMENDED READING LIST:

8. CMS. Laboratory Director Responsibilities. CLIA regulations, 2003.
INFORMATICS OBJECTIVES

**Basic Computer Skills**

**Skill Level I**
- Understand terms and concepts related to computer hardware and software. [PBL, SBP]
- Understand basic computer networking concepts. [PBL, SBP]
- Understand how to use word processing, spreadsheet, presentation graphics, and statistical software. [PBL, SBP]

**Laboratory Information System Concepts**

**Skill Level I**
- Understand the major features of a laboratory information system. [PBL, SBP]
- Know the basic data elements of a laboratory information system. [PBL, SBP]
- Be able to extract data from the laboratory information system. [PBL, ICS, SBP]

**Security and Privacy**

**Skill Level I**
- Understand HIPAA guidelines for security and privacy of protected health information. [PC, MK, PBL, SBP]

**The Internet and World Wide Web**

**Skill Level I**
- Know internet-related terms and concepts. [PBL, ICS, SBP]
- Be able to utilize the internet to access internet-based databases.
- Perform literature searches. [PC, MK, PBL, ICS, SBP]

**Communication and Standards**

**Skill Level I**
- Develop basic understanding of how the laboratory information system shares data with other networked systems within the enterprise. [PBL, ICS, SBP]

**Skill Level II**
- Develop basic understanding of laboratory instrument interfaces. [PC, MK, PBL, SBP]
- Understand data standards and encoding schemes, such as Health Level Seven (HL7), Logical Observation Identifier Names and Codes (LOINC), Systematized Nomenclature of Medicine by the CAP (SNOMED), International Classification of Diseases (ICD-9 and ICD10), and CPT. [PC, MK, PBL, SBP]
Emerging Technologies

Skill Level II

- Develop a basic understanding of telepathology systems and concepts. [PC, MK, PBL, ICS, SBP]
- Develop a basic understanding of bioinformatics concepts with an emphasis on the critical evaluation of evolving bioinformatics tools. [PC, MK, PBL, ICS, PRO, SBP]
- Develop a basic understanding of evolving multiparameter diagnostic approaches. [PC, MK, PBL, SBP]

<table>
<thead>
<tr>
<th>PGY 1 GOALS: By the end of the first year:</th>
</tr>
</thead>
<tbody>
<tr>
<td>The resident should achieve competency in Skill Level 1.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>PGY 2 GOALS: By the end of the second year:</th>
</tr>
</thead>
<tbody>
<tr>
<td>The resident should achieve competency in Skill Level 1 and some of Level 2.</td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>PGY 3 GOALS: By the end of the third year:</th>
</tr>
</thead>
<tbody>
<tr>
<td>The resident should achieve competency in Skill Level 1 and most of Level 2.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>PGY 4 GOALS:</th>
</tr>
</thead>
<tbody>
<tr>
<td>The resident should be competent in all Skills Levels 1 and 2 for laboratory management and informatics.</td>
</tr>
</tbody>
</table>

GENERAL

Throughout the entire duration of residency training the resident must also demonstrate the specific skills for Professionalism, Practice--Based Learning and Improvement, Interpersonal and Communication Skills and System-Based Learning as listed on pages 19-20.

DUTIES AND RESPONSIBILITIES OF THE RESIDENT

- Become familiar with the schedule of the formal activities cited above and to participate on a regular basis.
- Work with the appropriate faculty/administrative and management staff to become involved in the management and informatics activities in the department.
- Ascertain the specific management/informatics responsibilities associated with each service and to meet those responsibilities in a timely fashion.
- Attend all administrative meetings with the laboratory director.
- Participation in quality assurance and quality improvement activities is an essential part of the resident's curriculum. Quality assurance (QA) and quality improvement (QI) activities are not limited to a single rotation; but rather span the entire length of the residency. Residents in all clinical pathology rotations must attend monthly QI meetings. The activities are present in both the medical and administrative areas of the department.

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KUMC Pathology Residency Manual
Participation in these activities from both perspectives will provide insight into the pathologist's role and the Department's role in hospital QA/QI programs. During the first three months of starting the residency, a session is held with the administrative lab director. At this time the resident is given an overview of the QA/QI activities of the Department and the expectations of the resident's participation in those activities is outlined.
INTEGRATED CLINICAL PATHOLOGY - VAMC

INTEGRATED CLINICAL PATHOLOGY FACULTY (VA MEDICAL CENTER)

Sharad Mathur, MD   Associate Professor and Chief of Pathology and Laboratory Medicine
Ozlem Ulusarac, MD, PhD  Director of Chemistry, Immunology, and Microbiology

The integrated clinical pathology rotation is offered at the VA Medical Center. It is designed for PGY-2 or higher residents who have had initial rotations in different areas of clinical pathology. The rotation aims to expose the resident to various aspects of management and service work in an integrated clinical pathology laboratory. Residents will assume graded responsibility in subsequent months on this rotation.

Legend for Learning Activities for Residents

<table>
<thead>
<tr>
<th>Activity</th>
<th>Code</th>
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</thead>
<tbody>
<tr>
<td>Didactic lecture</td>
<td>DL</td>
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<tr>
<td>Faculty sign-out</td>
<td>FSO</td>
</tr>
<tr>
<td>Journal club</td>
<td>JC</td>
</tr>
<tr>
<td>Directly supervised procedure</td>
<td>DSP</td>
</tr>
<tr>
<td>Role modeling</td>
<td>RM</td>
</tr>
<tr>
<td>Lab inspections</td>
<td>LI</td>
</tr>
<tr>
<td>Interdisciplinary conference</td>
<td>IC</td>
</tr>
<tr>
<td>Online tools</td>
<td>OT</td>
</tr>
<tr>
<td>Unknown slide conferences</td>
<td>USC</td>
</tr>
<tr>
<td>Project</td>
<td>P</td>
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</table>

Legend for Evaluation Methods for Residents

<table>
<thead>
<tr>
<th>Method</th>
<th>Code</th>
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<tbody>
<tr>
<td>Report review</td>
<td>RR</td>
</tr>
<tr>
<td>Direct observation</td>
<td>DO</td>
</tr>
<tr>
<td>Checklist</td>
<td>CL</td>
</tr>
<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
</tr>
<tr>
<td>Standardized exam</td>
<td>SE</td>
</tr>
<tr>
<td>Practical slide exam</td>
<td>PSE</td>
</tr>
<tr>
<td>In-house written exam</td>
<td>IWE</td>
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<tr>
<td>360 multisource rating</td>
<td>360</td>
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<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
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</tbody>
</table>
### INTEGRATED CLINICAL PATH CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of diagnostic competence and the ability to provide appropriate and effective consultation in the context of clinical laboratory services.*

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gather essential and accurate information about patients using all relevant available modalities.</td>
<td>FSO, DSP, RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Act as a skilled consultant to other clinicians to develop a diagnostic plan based on specific clinical questions and relevant clinical and pathologic information. This should be accomplished both in the patient-specific setting and the broader context of developing appropriate clinical pathway algorithms for diagnosis.</td>
<td>DL, FSO, DSP, RM, IC</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Gain knowledge and technical skills to recognize, interpret, and explain pathologic processes in the clinical practice of laboratory medicine.</td>
<td>FSO, DL, JC, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as part of a multidisciplinary healthcare team in developing a therapeutic plan that includes laboratory monitoring of efficacy and toxicity.</td>
<td>DL, JC, DSP, RM, IC</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Provide expert consultation on the interpretation and follow-up of unusual or unexpected test results.</td>
<td>FSO, DL, DSP, RM</td>
<td>DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Consult as a clinical expert in laboratory medicine at multidisciplinary conferences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE, PCL</td>
</tr>
</tbody>
</table>

### INTEGRATED CLINICAL PATH CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive (e.g. epidemiological and social-behavioral) sciences and the application of this knowledge to laboratory medicine.*

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</thead>
<tbody>
<tr>
<td>Be able to use all relevant information resources to acquire and evaluate evidence-based information. Demonstrate proficiency in evaluating and presenting findings from appropriate peer-reviewed journals.</td>
<td>FSO, DL, JC, DSP, RM, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Develop and maintain a knowledge base in the basic and clinical sciences necessary for effective consultation in hematopathology.</td>
<td>DL, JC, SP, IC</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
<tr>
<td>Demonstrate sufficient knowledge to determine clinically optimal yet cost-effective testing and laboratory-based therapeutic strategies, including issues of turnaround time, test menu construction, and in-house referral diagnostic testing.</td>
<td>DL, JC, RM, IC, FSO</td>
<td>RR, DO, GR/FE, SE, IWE</td>
</tr>
</tbody>
</table>
Employ mathematics and statistics as appropriate to laboratory testing; understand and implement quality control (QC) and quality assurance procedures as required. | DL, JC, RM | DO, CL, GR/FE, SE, IWE |
---|---|---|
Recognize the unique aspects of laboratory medicine practice as modified by patient age and other patient population characteristics, especially aspects of pediatric and geriatric practice. | DL, JC, DSP, RM, IC | RR, DO, GR/FE, SE, IWE |
Demonstrate awareness and understanding of general and test-specific standards for method development and evaluation, such as those promulgated by the Clinical Laboratory Standards Institute (CLSI; formerly NCCLS), CAP, and similar organizations. | DL, RM, LI | DO, GR/FE, SE, IWE |
Demonstrate awareness and understanding of proficiency programs, such as those provided by CAP and similar organizations. | DL, RM, LI, OT | DO, GR/FE, SE, IWE |
Demonstrate knowledge of the principles of clinical research design, implementation, and interpretation. Understand the various levels of evidence in medicine and their translation into evidence-based practice. | JC, DSP, RM, P | DO, GR/FE, SE, IWE |
Be able to design a study that can be used to validate methodologies and parameters of clinical utility for the implementation and continuing use of new evidence-based analytes in the local setting. | DL, JC, DSP, RM, LI, P | DO, GR/FE, SE, IWE |

**INTEGRATED CLINICAL PATH CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT**

**Goal:**

*Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.*

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Demonstrate the ability to critically assess the scientific literature. | JC, RM | DO, GR/FE |
Demonstrate knowledge of evidence-based medicine and apply its principles in practice. | JC, RM | DO, GR/FE |
Use multiple sources, including information technology, to optimize lifelong learning and support patient care decisions. | JC, RM, OT | DO, GR/FE |
Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice. | DSP, RM | DO, GR/FE, SE, IWE |
Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety. | RM, IC | DO, GR/FE |
Demonstrate knowledge of how to establish continuing competency assessment for pathologists as well as for laboratory personnel. | RM, LI | DO, GR/FE |
Use proficiency programs to improve laboratory practices. | RM, LI, OT | DO, GR/FE |
### INTEGRATED CLINICAL PATH CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**  
*Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients’ families.*

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<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate the ability to communicate the vision of the microbiology/immunology service role to other clinicians as well as to other healthcare personnel and administrators to develop clinically advantageous and cost-effective strategies.</td>
<td>RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>RM, IC</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate skills in educating colleagues and other healthcare professionals: (1) demonstrate the ability to help other residents obtain proficiency in laboratory medicine; (2) demonstrate the ability to work well with technologists and to present laboratory medicine concepts to them effectively in continuing education settings and in the day-to-day laboratory environment; (3) demonstrate the ability to educate non-pathology clinicians and other healthcare workers, including pharmacists, nurses, residents, medical students, and others, about topics such as the fundamental principles of pathophysiology underlying test design/interpretation and the approach to choosing and interpreting laboratory tests; (4) demonstrate an understanding of the principles one must follow when educating other practicing pathologists through publications or seminars on new testing and therapeutic strategies, research discoveries, and other cutting-edge professional knowledge.</td>
<td>JC, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
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### INTEGRATED CLINICAL PATH CORE COMPETENCY: PROFESSIONALISM

**Goal:**  
* Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.

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<th>Evaluation Activities</th>
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</thead>
<tbody>
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<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
<td>DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Demonstrate positive work habits, including punctuality, dependability, and professional appearance. | RM | DO, GR/FE |
---|---|---|
Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest. | DSP, RM | DO, GR/FE |
Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter. | DL, DSP, RM, IC | DO, GR/FE |
Demonstrate knowledge of regulatory issues pertaining to the use of human subjects in research. | DL, OT | GR/FE, SE |
Demonstrate a commitment to excellence and ongoing professional development. | RM | DO, GR/FE |
Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team. | DSP, RM, IC | DO, GR/FE |

### INTEGRATED CLINICAL PATH CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

| Objectives: | Learning Activities | Evaluation Activities |
---|---|---|
Demonstrate understanding of the role of the microbiology/immunology laboratory in the healthcare system. | DL, LI, IC | DO, GR/FE |
Demonstrate the ability to design resource-effective diagnostic plans based on knowledge of best practices in collaboration with other clinicians. | RM | DO, GR/FE, SE, IWE |
Demonstrate knowledge of basic healthcare reimbursement methods. | DL | DO, GR/FE, SE, IWE |
Demonstrate knowledge of the laboratory regulatory environment, including licensing authorities; federal, state, and local public health rules and regulations; regulatory agencies such as the Centers for Medicare and Medicaid Services and the US Food and Drug Administration; and accrediting agencies such as The Joint Commission (TJC), CAP, and the ACGME. | DL, FSO, LI, OT | DO, GR/FE, SE, IWE |
Understand and implement policies to continually improve patient safety as they relate to the clinical laboratory. | FSO, RM, LI, P | DO, GR/FE |
ADDITIONAL SPECIFIC OBJECTIVES

[Pathology Milestones]
PC = Patient Care  
MK = Medical Knowledge  
PBL = Problem Based Learning  
ICS = Interpersonal and Communication Skills  
PRO = Professionalism  
SBP = Systems Based Practice  
For expanded definitions, see pages 30-31.

Chemistry

Skill Level II
- Understand the general principles of assay calibration, QC, and the need for calibration verification. [PC, MK, PBL]
- Understand the causes of both positive and negative interferences as well as how to detect and avoid them. [PC, MK]
- Identify factors influencing separation and resolution in electrophoresis and chromatography, including mechanism of separation and mobile/stationary phases. [PC, MK]

Hematology

Skill Level II
- Interpret results of automated and manual cell counts and understand the relevant technical limitations. [PC, MK, PBL]
- Recommend appropriate steps for abnormal sample processing, analysis, and result reporting. [PC, MK, PBL, ICS, SBP]
- Review abnormal results and correlate results with peripheral blood smear findings and clinical history. [PC, MK]
- Recognize abnormal RBC, WBC, and platelet morphology; formulate a differential diagnosis and suggest appropriate laboratory testing for follow-up. [PC, MK]
- Recognize technical artifacts in WBC, RBC, and platelet morphology. [PC, MK]
- Recognize infectious disorders that can be diagnosed by blood smear. [PC, MK]
- Correlate peripheral blood smear findings with bone marrow morphology. [PC, MK]
- Interpret results of body fluid analyses in the appropriate clinical context. [PC, MK, SBP]
- Recognize malignant cells and recommend appropriate confirmatory tests. [PC, MK]
- Correlate abnormal body fluid cell morphology with cytology, flow cytometry, and other relevant diagnostic test results. [PC, MK, SBP]
- Identify body fluid crystals. Distinguish between urate and calcium pyrophosphate crystals, using polarized light. [PC, MK]
• Understand the pathophysiology, clinical findings, etiology, and expected bone marrow morphology for vitamin deficiency anemias, hemoglobinopathies, thalassemias, aplastic anemia, red cell aplasia, leukemias, myeloproliferative disorders, myelodysplastic syndromes, plasma cell dyscrasias, and mast cell diseases. [PC, MK]

• Integrate morphology, cytochemistry, immunophenotype, and molecular and cytogenetics in the differential diagnosis of acute and chronic leukemia, lymphoma, and myeloproliferative and myelodysplastic diseases. [PC, MK, SBP]

• Integrate peripheral blood smear and bone marrow findings and render a preliminary diagnosis. [PC, MK, PBL, ICS, SBP]

• Know the post-therapy findings seen after treatment for leukemia and the temporal relationships to marrow regeneration post therapy. [PC, MK, SBP]

• Recognize the bone marrow manifestations of infections (e.g., viral, fungal, and hemophagocytic syndromes). [PC, MK]

• Recognize the bone marrow manifestations of noninfectious systemic diseases (e.g., alcoholism, collagen vascular disease, and nonhematologic malignancies). [PC, MK]

Microbiology

Skill Level II

• Know the media used for isolation of less common or fastidious bacteria. [PC, MK]

• Understand the advantages and disadvantages of methods used to identify bacteria, including automated systems and manual methods (including biochemical reactions such as oxidase, catalase, PYR, lactose fermentation, and metabolism of glucose and other carbohydrates). [PC, MK]

• Acquire advanced skills in microscopy, including the ability to read and interpret respiratory and wound Gram stains and fluorescent stains. [PC, MK]

Transfusion Medicine

Skill Level II

• Demonstrate proficiency in evaluating and recommending treatment plans for complex transfusion reactions. [PC, MK, PBL, ICS, PRO, SBP]

• Demonstrate familiarity with the appropriate use of highly specialized blood components. [PC, MK, SBP]

• Compare and contrast the various means of performing blood utilization reviews. [PC, MK, PBL, SBP]

Laboratory Management

Skill Level I

• Appreciate the conflicting responsibilities and rewards of pathologists, administrators, and technologists, and even the competing interests within each group as necessary to the positive functioning of the laboratory. [PBL, ICS, PRO, SBP]

• Understand the nature of the relationships between pathologists, hospitals, and medical staffs, including a
basic understanding of contracts, decision-making, and effective negotiation. [PBL, ICS, PRO, SBP]

• Develop skills to project an environment of patient-oriented and ethical service. [PC, MK, PBL, ICS, PRO, SBP]

• Understand the organization for the laboratory, including preanalytical sample acquisition, accessioning and processing, structure of analytical units, and postanalytical sample resulting. [PC, MK]

• Recognize the different skill sets required of personnel in all of these areas. Be able to analyze work flow in the laboratory. [PC, MK, PBL]

• Understand the role of quality assurance, quality management, and process improvement principles in laboratory operation and planning. [PC, MK, PBL, SBP]

• Understand the role of interlaboratory proficiency surveys, such as the CAP proficiency surveys. [PC, MK, PBL, SBP]

• Recognize sources of preanalytical variation and the role of biological variability in laboratory assessment. [PC, MK, PBL, SBP]

• Know how to employ appropriate use of delta checks in detecting preanalytical, analytical, and postanalytical errors. [PC, MK, PBL, SBP]

Skill Level II

• Understand the nature and behavior of costs in the laboratory, including test-cost accounting. [PBL, SBP]

• Understand how to monitor utilization and become familiar with strategies to effectively manage utilization in a healthcare organization. [PBL, ICS, PRO, SBP]

• Be able to participate in a quality improvement process. [PBL, ICS, PRO, SBP]

ADDITIONAL SPECIFIC SKILLS

• Evaluate abnormal peripheral blood smears (Wright-Giemsa stain). [PC, MK]

• Evaluate abnormal body fluid specimens (Wright-Giemsa stain). [PC, MK]

• Evaluate bone marrow aspiration and biopsy specimens including. [PC, MK]

• Wright-Giemsa stained peripheral blood smears, bone marrow aspirate smears, and bone marrow biopsy touch imprints.

• H&E stained bone marrow biopsy and aspirate clot sections.

• Iron stained aspirate smear and clot section.

• Reticulin stained biopsy section.

• Other special and/or immunohistochemical stains.

• Prepare a written bone marrow report including results of iron, reticulin, and other special stains, immunohistochemical stains, flow cytometric results, and cytogenetic and molecular genetic results.

• Know the guidelines for utilization of various blood products. [PC, MK]

• Assess appropriateness of blood product orders using MSBOS and blood product utilization guidelines. [PC, MK, PBL, SBP]

• Assess appropriateness of initial requests for irradiated, CMV-negative, and other special blood products. [PC, MK, SBP]
• Evaluate suspected transfusion reactions. [PC, MK, SBP]
• Prepare a written transfusion reaction report incorporating clinical and laboratory information and provide recommendations. [PC, MK, PBL, ICS, PRO, SBP]
• Evaluate unusual serologic results in the blood bank and recommend further work-up. [PC, MK]
• Understand the function of the Transfusion Committee (TC). [PBL, SBP]
• Work-up and present transfusion audit cases at TC meetings. [PC, MK, PBL, ICS, PRO, SBP]
• Understand the utility and principle of routine media used in the clinical microbiology lab. [PC, MK]
• Understand the rationale for selecting particular media for different specimens. [PC, MK]
• Observe and understand the process of receiving, accessioning and plating a specimen for culture. [PC, MK]
• Understand the principle and procedure for rapid tests used in the clinical microbiology lab. [PC, MK]
• Understand principle and procedure for screening sputum specimens for culture. [PC, MK]
• Understand the principle and procedure for Gram stain procedure. [PC, MK]
• Understand the principle of the growth detection system for blood culture bottles. [PC, MK]
• Understand the principle and procedure for performing Vitek and Vitek-MS identification and susceptibility systems. [PC, MK]
• Understand the function of the Infection Control Committee. [PC, MK, PBL, SBP]
• Interpret results of serum and urine protein electrophoresis and immunofixation studies and prepare written reports. [PC, MK, PBL, ICS, SBP]
• Understand the problems encountered in daily practice in various areas of the clinical pathology laboratory. [PC, MK, PBL, SBP]
• Understand the role of the clinical pathology laboratory and the clinical pathologist in providing optimal patient care services in a cost efficient manner. [PBL, SBP]
• Critically analyze laboratory utilization practices and assess appropriateness of reference laboratory (send out) test orders. [PBL, SBP]
• Provide consultation to clinical colleagues about issues related to test ordering, performance characteristics of tests, and interpretation of test results in hematology, coagulation, chemistry, immunology, therapeutic drug monitoring, and microbiology. [PC, MK, PBL, ICS, PRO, SBP]

DUTIES AND RESPONSIBILITIES OF THE RESIDENT

• Be present in the laboratory or be available on pager between 8 a.m. and 5 p.m., weekdays.
• Review bone marrow, peripheral smear, and body fluid cases with the staff pathologist.
• Review electrophoresis results with staff pathologist.
• Review transfusion reactions with staff pathologist.
• Review reference laboratory test orders for appropriateness and medical necessity.
• Attend Transfusion Committee meeting (TC) (monthly) and present transfusion audit cases.
• Attend Infection Control Committee meeting (monthly) and assist staff microbiologist in addressing laboratory issues.
• Attend Quality Improvement Committee meeting (monthly).
• Attend Pathology-Oncology-Surgery Conference (monthly).
• Attend Cancer Conference (weekly).
• Complete learning objectives stated above through participation in laboratory work, attendance at departmental and interdepartmental meetings, review of textbooks and journal articles, and participation in laboratory studies.
• Prepare and present an in-service on a clinical pathology topic.
• Review and prepare a study guide for one chapter of Henry’s CP book.

Resident Recommendations

Reference laboratory test orders
1. When reviewing reference laboratory tests, look up the patient history in CPRS to determine whether the test is medically appropriate and necessary.
2. If the resident is unsure about the nature/specifics of the test, the resident can look up the test on the Quest diagnostics website (http://www.questdiagnostics.com/testcenter/testguide.action). This will give the resident clinical indications, method, and interpretive information for the test.
3. Clinlab Navigator (http://www.clinlabnavigator.com/) is also a good resource for lab test indications and interpretation.

Serum and Urine Protein Electrophoresis
• For every patient, look up previous SPEP and UPEP results.
  o In CPRS, go to labs, look up cumulative labs, select “all results”, then select SPEP or UPEP to see the previous results.
• Also go to the problems tab to look for possible causes of SPEP/UPEP abnormalities.
  o Diabetes, neuropathy, renal disease, hematologic malignancies, etc.
• Examine the SPEP gel and printouts to look for monoclonal spikes, decreased albumin, increased or decreased immunoglobulins.

General Chemistry
• Aside from reviewing reference laboratory orders and electrophoresis cases, it is a good idea to use resident free time to do some chemistry studying.
• Both Tietz and Henry are large, meaty reference books that are great when the resident has a specific question that needs to be answered.
• For a more broad chemistry overview, study companion and question books provide a great introduction and outline to the information that is pertinent for both the board examination and the RISE.
  o ASCP Quick Compendium and the Quick Compendium Companion.
  o Appleton & Lange’s Outline Review Clinical Chemistry.

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Additionally, there are many electronic and online resources that are helpful to review.

- Mayo Clinic teleconferences – several of these are available with Dr. Tilzer.
- American Association for Clinical Chemistry (AACC) has numerous resources for resident education including case studies, journal clubs, pearls of laboratory medicine, and podcasts. ([http://www.aacc.org/publications/clin_chem/clinchemtraincouncil/Pages/Default.a](http://www.aacc.org/publications/clin_chem/clinchemtraincouncil/Pages/Default.a)).
- Clinlab Navigator ([http://www.clinlabnavigator.com/](http://www.clinlabnavigator.com/)) has brief descriptions of tests and test utilization as well as excellent resources for transfusion medicine.
- American Society for Clinical Pathology (ASCP) and College of American Pathologists (CAP) have resources for residents as well. They also offer CP educational resources for purchase.

Hematology

- Review peripheral smears and examine the patient's history for possible causes of any abnormal findings.
- Review and count bone marrow cases and fill out the bone marrow examination worksheet.
- Prepare for sign-out by reading about resident cases in the 2008 WHO book and Foucar textbooks.

Blood Banking

- To prepare for the monthly TC meeting, get the list of patients from Dr. Mathur and look up their histories in CPRS.
- Document any indications for transfusion (cardiac history, active bleeding, intraoperative transfusion, symptomatic anemia, etc.).
- Document the CBC trends including admission values, point of transfusion, and post transfusion trends.
- Be prepared to briefly present the patient and pertinent issues and to discuss the appropriateness of the transfusion.
- Clinlab Navigator ([http://www.clinlabnavigator.com/](http://www.clinlabnavigator.com/)) is a good resource for transfusion guidelines and blood banking questions.

Microbiology

- Good texts for the review of microbiology include: ASCP Quick Compendium and Koneman.
- Additionally, bacterial board review PowerPoints are available on the computer L drive in the “Resident Microbiology” folder.

Laboratory Management

- Read through the first section of Henry and relevant chapters of Quick Compendium.
- Resident should familiarize themselves with the following:
  - Six Sigma steps.
  - LEAN management.
  - Maslow's hierarchy.
Laboratory design and service models.
Laboratory regulations.
Laboratory related governmental and nongovernmental organizations.
Suggested guidelines for record/specimen retention.
The 4 categories of lab tests (CLIA standards).
Westgard Rules.
The three types of errors in laboratory medicine.
Workflow analysis.
Point-of-care laboratory regulations.
LIS standards.
Types of costs.
Reimbursement and coding.

RECOMMENDED READING LIST:


RESIDENT EVALUATION

- The resident must show active participation in the functions of the clinical pathology laboratory.
- The resident must be responsible for his/her duties and be available to the laboratory and clinical staff for consultations.
- The resident must show improvement of knowledge in the areas outlined above.
GOALS AND OBJECTIVES

The goal of the rotation in the cytogenetics laboratory is to become competent in the interpretation of basic chromosome abnormalities, and to learn consultative skills that aid the clinician in the diagnosis of chromosomal syndromes.

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<tr>
<th>Legend for Evaluation Methods for Residents</th>
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</thead>
<tbody>
<tr>
<td>Report review</td>
<td>RR</td>
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<tr>
<td>Direct observation</td>
<td>DO</td>
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<tr>
<td>Checklist</td>
<td>CL</td>
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<tr>
<td>Global rating/faculty evaluation</td>
<td>GR/FE</td>
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<tr>
<td>Standardized exam</td>
<td>SE</td>
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<tr>
<td>Practical slide exam</td>
<td>PSE</td>
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<tr>
<td>In-house written exam</td>
<td>IWE</td>
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<tr>
<td>360 multisource rating</td>
<td>360</td>
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<tr>
<td>Portfolios</td>
<td>PF</td>
</tr>
<tr>
<td>Procedures and case logs</td>
<td>PCL</td>
</tr>
</tbody>
</table>
### CYTOGENETICS CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of interpretive competence in cytogenetics.*

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<tr>
<th>Objectives:</th>
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</thead>
<tbody>
<tr>
<td>Correlate chromosomal abnormalities with specific hematologic disorders such as myelodysplastic syndromes, hematologic malignancies, and myeloproliferative disorders.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Recognize abnormal karyotyping in prenatal specimens, including, but not limited to, Turner syndrome, Klinefelter syndrome, trisomy 21, trisomy 13 and trisomy 18.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Understand the use of fluorescence in situ hybridization (FISH) analysis for common disorders involving aneuploidies, microdeletions, or chromosomal translocations, including hematologic disorders such as acute promyelocytic leukemia and chronic myelogenous leukemia.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Recognize the major chromosomal abnormalities and their association with congenital syndromes, human malignancies, and spontaneous abortion.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
</tbody>
</table>

### CYTOGENETICS CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive sciences and the application of this knowledge to cytogenetics.*

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<tr>
<th>Objectives:</th>
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<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Have awareness of sample types, preparation, and storage conditions for cytogenetic tests.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Understand sample preparation from peripheral blood, bone marrow, amniocytes, chorionic vili, skin, and products of conception for karyotyping.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Understand basic cell and tissue culture techniques, harvesting, slide preparation, banding, staining and microscopic analysis for karyotyping.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Have knowledge of different FISH probe strategies (dual fusion, break-apart, etc).</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
<tr>
<td>Recognize the major chromosomal abnormalities and their association with congenital syndromes, human malignancies, and spontaneous abortion.</td>
<td>FSO, RM, OT, DL</td>
<td>DO, GR/FE, IWE</td>
</tr>
</tbody>
</table>
## CYTOGENETICS CORE COMPETENCY: PRACTICE-BASED LEARNING & IMPROVEMENT

**Goal:**
Demonstrate the ability to investigate and evaluate their diagnostic and consultative practices, appraise and assimilate scientific evidence and improve their patient care practices.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
<th>Evaluation Activities</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demonstrate the ability to critically assess the scientific literature.</td>
<td>JC, RM, OT, P</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of evidence-based medicine and apply its principles in practice.</td>
<td>FSO, JC, RM, OT</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Develop personally effective strategies for the identification and remediation of gaps in medical knowledge needed for effective practice.</td>
<td>DSP, RM, USC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Use laboratory problems and clinical inquiries to identify process improvements to increase patient safety.</td>
<td>FSO, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

## CYTOGENETICS CORE COMPETENCY: INTERPERSONAL & COMMUNICATION SKILLS

**Goal:**
Demonstrate interpersonal and communication skills that result in effective information exchange and teaming with other healthcare providers, patients and patients' families.

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<thead>
<tr>
<th>Objectives</th>
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<tbody>
<tr>
<td>Demonstrate the ability to provide direct communication to the referring physician or appropriate clinical personnel when interpretation of a laboratory assay reveals an urgent, critical, or unexpected finding and document this communication in an appropriate fashion.</td>
<td>FSO, DSP, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Choose effective modes of communication (listening, nonverbal, explanatory, questioning) and mechanisms of communication (face-to-face, telephone, e-mail, written), as appropriate.</td>
<td>FSO, RM, IC, USC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Conduct both individual consultations and presentations at multidisciplinary conferences that are focused, clear, and concise.</td>
<td>RM, IC, USC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

## CYTOGENETICS CORE COMPETENCY: PROFESSIONALISM

**Goal:**
Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.

<table>
<thead>
<tr>
<th>Objectives</th>
<th>Learning Activities</th>
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<tbody>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>
Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.  FSO, DSP, RM, IC  DO, GR/FE

Demonstrate positive work habits, including punctuality, dependability, and professional appearance.  FSO, RM  DO, GR/FE

Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.  FSO, DSP, RM  DO, GR/FE

Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.  DL, FSO, DSP, RM, IC  DO, GR/FE

Demonstrate a commitment to excellence and ongoing professional development.  RM  DO, GR/FE

**CYTOGENETICS CORE COMPETENCY: SYSTEM-BASED PRACTICE**

**Goal:**
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

**Objectives:**

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<tr>
<td>DL, FSO, LI, IC</td>
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<tr>
<td>FSO, RM, LI</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>RM, LI, OT</td>
<td>DO, GR/FE</td>
</tr>
</tbody>
</table>

**SPECIFIC QUESTIONS TO BE DISCUSSED WITH THE DIRECTOR**

- **t(9;22)** - What diseases is this associated with? What percentage of cases within each disorder is it present? What is the clinical significance when it is present in each disease? What are the genes that are located at the breakpoints? What is the molecular basis of the abnormality? What is the major breakpoint and what is the minor breakpoint? What is Gleevec? What is the mechanism of action of Gleevec?
- **t(15;17)** - What disease is this associated with? What gene is located at the breakpoint on chromosome 17? How is this disorder treated? What is the prognosis?
- **t(8;14)** - What disorder(s) is this associated with? What genes are located at each breakpoint? What alternative translocations are associated with the t(8;14)? What is the molecular basis of the abnormality?
- **t(8;21)** - What disorder is this associated with? What genes are involved? What is the prognosis?
- **Inv(16)** - What disorder is this associated with? What genes are involved? What is the prognosis?
- **t(9;11)** - Associated disorder? What gene is located at 11q23? What is the prognosis?
- Inv(3)(q21q26.2) – prognosis?
- t(4;11) - Associated disorders? What is the prognosis?
- What is the most common situation in which FLT3, NPM1, and CEBPA mutation assays are ordered? What is the clinical significance of each mutation?
- KIT mutations are clinically significant in AMLs with what cytogenetic abnormalities? How does the KIT mutation change the prognosis in AML? What other disorders are KIT mutations seen in?
- Describe diseases with eosinophilia and abnormalities associated with PDGFRα, PDGFRβ, FGFR1.
- t(2;5) - Associated disorder? What is the important gene?
- t(11;14) - Associated disorders? What genes are located at each breakpoint? What are the functions of the genes?
- t(14;18) – What are the associated disorders? What genes are located at each breakpoint? What are the functions of the genes?
- t(11;18) – What is the associated disorder? What is the clinical significance?
- t(12;21) – What is the associated disorder? Associated genes? Prognosis?
- del(5q) – What is(are) the associated disorder(s)? What are the characteristics of the 5q- syndrome? Is 5q- found outside of the 5q- syndrome?
- -7 – What are the associated disorders? What is the prognosis?
- Loss of the Y chromosome - What is the clinical significance when seen in a bone marrow specimen?
- What are the most common additional chromosome abnormalities associated with CML in an accelerated phase or blast crisis?
- What chromosome abnormality is often associated with secondary AML following treatment with topoisomerase II inhibitors?
- What chromosome abnormalities are often associated with secondary AML following chemotherapy with alkylating agents?
- What are the cytogenetic and clinical prognostic factors associated with childhood ALL?
- What is Fanconi anemia? What chromosome abnormalities are associated with the disease? What assays are needed to diagnosis the chromosome abnormalities?
- What are the cytogenetic prognostic factors associated with multiple myeloma? What makes up a MM FISH panel?
- What are the cytogenetic prognostic factors associated with chronic lymphocytic leukemia? What makes up a CLL FISH panel?
- What is the UroVysion FISH assay used for?
- Describe the cytogenetic abnormalities associated with the following solid tumors:
  - Ewings sarcoma.
  - Alveolar rhabdomyosarcoma.
  - Myxoid liposarcoma.
  - Synovial sarcoma.
  - Alveolar soft part sarcoma.
• Clear cell sarcoma.
• Extraskeletal myxoid chondrosarcoma.

- What is the clinical significance of deletion of 1p and 19q in oligodendroglioma?
- What is the clinical significance of gene amplification for:
  - HER2 in breast carcinoma.
  - MYCN in neuroblastoma.
- Describe the use of the following FISH probes: MDM2, EWSR1, DDIT3, SS18/SYT, FOX01/FKHR, and FUS.
- Why is FISH for ALK and ROS1 performed in adenocarcinoma of the lung? What other molecular assay is also run in these tumors?
- Describe the clinical cytogenetic abnormalities and picture for Down, Edward and Patau syndromes.
- What other cytogenetic abnormality can cause Down syndrome (other than trisomy 21)? What is the importance in differentiating between the two?
- What are the common serum screening tests for Down syndrome?
- What is the incidence in the general population of a balanced translocation carrier? What is the significance to the carrier and to the family?
- Describe cell free fetal DNA screening and the pros and cons of the assay.
- Briefly describe the *Cri du Chat* syndrome.
- Briefly describe the deletion 22q11.2 syndrome.
- What is genomic imprinting? Describe how Prader-Willi and Angelman diseases are associated with genomic imprinting? How are ovarian teratomas and partial molar pregnancies related to genomic imprinting?
- Describe the cytogenetic and clinical pictures of Turner and Klinefelter syndrome.
- If a portion of a Y chromosome is present in a Turner patient, what is the patient at risk for developing?
- Describe the clinical, cytogenetic and molecular findings in Fragile X syndrome. What is a pre-mutation?
- Describe the role of chromosome abnormalities in spontaneous abortions.
- What is high resolution chromosome analysis?
- What is comparative genomic hybridization array analysis? How does it complement routine cytogenetic analysis?
- What is a break-apart FISH probe? What is a dual fusion FISH probe?

**DUTIES AND RESPONSIBILITIES OF THE RESIDENT**

- Set up, harvest, and analyze a blood specimen.
- Discuss above questions with director.
- Review each case coming through the lab during the month. Verify the correct test has been ordered and the correct procedure is being performed. Anticipate the possible cytogenetic findings. Be prepared to discuss the clinical significance of the findings. Sign each report below the director's signature line. Be present to discuss each case when director is signing out.
- Check in hematology and surgical pathology for diagnosis on all hematology cases coming through the lab. Be ready to discuss the relationship between the diagnosis and the cytogenetic result.
- Spend as much time as possible in the cytogenetics laboratory (if gone to a conference, leave page number with a tech). The resident will be assigned a desk in the analysis area for reading and where the resident can become involved with cases as they arrive and are being analyzed.
- The resident will be given both a pre-test (first day of rotation) and a post-test (last day of rotation) to determine the increased competency obtained during the rotation.

RECOMMENDED READING LIST:

The overall goal of the molecular pathology rotation is to become competent in the interpretation of molecular testing and to learn consultative skills that in the areas of molecular pathology, toxicology and histocompatibility testing. One rotation is spent at Children’s Mercy Hospital (three weeks in the molecular diagnostic laboratory and one week in clinical toxicology lab). Several days are spent in the KU Pathology Molecular Diagnostic Laboratory and/or the KU Hospital Molecular Diagnostic Laboratory (see Microbiology rotation description). And two to four days are spent at the Midwest Transplant Network to learn the testing concepts in histocompatibility testing for transplants.
## MOLECULAR PATHOLOGY CORE COMPETENCY: PATIENT CARE

**Goal:**
*Residents must demonstrate a satisfactory level of competence in molecular pathology, toxicology and histocompatibility testing.*

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<th>Objectives</th>
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<tbody>
<tr>
<td>Demonstrate a satisfactory level of competence in interpreting molecular test results (list of example molecular tests is below).</td>
<td>FSO, DSP, JC, RM, DL, IC</td>
<td>DO, GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate a satisfactory level of competence in interpreting toxicology test results (list of example molecular tests is below).</td>
<td>FSO, DSP, RM, DL</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a satisfactory level of competence in interpreting histocompatibility test results (list of example molecular tests is below).</td>
<td>FSO, DSP, RM, DL</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate competence in consultation for appropriate molecular pathology testing based on the clinical presentation.</td>
<td>FSO, DSP, JC RM, DL, IC</td>
<td>DO, GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate competence in consultation for appropriate toxicology testing based on the clinical presentation.</td>
<td>FSO, DSP, RM, DL</td>
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<tr>
<td>Demonstrate competence in consultation for appropriate histocompatibility testing based on the clinical presentation.</td>
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## MOLECULAR PATHOLOGY CORE COMPETENCY: MEDICAL KNOWLEDGE

**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive sciences and the application of this knowledge to molecular pathology.*

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<tbody>
<tr>
<td>Demonstrate basic knowledge in molecular testing techniques.</td>
<td>DL, JC, FSO, DSP, IC</td>
<td>DO, GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate knowledge of common genetic disorders and molecular tests associated with the disorders.</td>
<td>DL, FSO, DSP, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate knowledge of the use of appropriate molecular tests for neoplastic, infectious, and constitutional disorders.</td>
<td>DL, JC, FSO, DSP, IC</td>
<td>DO, GR/FE, SE</td>
</tr>
<tr>
<td>Demonstrate knowledge of appropriate specimen handling, storage and use for specific molecular, toxicology and histocompatibility testing.</td>
<td>DL, JC, FSO, DSP, IC</td>
<td>DO, GR/FE</td>
</tr>
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Demonstrate knowledge of the use of histocompatibility testing for transplant purposes.  

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# MOLECULAR PATHOLOGY CORE COMPETENCY: PROFESSIONALISM

**Goal:** 
*Demonstrate a commitment to carrying out professional responsibilities, adherence to ethical principles and sensitivity to a diverse patient population.*

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<tr>
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</thead>
<tbody>
<tr>
<td>Demonstrate knowledge of regulatory and Health Insurance Portability and Accountability Act (HIPAA) issues pertaining to the use of genetic testing in human research and clinical molecular practice.</td>
<td>FSO, DSP, RM</td>
<td>RR, DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate compassion: be understanding and respectful of patients, their families, and the staff and physicians caring for them.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Interact with others without discriminating on the basis of religious, ethnic, gender identity, or educational differences.</td>
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<td>DO, GR/FE</td>
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<td>Demonstrate positive work habits, including punctuality, dependability, and professional appearance.</td>
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<td>Demonstrate a responsiveness to the needs of patients and society that supersedes self-interest.</td>
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<td>Demonstrate principles of confidentiality with all information transmitted both during and outside of a patient encounter.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a commitment to excellence and ongoing professional development.</td>
<td>RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate interpersonal skills in functioning as a member of a multidisciplinary healthcare team.</td>
<td>FSO, DSP, RM, IC</td>
<td>DO, GR/FE</td>
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# MOLECULAR PATHOLOGY CORE COMPETENCY: SYSTEM-BASED PRACTICE

**Goal:** 
*Residents must demonstrate an awareness and responsiveness to the larger context and system of healthcare and the ability to call on system resources to provide pathology services that are of optimal value.*

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<th>Objectives:</th>
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<tr>
<td>Demonstrate understanding of the role of the laboratory in the healthcare system.</td>
<td>DL, FSO, IC</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Show a working knowledge of the basic principles of quality assurance, quality control, continuous quality improvement, and outcomes analysis, as they apply to molecular pathology.</td>
<td>FSO, RM</td>
<td>DO, GR/FE</td>
</tr>
<tr>
<td>Demonstrate a familiarity with standards set forth by the CAP and TJC for laboratory certification.</td>
<td>RM, OT</td>
<td>DO, GR/FE, SE</td>
</tr>
</tbody>
</table>
Understand the principles involved in public-health perinatal genetic testing paradigms.

RM, OT

DO, GR/FE, SE

SPECIFIC LEARNING OBJECTIVES

Acquisition of Knowledge of Specific Tests Using Molecular Biology Methods

- Understand basic molecular biology concepts.
- Know molecular testing methods for inherited causes for thrombophilia, such as factor V Leiden, prothrombin 20210 mutation, MTHFR, and platelet glycoprotein III polymorphisms (PIA 1/2).
- Understand molecular testing and interpretation for cystic fibrosis diagnosis and screening.
- Understand molecular testing for hematologic malignancies, including non-Hodgkin lymphomas (T-and B-cell gene rearrangements) and chronic myelogenous leukemia (bcr-abl detection and quantitation for therapeutic monitoring), and other translocation detection or quantitation assays.
- Understand molecular diagnostic tests for detection and speciation of pathogenic organisms, including Chlamydia trachomatis, N. gonorrhoeae, M. tuberculosis, high-risk human papillomaviruses, and viruses that cause encephalitis/meningitis (HSV and enteroviruses).
- Understand qualitative and quantitative methods used to determine viral load in human immunodeficiency virus, cytomegalovirus, Epstein–Barr virus, and hepatitis C virus, as well as human immunodeficiency virus and hepatitis C virus genotyping to direct therapy.
- Be familiar with molecular testing for trinucleotide repeats diseases, such as fragile X.
- Understand pharmacogenomic testing for cytochrome p450 mutations and other mutations that affect sensitivity to chemotherapeutic agents, such as thiopurine methyltransferase (TPMT), or other drugs.
- Be familiar with molecular testing for hereditary hemochromatosis, including the C282Y and H63D polymorphisms.
- Understand the principles behind human identity testing for transplant (see also the Immunology and Immunogenetics section).

[Pathology Milestones]

PC = Patient Care
MK = Medical Knowledge
PBL = Problem Based Learning
ICS = Interpersonal and Communication Skills
PRO = Professionalism
SBP = Systems Based Practice

For expanded definitions, see pages 30-31.

Specific Analytical and Technical Training Learning Objectives for Molecular Pathology

- Have awareness of sample types, preparation, and storage for molecular biology tests.
- Understand applicability of testing to samples of blood, bone marrow, body fluids (e.g., CSF, pleural, and peritoneal samples), lymph node, and spleen.
- Understand storage media and conditions for cells, DNA, and RNA.

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KUMC Pathology Residency Manual
• Understand DNA extraction and purification from a variety of biological specimens.
• Have knowledge of restriction endonuclease digestion of purified DNA or amplified DNA.
• Understand electrophoretic separation of DNA fragments, native DNA gel electrophoresis for verification of DNA quality, photographic documentation of gels, and capillary electrophoresis methods. [PC, MK]
• Have knowledge of total cellular RNA extraction, quantitation, separation of mRNA, and reverse transcription to generate cDNA.
• Understand southern blot DNA hybridization.
• Understand DNA sequencing.
• Have experience and knowledge of in vitro DNA amplification using the PCR and alternative amplification systems, as well as awareness of methods to prevent contamination.
• Understand varying means of analyzing PCR products, e.g., electrophoresis, sequencing, and restriction enzyme digestion.
• Understand mutation detection and scanning technologies for single-and multiple-mutation platforms.
• Understand real-time quantitative PCR and reverse-transcription-PCR.
• Understand DNA and gene expression microarrays.
• Be aware of the legal, ethical, and social implications of genetic testing.

Consultation and Presentation of Cases Using Molecular Techniques/Data

• Understand and use pedigrees for familial genetic assessments.
• Interpret and report molecular results in association with pathologic and laboratory findings and clinical history to reach a final diagnosis.
• Assess the sensitivity and specificity of testing for an individual patient's disease state.

RECOMMENDED READING LIST:

Specific Toxicology Learning Objectives:

• Be able to answer the following questions: What is the meaning of comprehensive drug screening? How comprehensive drug screen differ between laboratories? What drugs are identified on a comprehensive drug screen? What is the best sample for drug screening? Why? What are the most common drugs seen in overdose situations (pediatrics vs adults)?

• Understand basic principles of instrumentation/methods used in toxicology laboratory including:
  o Spot tests.
  o HPLC.
  o GC-FID/MS.
  o Atomic Absorption spectrometry.

• What is the specimen of choice for different heavy metal testing?

• Learn about lead testing and interpretation of lead levels. Interaction of lead with other metals in the body, particularly iron. Role of Zinc-Protoporphyrin (ZPP) in lead toxicity diagnosis. Advantages and disadvantages of ZPP testing?

• What are commonly abused drugs in adolescents?

• What are the alternate samples for drugs of abuse testing? Saliva, meconium, hair etc. Advantages and disadvantages of alternate samples.

• What are common drugs of abuse?

• What are the forensic/legal issues in laboratory testing for drugs of abuse?

• Understand specific overdoses and their antidotes.
  o Alcohols (ethanol, methanol, isopropanol)
    i. Ethylene glycol.
    ii. Estimation of ethanol alcohol conc.
    iii. Various methods (lab Vs breath analyzer).
    iv. Plasma Vs Whole Blood Vs Urine.
    v. Management.
  o Heavy Metals (Ar, Cd, Fe, Lead, Mercury)
    i. Specimen of choice.
    ii. Mechanism of action.
    iii. Lab tests.
    iv. Antidotes (Dimercaprol, EDTA, Penicillamine, Deferoxamine, Succimer).

• Analgesics/ Antipyretic(Acetaminophen, salicylate, Ibuprofen).

• Antidepressants (TCAs, Newer).

• Anticonvulsants (Phenytoin, Phenobarb, Carbamazepine, Valproate etc.).

• Cardiovascular drugs (Digoxin, Beta-blockers, Disopyramide, Quinidine).

• Drugs of abuse
  o Cocaine.
  o Amphetamines.
  o Marijuana.
- PCP (Phencyclidine).
- Opiates.
- Propoxyphene.
- Methadone.
- LSD (Lysergic Acid Diethylamide).
- Others: Nitrite, Nitrates, CO, Pesticides, OTCs, Plants, Hydrocarbons.

Specific Learning Objectives for Histocompatibility Testing (Midwest Transplant Network)
- Demonstrate an understanding of ethical, socioeconomic and medical/legal issues that affect patient care in organ procurement and histocompatibility testing.
- Use information technology to support patient care decisions, ensure quality assurance, and educate medical professionals and the public concerning histocompatibility testing.
- Observe and understand HLA typing by molecular methods and HLA antigen testing by serology.
- Demonstrate an understanding of crossmatching by flow cytometry using T cells and B cells.
- Perform a crossmatch using the anti-human globulin (AHG) technique.
- Observe and understand VNTR/STR chimerism assays.
- Know and apply basic science knowledge to understand basic concepts of histocompatibility testing.

DUTIES AND RESPONSIBILITIES OF THE RESIDENT
- Be present in the lab for significant amounts of time to allow exposure to routine molecular laboratory and toxicology work.
- Meet the general objectives outlined above and complete any work on unknowns, reading assignments or project assignments during the rotation.
- Attend required conferences at KUMC and at CMH.

RESIDENT EVALUATION
- The resident must show active participation in the laboratory.
- The resident must show improvement in knowledge of diseases that are diagnosed by molecular and toxicology methods.
- The resident must show improvement in knowledge of molecular and toxicology laboratory skills.
DERMATOPATHOLOGY/NEUROPATHOLOGY

DERMATOPATHOLOGY FACULTY
Garth Fraga, MD  
Associate Professor, Subspecialty: Dermatopathology

NEUROPATHOLOGY FACULTY
Kathy Newell, MD  
Associate Professor and Director, Neuropathology

The dermatopathology/neuropathology rotation is a combined rotation. Residents will participate in both areas each day. The dermatopathology portion of the rotation is described first, followed by the neuropathology portion of the rotation.

**Dermatopathology**

Dermatopathology is a joint subspecialty of both pathology and dermatology. The skin is the most visible organ and even small abnormalities are easily recognized, unlike visceral abnormalities. Skin biopsies are relatively easy to perform and fairly risk-free.

As a result of these factors, skin biopsies often comprise a large proportion of all biopsy samples. Dermatopathologists apply histologic techniques to diagnose these biopsies. Dermatopathology can be a challenging field to master because there are more than 1,500 described skin diseases.

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GOALS AND OBJECTIVES

The goal of this rotation is for the resident physician to acquire a satisfactory level of diagnostic competence in dermatopathology.

Objectives of this rotation include:

- Learn how to recognize and manage common dermatopathology problems by daily attendance at the noon dermatopathology sign-out session. The resident physician should become comfortable diagnosing most melanocytic and epidermal neoplasms and common inflammatory skin diseases (FSO, DO). [PC, PBL, ICS, PRO, SBP]
- Acquire dermatopathology knowledge by reading *Dermatopathology* by Elston and Ferringer from cover to cover (P, PSE). [MK]
- Supplement dermatopathology fundamentals by listening to *Dermatopathology: A Comprehensive Review* video series provided by Dr. Fraga and attending the dermatopathology teaching conferences every Thursday at 1 p.m. and the second Friday of the month at 8 a.m. (DL, IC, USC; DO, PSE). [PC, MK, PBL]
- Apply resident skills to slide study boxes of epidermal and melanocytic tumors provided by Dr. Fraga (USC, PSE). [PBL]
- Learn techniques for trimming in skin samples and sectioning paraffin blocks by participating in the prosection of skin biopsies in the gross room and the handling of trimmed in skin specimens in the histology laboratory (DSP, DO). [PC, ICS, SBP]
- Successful performance on a unknown slide test administered by Dr. Fraga at the end of the rotation (USC, PSE).

DUTIES AND RESPONSIBILITIES OF THE RESIDENT

Since this is a combined rotation with neuropathology, there will be some flexibility in order to maximize the learning experience of the resident. For example, *the resident should attend all neuropathology frozen sections, even if they conflict with a previously scheduled dermatopathology activity.*

- The resident should attend the daily noon sign-out of cases in Dr. Fraga's office.
- The resident should help prepare and attend the weekly, 1 p.m. Thursday dermatopathology conference for the dermatology residents.
The resident should help prepare and attend the monthly, 8 a.m., dermatopathology conference for pathology residents.

The resident should attend the 1-6 p.m., Kansas City Dermatological Society meeting on the third Thursday of the month.

Provided there is no previously scheduled neuropathology activity, the resident should spend afternoons assisting with the trimming in of skin samples in the gross room. Scheduled neuropathology activities such as case review with Dr. Newell should take precedence over trimming in skin samples, which can be handled by the gross room technicians.

The resident should arrange to come in early to the histology laboratory at least one morning to assist with the embedding and microtomy of skin samples.

The resident should attend all KU pathology teaching conferences expected of a resident on an AP KU elective (AP and CP core lectures, AP/CP conference, board review, CPC, Friday specialty conference, Grand Rounds, Journal Club and Monday/Thursday, Surgical Pathology conferences).

RESIDENT EVALUATION

The resident physician's patient care skills, medical knowledge, practice-based learning and improvement ability, interpersonal and communication skills, professionalism and systems-based practice skills will be evaluated by Dr. Fraga using a standard pathology rotation evaluation form on a scale from needs improvement, meets expectations, to exceeds expectations.

RECOMMENDED READING LIST:


NEUROPATHOLOGY

Neuropathology focuses on the study of neoplastic and non-neoplastic conditions of the central and peripheral nervous systems. Categories include vascular disease, trauma, developmental disorders, demyelinating and dysmyelinating disorders, neurodegenerative diseases, infections, metabolic and toxic disorders, muscle and nerve disorders, and primary and secondary tumors. In the study of neuropathologic disease, it is necessary to have an appreciation of gross and microscopic anatomy of the nervous system; therefore, continual review of neuroanatomy is an essential component of neuropathology. In surgical neuropathology, correlation with neuroradiology is emphasized. This rotation will focus primarily on neurosurgical specimens, with a minor emphasis on autopsy material. Individualization of topics for study, based on resident needs and interests, can be accommodated.

GOALS AND OBJECTIVES

The major goal of this rotation designed for residents in pathology is to provide exposure to and education about the major categories and classification of neuropathologic disease. The rotation will stress general principles and special techniques in approaching the study of these diseases and introduce tools and resources to enable successful independent and collaborative evaluations of neuropathologic entities.
DUTIES AND RESPONSIBILITIES OF THE RESIDENT

- Participate in the evaluation of neurosurgical specimens, including intraoperative consultations, with the attending neuropathologist.
- Review neurosurgical cases independently and with the attending neuropathologist.
- Review assigned cases of special neuropathological interest from the autopsy, surgical, or consult services.
- Study suggested textbooks or other references to learn about current cases.
- Review learning materials in the neuropathology computer modules available on the rotation to enhance and supplement other reading materials.
- Learn about special stains, immunohistochemical stains, and ancillary tests, including genetic markers, indicated in the current diagnostic and prognostic evaluation of neuropathologic specimens.
- Perform computer-based literature searches to review new findings with respect to diagnosis, treatment, or etiology of neuropathological diseases.
- Develop the necessary skills and education to initiate and coordinate a competent postmortem examination of the central nervous system.
- Participate with the attending neuropathologist in selected weekly and monthly clinical and teaching conferences at KUMC that focus on gross and microscopic neuropathology.

CONFERENCES

Neuro-Oncology Tumor Board
Time/Location: Monday (every other), 7 a.m.; 4893 Eaton.
Primary audience: Neurosurgeons, pathologists, radiation oncologists, neuroradiologists, medical oncologists, neurologists, and trainees.
Purpose: Bi-weekly review of patients with central or peripheral nervous system tumors for preoperative and/or postoperative treatment planning.

Surgical Pathology Unknown Conference
Time/Location: Fourth Friday, 8:30 a.m.; Surgical Pathology Laboratory, KU Hospital.
Primary audience: Pathologists, pathology residents, students.
Purpose: Monthly review of classic and unique/rare cases from current neurosurgical specimens, including consultation cases, often presented by theme. Autopsy neuropathology material will be reviewed quarterly.

Neuropathology Conference
Time/Location: Second Friday, 12 p.m.; Surgical Pathology Laboratory, KU Hospital.
Primary audience: Residents in neurosurgery, neurology, and pathology.
Purpose: Monthly neuropathology teaching conference.

KU Alzheimer Disease Center Neuropathology Case Review
Time/Location: Second Friday (alternating months), 9 a.m.; Surgical Pathology Laboratory, KU Hospital.
Primary audience: Neurologists, neuroradiologists, pathologists, nurses, trainees, students, and ADC personnel.
Purpose: Bi-monthly neuropathology case-correlation teaching conference.
**Pituitary Tumor Board**

**Time/Location:** First Thursday, 7:30 a.m.; 4th floor MOB conference room.

**Primary audience:** Endocrinologists, ENT surgeons, neurosurgeons, pathologists, neuroradiologists.

**Purpose:** Review of patients with pituitary-sellar region tumors for preoperative and/or postoperative treatment planning.

**Epilepsy Surgery Conference**

**Time/Location:** Tuesday, by arrangement, 1 p.m.; 5089 Delp.

**Primary audience:** Neurologists, neurosurgeons, pathologists, neuroradiologists, neuropsychologists.

**Purpose:** Review of patients with chronic seizure disorders being considered for surgical intervention, including pre-surgical epilepsy monitoring and correlation with neuropathology.

**SCHOLARLY ACTIVITIES**

Research within the neuropathology laboratory is an option that residents may choose for elective activities. Projects are available in the areas of tumors, demyelinating disorders, and neurodegenerative disease.

**RESIDENT EVALUATION**

Residents will be evaluated on their overall participation in the rotation and demonstrated ability to provide useful consultation to the clinical service teams, medical knowledge, application of this knowledge to efficient/quality patient care, and gross and microscopic diagnostic, technical and observational skills. Residents are also evaluated on their interpersonal skills, professional attitudes, and reliability, with members of the teaching faculty, peers, laboratory staff, and clinicians. They are further evaluated on their initiative in fostering quality patient care and use of the medical literature, attendance at relevant selected conferences, preview of cases and formulation of differential diagnoses prior to formal discussions, clinicopathological correlations, completion of assigned readings relevant to daily cases, and research of new information on neuropathological processes by searching the literature.

**RECOMMENDED READING LIST:**

**General Neuropathology**

RECOMMENDED READING LIST:

Surgical Neuropathology\Tumors


Neurodegenerative Disorders


Developmental Neuropathology

ANATOMIC PATHOLOGY ELECTIVE

The elective in anatomic pathology is designed to allow a resident to obtain in-depth training in any specific area or subspecialty within anatomic pathology in which a faculty member is willing to provide additional training. Residents must obtain permission from the appropriate faculty member prior to scheduling the elective.

Legend for Learning Activities for Residents

| Didactic lecture | DL |
| Faculty sign-out | FSO |
| Journal club | JC |
| Directly supervised procedure | DSP |
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| Lab inspections | LI |
| Interdisciplinary conference | IC |
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Legend for Evaluation Methods for Residents

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ANATOMIC PATHOLOGY CORE COMPETENCY: PATIENT CARE

Goal:
Residents must demonstrate an increased level above the general residency competency within the chosen specific area or subspecialty within anatomic pathology.

| Objective: Develop in-depth diagnostic competency within the specific area of choice. | Learning Activities | Evaluation Activities |
| | FSO, DSP, RM | RR, DO, GR/FE |

KUMC Pathology Residency Manual
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**Goal:**
*Demonstrate knowledge about established and evolving biomedical, clinical and cognitive sciences and the application of this knowledge to specific area of pathology.*

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### DUTIES AND RESPONSIBILITIES OF THE RESIDENT

- These will be related to the specific project and will be determined by the elective mentor.

### RESIDENT EVALUATION

- The resident will be evaluated by the elective faculty mentor using the global rating faculty evaluation form.
CLINICAL PATHOLOGY ELECTIVE

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- These will be related to the specific area and will be determined by the elective mentor.

**RESIDENT EVALUATION**

- The resident will be evaluated by the elective faculty mentor using the global rating faculty evaluation form.
RESEARCH ELECTIVE

Research to advance knowledge in pathology is one of our department's missions. All residents are required to produce at least a poster or platform research presentation during their residency. They are encouraged to publish at least one manuscript in a peer-reviewed journal. A research elective allows for dedicated time to be applied towards a focused research project. **A research project can require considerable advance planning, and residents are strongly encouraged to begin the process of applying for the elective at least six months in advance of the planned rotation.**

[Pathology Milestones]

PC = Patient Care  
MK = Medical Knowledge  
PBL = Problem Based Learning  
ICS = Interpersonal and Communication Skills  
PRO = Professionalism  
SBP = Systems Based Practice  
For expanded definitions, see pages 30-31.

GOALS AND OBJECTIVES

The goal of this rotation is for the resident physician to acquire appropriate research skills, techniques and expertise in designing and carrying out pathology-related research.

The objectives of this rotation include:

- Learn how to develop and test a scientific hypothesis. [MK, PBL]  
- Learn how to review and evaluate the scientific literature. [PBL]  
- Learn about research ethics, informed consent and the process for obtaining institutional approval for a research project. [PBL, SBP]  
- Acquire skills in laboratory and clinical research. [PBL]  
- Learn biostatistics and data analysis. [PBL]  
- Develop communication skills in presenting research results in both national meetings and in peer-reviewed journals. [ICS]

DUTIES AND RESPONSIBILITIES OF THE RESIDENT

- The research mentor will determine the specific daily duties and responsibilities of the resident physician to ensure the successful completion of the research project.  
- The resident physician is required to participate in educational conferences at KUMC ensuring at least 80% attendance over a six-month period.
RESIDENT EVALUATION

The research mentor will complete a standardized resident rotation evaluation form at the end of the rotation. He/she will provide a full comprehensive evaluation of the resident’s performance on the rotation and provide documentation of progress in terms of personal growth and research outcomes to the Resident Education Committee.

GUIDELINES

Research projects may be composed of a variety of styles including, but not limited to those listed below.

- A series of case studies with similar theme or disease process (i.e., hospital acquired pneumonia, or a predicative marker in breast cancer). Please note: an individual case report is not considered a research project.
- A quality assurance project that resulted in a change in practices in pathology (i.e., evaluation of testing patterns within certain medical specialty areas).
- Evaluation of new tests or markers when compared to established practice.
- A basic science project.

It is understood that many research projects will not be completed within one block and may be continued part time while the resident is on other rotations or may be continued with another research rotation.

PROCEDURE

The academic year prior to scheduling a research rotation, the resident should identify a faculty sponsor, and have the faculty member confirm that they will serve as the resident’s preceptor and agree to be responsible for the research activity during the scheduled block(s).

- **At least three months prior to the research elective**, the resident and faculty member are required to submit a petition for a research elective to the Resident Education Committee for approval.

- **The petition must include the following elements:**
  - Statement of the hypothesis to be tested.
  - Description of the method to be used.
  - Description of the data to be collected.
  - Description of the methods of data analysis.
  - How the results will be reported (e.g. peer-reviewed journal, national meeting poster/presentation).
  - Will the research require the use of human material and if so, has the project obtained approval from the Institutional Research Committee (IRB).
  - Will the research require funding, and if so has the funding been obtained.
• The petition should be signed and dated by:
  - The Resident.
  - The Preceptor.
  - The Residency Program Director or Associate Residency Program Director.

• The REC will review the proposal and give final approval for the research elective.

At the end of the rotation the resident should submit a progress report to the faculty preceptor which will be used as part of the evaluation for the month. The faculty sponsor must submit the progress report along with a written evaluation of the resident's performance to the REC. This written evaluation is in addition to the standardized evaluation in MedHub. The resident is strongly encouraged to present their findings at the subsequent AP/CP Conference.

Approved by REC, 1.22.2015

RECOMMENDED READING LIST: